

# **Case Study: Alternative Approach of Validating ADA Assays for Bispecific Antibodies**

Lysie Champion

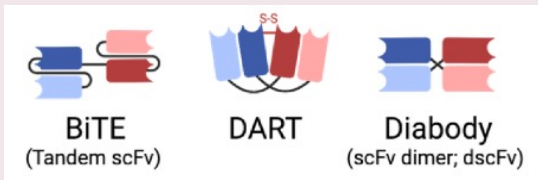
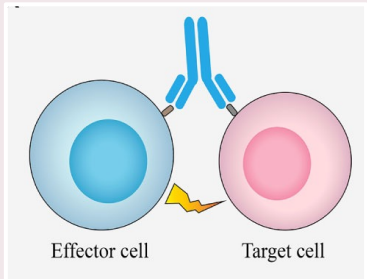
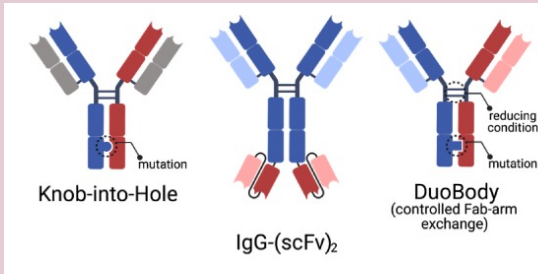
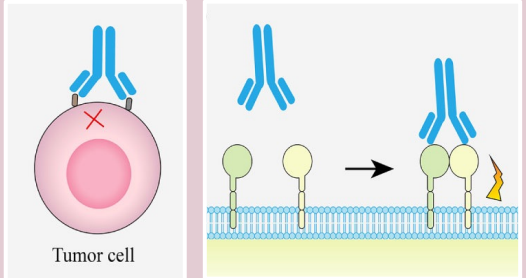
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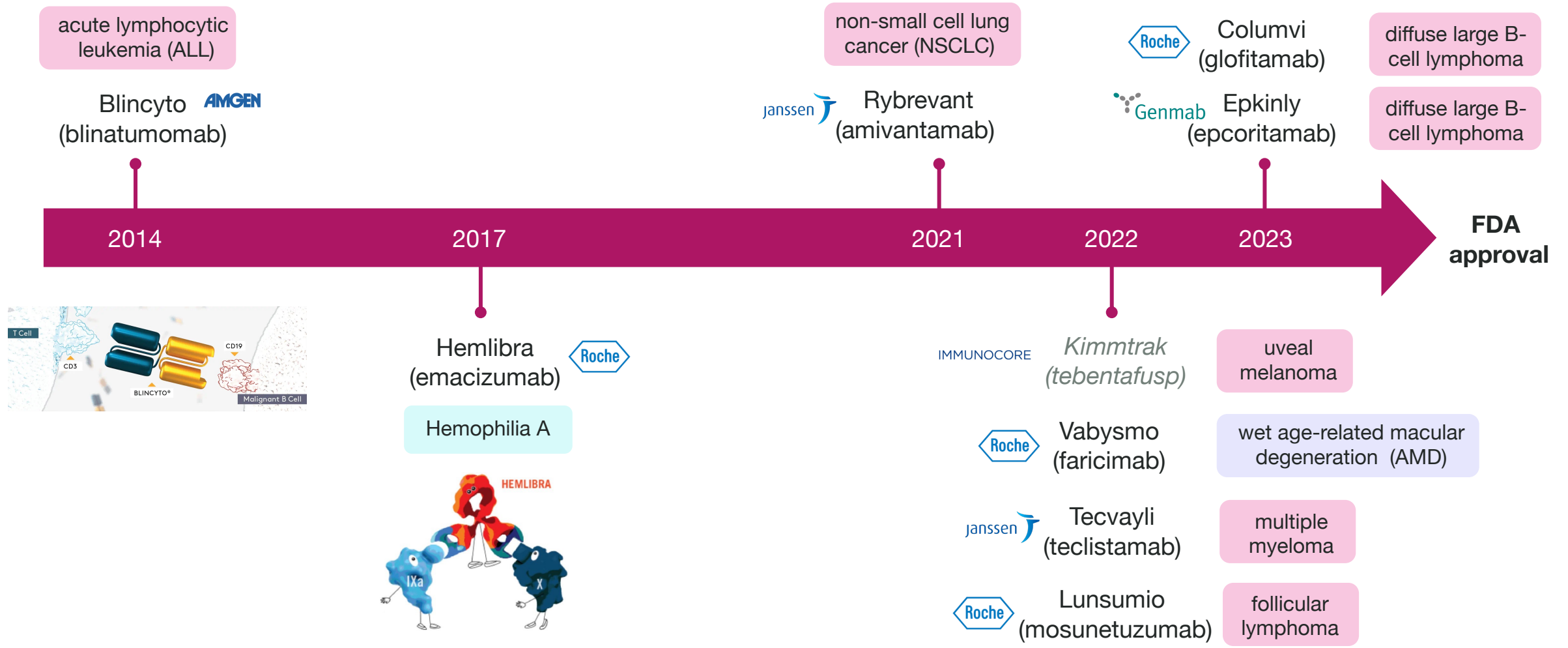
# What Is a Bispecific Antibody?

