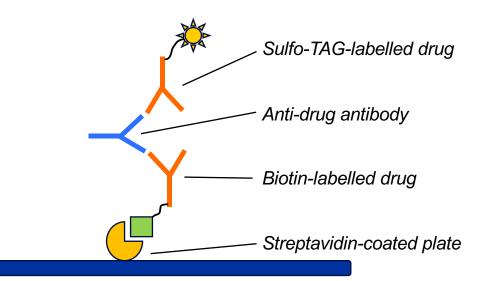


### Table of Contents

- // ADA Assay Development
- // Case study 1
- // Case study 2
- // Case study 3

### ADA ASSAY DEVELOPMENT

- ▶ Use pre-clinical ADA assay data: interpretation TK data
- Use of clinical ADA data: patient safety, interpretation PK data
- Standard (pre-) clinical ADA assay
  - Bridging assay
  - ECL detection
  - One tier approach (pre-clinical assay)
  - Three tier approach (clinical assay)
- ▶ Typical challenges
  - Sensitivity
  - Selectivity
  - Drug tolerance



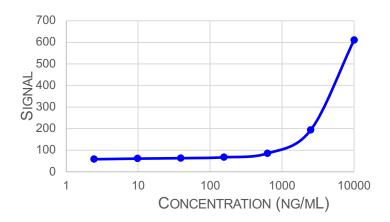




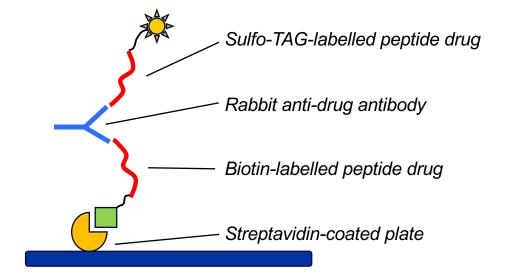
Assay development pre-clinical ADA assay for peptide drug



- Drug: peptide
- ▶ Positive control: rabbit anti-drug antibody
- Bridging assay format with ECL detection



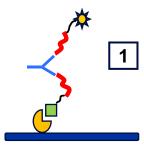
- → Signals low
- → Sensitivity potential issue

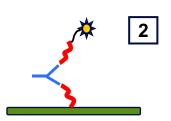


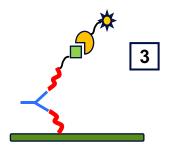


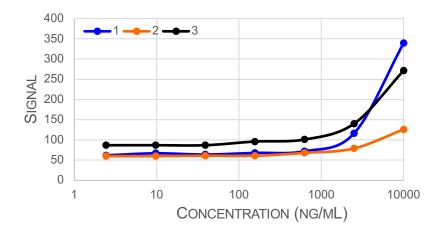
### CASE STUDY 1

ADA assay for peptide drug



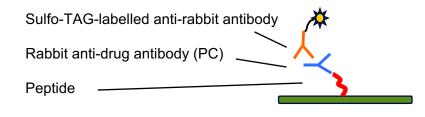


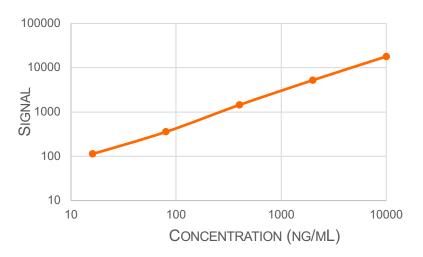




- $\rightarrow$  Signals low
- → Poor sensitivity
- → Steric hindrance of labelled peptide





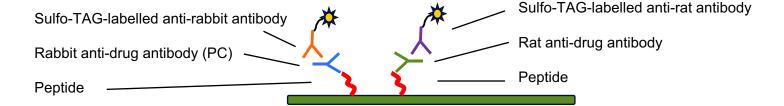


- MRD tested → 50 (2% matrix)
- Sensitivity assessment → <250 ng/mL</p>
- Response single individuals above response NC
  - Other blocking buffer
  - Higher coating concentration
- Acid dissociation step to improve drug tolerance



#### **DETECTION POSITIVE CONTROL**

#### **DETECTION RAT ANTI-DRUG ANTIBODIES**



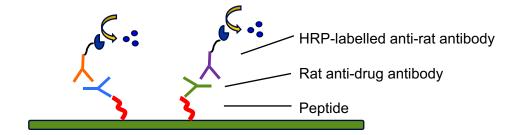
	Anti-rabbit antibody	Anti-rabbit antibody + Anti-rat antibody
NC	200	75,000
LPC	400	80,000
HPC	7,000	90,000

- Very high background levels caused by the Sulfo-TAG-labelled anti-rat antibodies
- Other blocking buffer reduced background only
   5- to 10-fold

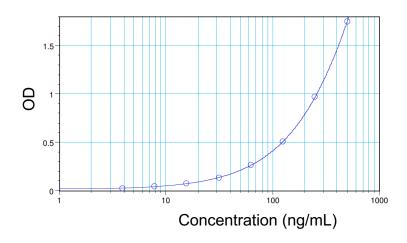




Switch from ECL to ELISA



- Tweaking capture and detection concentrations:
  - Good sensitivity
  - Good drug tolerance
- Rat IgG and IgM as controls for performance anti-rat antibodies





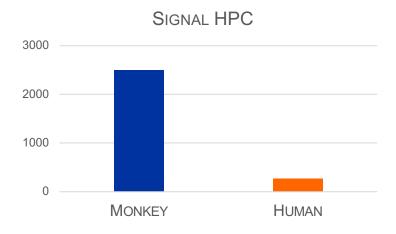
- ADA assay for peptide drug more challenging due to small peptide and bulky labels
- ▶ Check early in method development if positive control can bind to labelled peptide



FROM PRE-CLINICAL TO CLINICAL ASSAY: LOSS OF SIGNAL



- ADA assay monkey serum
- Bridging assay
- ▶ Positive control: purified rabbit polyclonal
- ▶ Drug: humanized mAb
- **ECL**

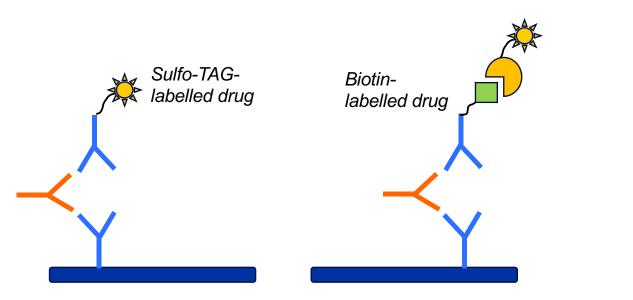


▶ Transfer of assay from monkey serum to human serum : loss of signal



### CASE STUDY 2 FROM PRE-CLINICAL TO CLINICAL ASSAY: LOSS OF SIGNAL

▶ Performance issues capture or detection reagents?

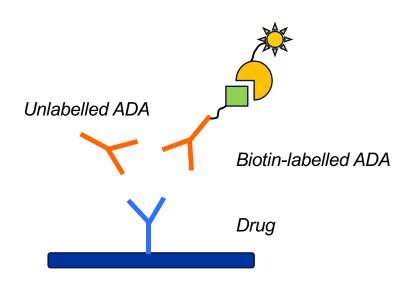


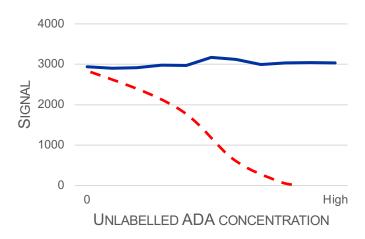
→ Low signals



# CASE STUDY 2 FROM PRE-CLINICAL TO CLINICAL ASSAY: LOSS OF SIGNAL

▶ Alternative assay set-up: competitive ADA assay



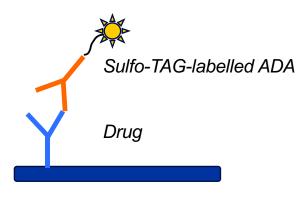


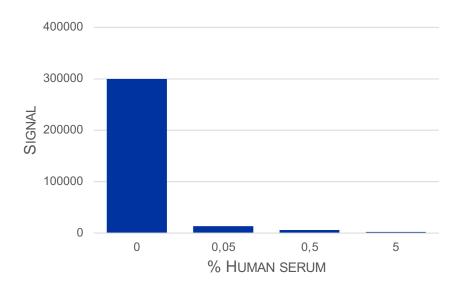
- → Signal in assay buffer 100-fold higher.
- → No competition. A-specific signal.
- → Matrix effect?