Bispecific antibodies (BsAbs) have been engineered to contain two distinct binding domains that can bind to two antigens or two epitopes of the same antigen simultaneously.

## Classifications

half-life <i>in vivo</i>		mechanism of action	
<ul style="list-style-type: none"> <li>• <b>Short-lived, small (&lt; 50 kDa) proteins</b></li> <li>• non-IgG-like, fragment (scFv)-based (lacking human Fc)</li> </ul> <p>→ half-life of few hours</p>	 <p>BiTE (Tandem scFv)      DART      Diabody (scFv dimer; dscFv)</p>	<ul style="list-style-type: none"> <li>• <b>Cell-bridging bsAbs: acting in-trans</b> (designed for cancer treatment by linking immune cells to malignant cells)</li> </ul>	 <p>Effector cell      Target cell</p>
<ul style="list-style-type: none"> <li>• <b>Long-lived, large bsAbs (&gt; 150 KDa)</b></li> <li>• IgG-like (antibodies Abs)</li> </ul> <p>→ half-life of up to several days</p>	 <p>Knob-into-Hole (mutation)      IgG-(scFv)<sub>2</sub>      DuoBody (controlled Fab-arm exchange) (reducing condition, mutation)</p>	<ul style="list-style-type: none"> <li>• <b>Antigen-crosslinking bsAbs, non cell-bridging: acting in-cis</b> (e.g. blocking signals of cell growth, activation of immune cells, co-factor mimetic)</li> </ul>	 <p>Tumor cell</p>

# Bispecific Antibodies as Biotherapeutics



# Bispecific Antibodies: Bioanalytical Challenges

- **ADA: domain specificity characterization of the immune response**  
Comparison of immunogenicity between moieties and related to novel / artificial structures (e.g. linker)

- Is domain specificity required? Regulatory recommendations:

- **FDA 2019 Guidance (Section IV.A.3):**

”For multi-domain therapeutic protein products, the sponsor may need to investigate whether the ADA binds to specific clinically relevant domains in the protein.”

- **EMA 2017 Guideline (Section 7.5)**

”The evaluation of this (*immune*) response, in particular, the characterization of the specificity of the induced antibodies is challenging and may require multiple assays for measuring immune responses to various moieties.”

→ risk assessment (safety – moiety with endogenous counterpart; clinical phase)

- If domain specificity characterization is required:

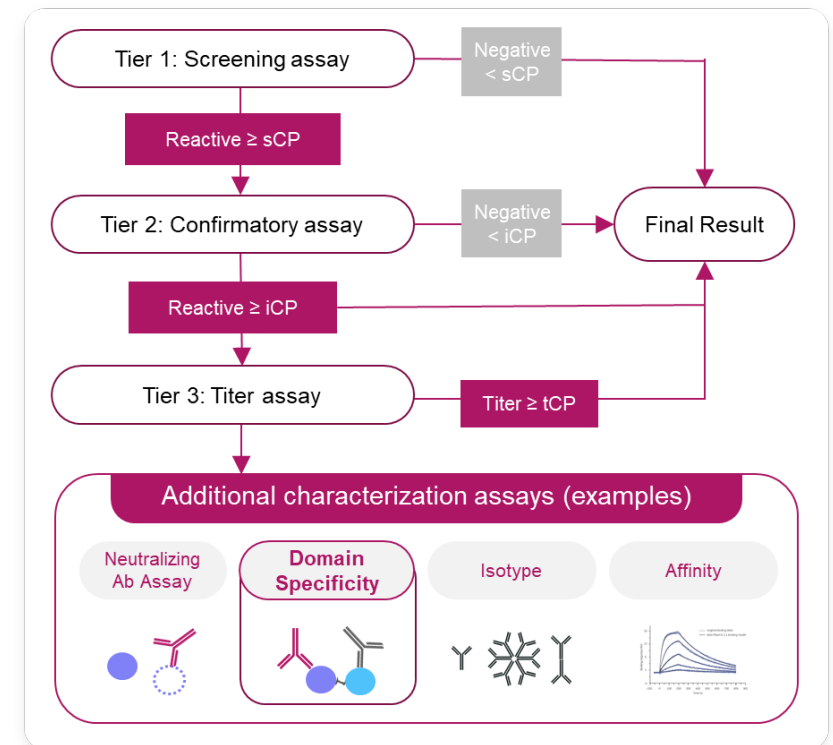
→ Development of multiple domain characterization assays to measure immune responses to different domains of the molecules

→ Additional reagent generation:

- domain-specific positive control
  - domain-specific inhibitors for the characterisation assay

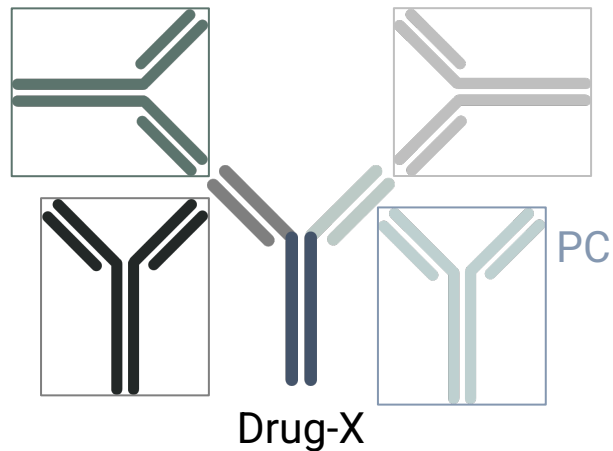
→ Assay development and validation is more time/ resource consuming

## ADA testing: the multi-tiered approach



# Positive Control Strategies

## Polyclonal Antibody as PC

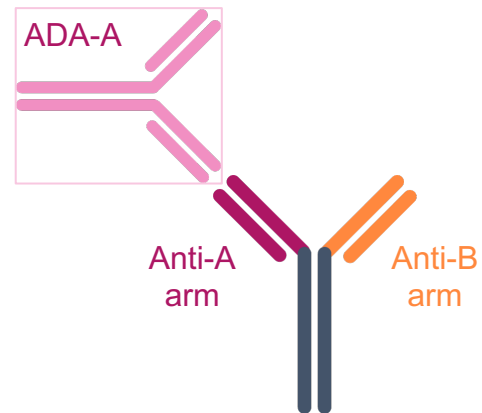


1 pAb used in all tiers

Life-cycle management of pAb PC is to be considered

## Domain-specific Monoclonal Antibody as PC

### Case Study 1



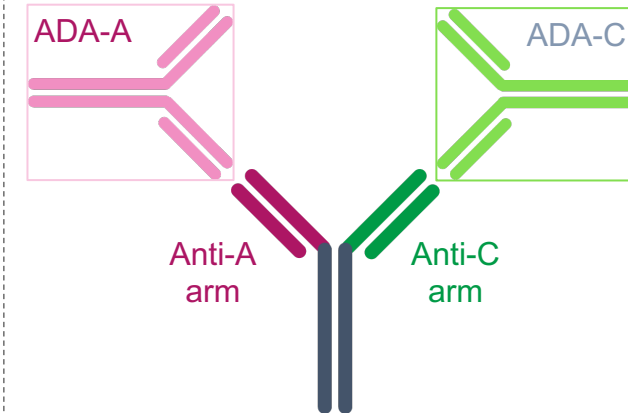
**Bispecific antibody 1  
(Drug-AB)**

1 mAb used as main PC in all tiers

2<sup>nd</sup> mAb needed in charact. assay

→ 2 independent mAb PCs

### Case Study 2



**Bispecific antibody 2  
(Drug-AC)**



1 mAb mix used in all tiers

→ 1 Pseudo-pAb PC

# Assay Requirements and Reagent Availability

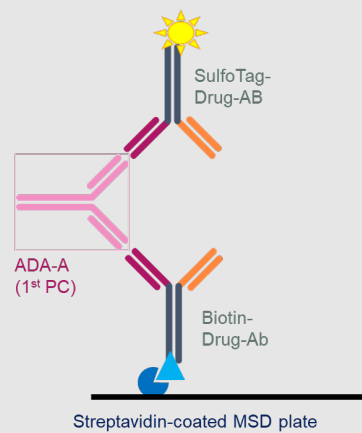
## Assay requirements (Drug-AB and Drug-AC)