### ▶ Evaluation impact human serum



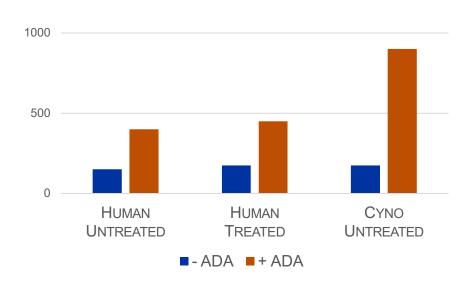


→ Interaction human serum components and ADA



### CASE STUDY 2 FROM PRE-CLINICAL TO CLINICAL ASSAY: LOSS OF SIGNAL

- Loss of signal caused by serum protein interference in human serum?
- ▶ Removal of serum proteins from human serum through thiophilic resin antibody purification



→ Antibodies interfering with ADA





#### Positive control fractions

- hIgG column eluate → used as positive control in monkey assay
- hlgG column flowthrough

#### Positive control in eluate fraction

- Binds to all human IgG antibodies
- Will only bind to antibody drug in monkey serum
- Will bind to drug and other IgG antibodies in human serum

### Positive control in flowthrough fraction

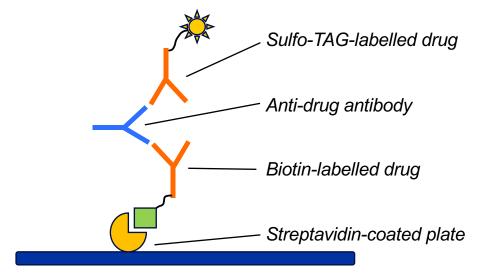
- Includes the idiotype-binding antibodies
- Used for development of ADA assay for human serum





## CASE STUDY 3 IMPAIRED ASSAY PERFORMANCE OVER TIME

- Validated ADA assay for human serum on ECL
- Over time assay performance diminished
  - → Poor range (NC ↑, HPC ↓)
  - → Lower sensitivity
  - → Plate-specific cut point too high



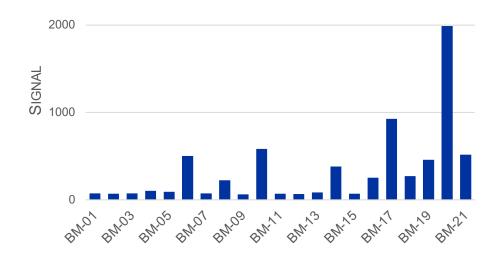




Impaired performance of ADA assay

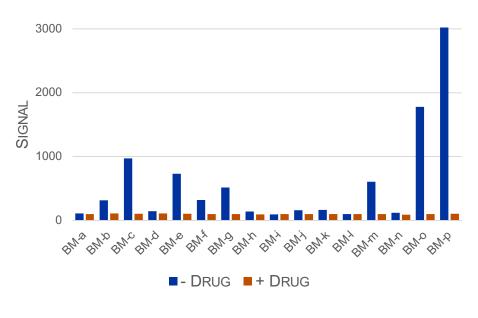
	Validation	Modified
NC	150	360
LPC	290	430
HPC	9000	4000

- ▶ New biotin- and Sulfo-TAG-labelled drug
- New negative control pool needed
- ▶ Testing individual blank human serum samples
  - → High variation between individual samples
  - → Acid dissociation did not improve results: pre-existing antibodies?



# CASE STUDY 3 IMPAIRED ASSAY PERFORMANCE OVER TIME

Confirmation high variation caused by pre-existing antibodies in blank human serum samples



→ 40-50% of samples have pre-existing antibodies



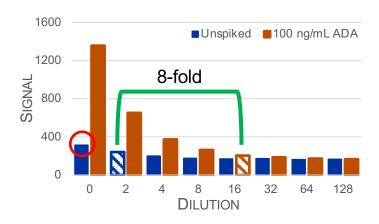


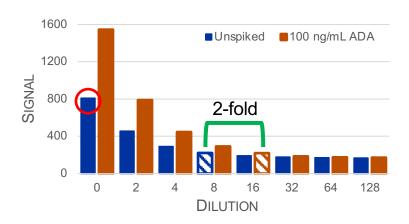
### Approach cut point determination

- Pre-screen large number of blank human serum samples
- Select samples with low response

### Identification of drug-induced ADA response

- Titer sensitivity will be determined using individual human serum samples with pre-existing antibodies





→ The drug-induced antibody response may not be detectable at very high pre-existing antibody concentrations



- ▶ Better sensitivity of new labelled drug resulted in increased background of negative control
- ▶ Higher negative control response due to more pronounced pre-existing antibody signals
  - → Pre-screen of blank human serum samples needed before cut point determination
  - → Titer sensitivity experiments needed for interpretation of study sample data
- Impaired HPC performance not related to pre-existing antibodies