- $\leq 100$  ng/mL sensitivity
- Low free drug interference
- Likely low target interference
- Typical MSD bridging assay
- No need for sample pre-treatment like acid dissociation

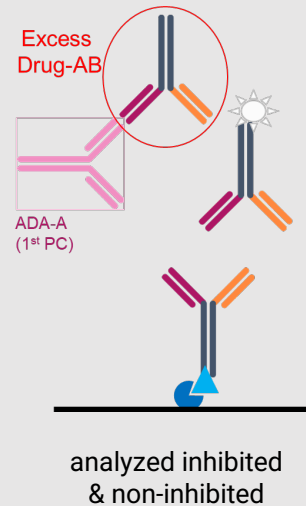
Reagents	Drug-AB (Case Study 1)	Drug-AC (Case Study 2)
Positive controls	<ul style="list-style-type: none"> <li>• mAb targeting anti-A arm (ADA-A)</li> <li>• mAb targeting anti-B arm (ADA-B)</li> </ul>	<ul style="list-style-type: none"> <li>• mAb targeting anti-A arm (ADA-A)</li> <li>• mAb targeting anti-C arm (ADA-C)</li> </ul>
Capture and Detection	• Biotin- and SulfoTag-Drug-AB	• Biotin- and SulfoTag-Drug-AC
Reagents for domain characterization	<ul style="list-style-type: none"> <li>• Anti-A arm</li> <li>• Anti-B arm</li> </ul> 	<ul style="list-style-type: none"> <li>• Anti-A arm</li> <li>• Anti-C arm</li> </ul> 

# Case study 1 – Two Independent mAb Positive Controls

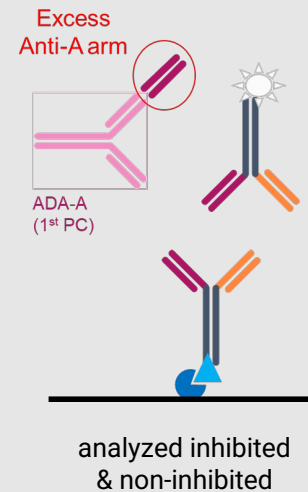
## Screening / Titer



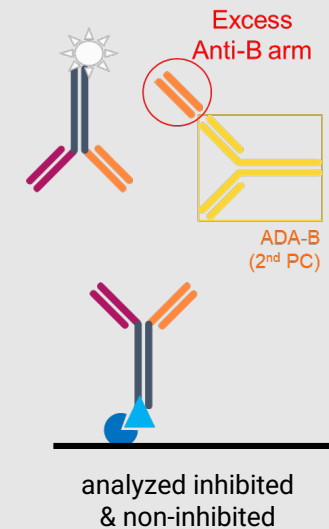
## Confirmatory



## Characterization A



## Characterization B

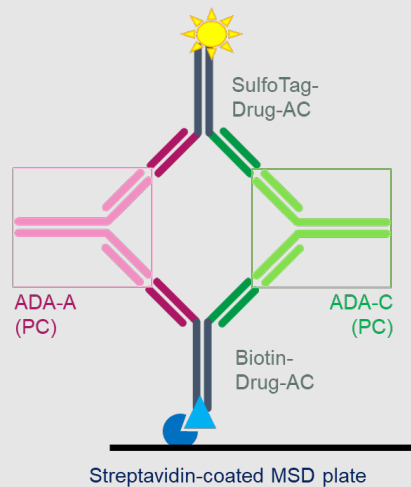


Reference item 1  
(ADA-A)  
→ main PC  
→ LPC, MPC, HPC

Reference item 2  
(ADA-B)  
→ 2<sup>nd</sup> PC  
→ LPC only

# Case study 2 – One Pseudo-pAb Positive Control

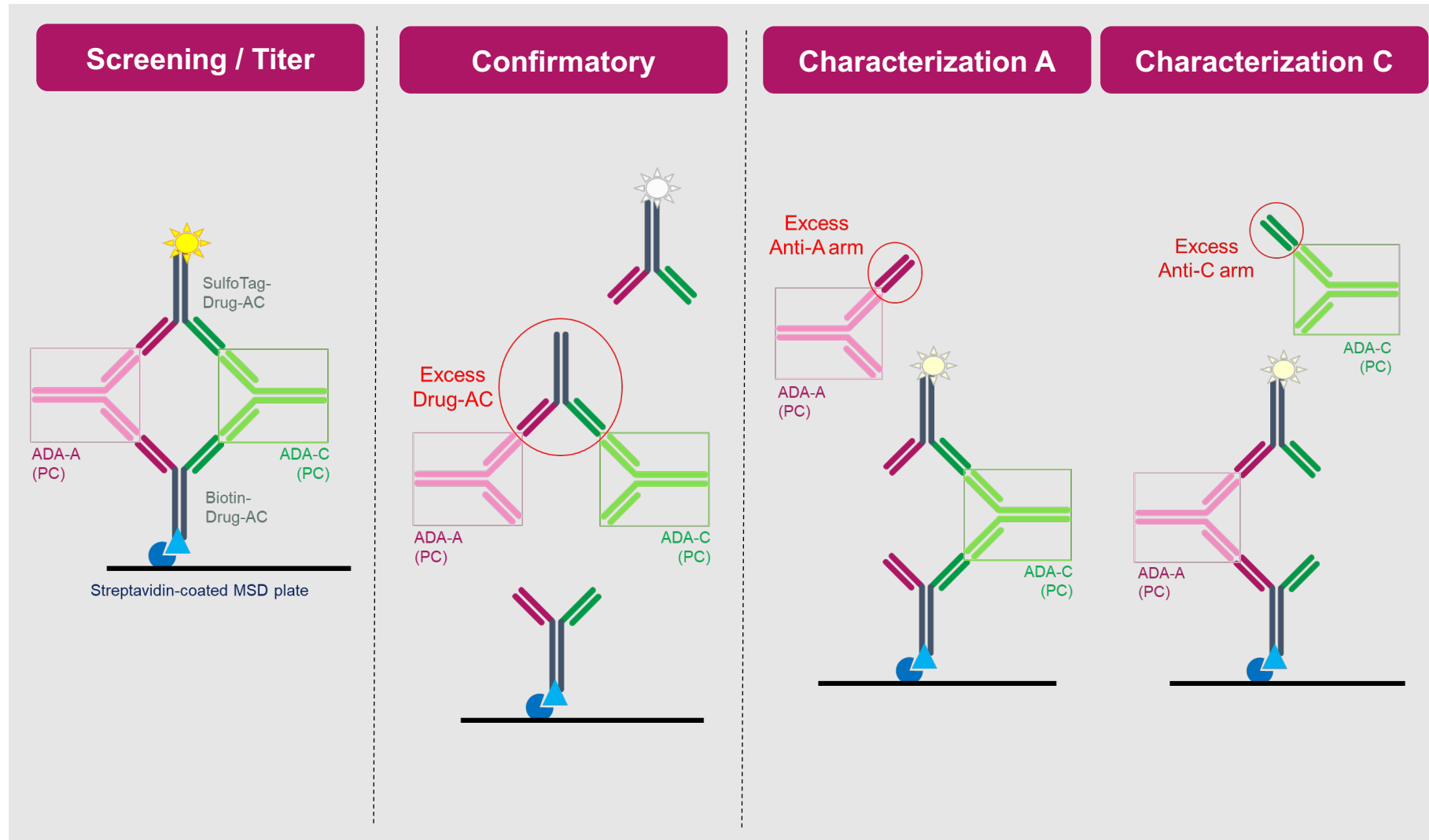
## Screening / Titer



- Reference item 1 (ADA-A) and Reference item 2 (ADA-C) were mixed 1:1 to create a pseudo-polyclonal antibody as positive control
- The Ab mix was used to prepare positive control (LPC, MPC and HPC) and validation samples  
→ only 1 positive control



# Case study 2 – One Pseudo-pAb Positive Control



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# ADA Assay Development for Bispecific Antibodies

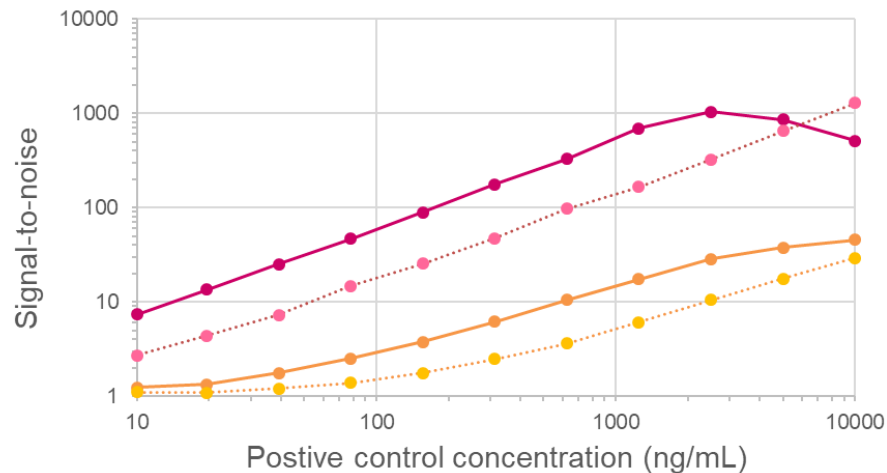
# Development Milestones for BsAb ADA Assay

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1. Titration of capture and detection reagents
2. Selection of positive control
3. Testing sample pre-treatment if required (drug-tolerance)
4. Selection of MRD
5. Establishment of confirmatory and domain characterization assays
6. Evaluation of key assay performance
  - Cut points
  - Sensitivity
  - Selectivity
  - Drug (target) interference
  - (Stability)

# Selection of Positive Control

## Drug-AB

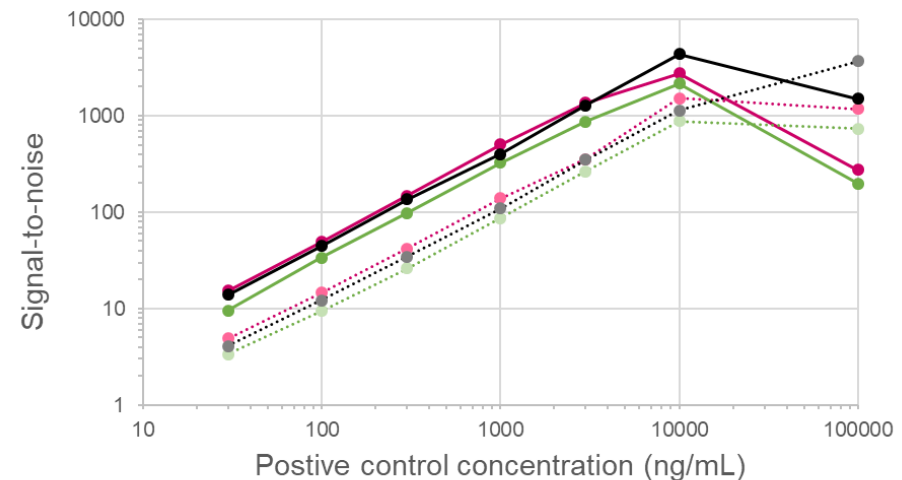


—●— ADA-A MRD 12.5      ·····●····· ADA-A MRD 50  
—●— ADA-B MRD 12.5      ·····●····· ADA-B MRD 50

Sensitivity: ADA-A >> ADA-B

→ ADA-A selected as main PC based on sensitivity (<< 100 ng/mL)

## Drug-AC



—●— ADA-A MRD 12.5      ·····●····· ADA-A MRD 50  
—●— ADA-C MRD 12.5      ·····●····· ADA-C MRD 50  
—●— ADA-A + ADA-C MRD 12.5      ·····●····· ADA-A + ADA-C MRD 50

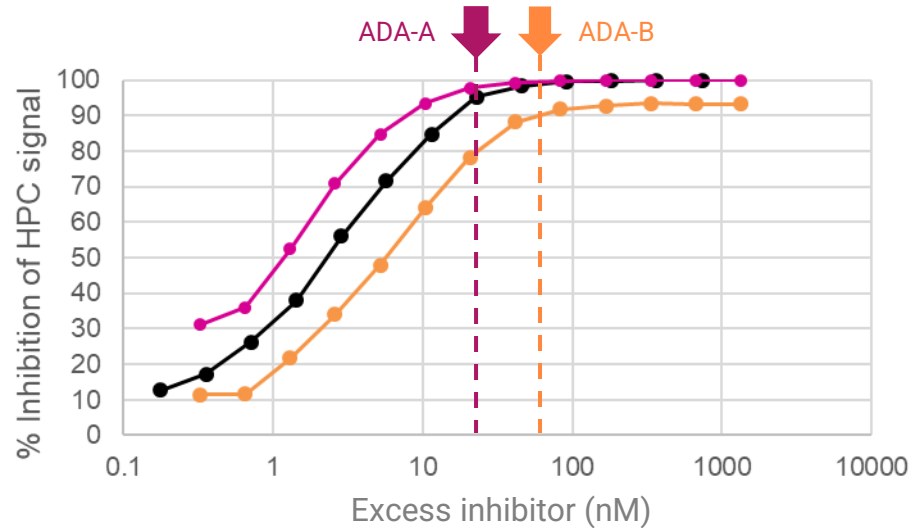
Sensitivity: ADA-A > ADA-C

→ mAb mix selected as PC based on sensitivity (<< 100 ng/mL) and dilution linearity ("hook effect" reduced)

Remark: if drug interference is expected, positive controls should also be evaluated in the presence of free drug

# Determination of Excess Inhibitor Levels in the Confirmatory and Characterization Assays

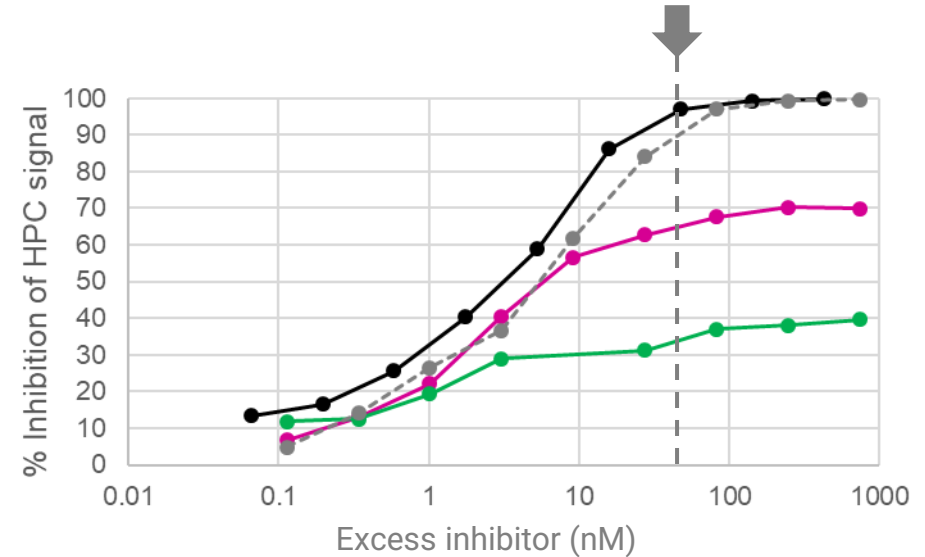
## Drug-AB



- Drug-AB inhibiting ADA-A (Confi.)
- Anti-A arm inhibiting ADA-A (Chara. A)
- Anti-B arm inhibiting ADA-B (Chara. B)

- Different PCs → different excess inhibitor conc.
- Full inhibition

## Drug-AC



- Drug-AC (Confi.)
- Anti-A arm (Chara. A)
- Anti-C arm (Chara. C)
- Anti-A arm + Anti-C arm

- Only one PC → similar excess inhibitor concentrations
- Partial inhibition (correlates with mAb binding affinities)

# Final Assay Parameters

	Drug-AB	Drug-AC
Assay format	MSD bridging assay (no acid dissociation)	
Positive controls	Two positive controls strategy <ul style="list-style-type: none"> <li>• ADA-A for all assay tiers but Char<sub>B</sub></li> <li>• ADA-B (LPC) for Char<sub>B</sub> (LPC)</li> </ul>	Antibody mix strategy (1 PC) <ul style="list-style-type: none"> <li>• ADA-A + ADA-C (1:1 mix) for all assay tiers</li> <li>• Pseudo-polyclonal Ab</li> </ul>
MRD (sample dilution)	25	50
Provisory LPC (pLPC)	pLPC <sub>A</sub> : 5 ng/mL pLPC <sub>B</sub> : 10 ng/mL	pLPC: 5 ng/mL

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# ADA Assay Validation for Bispecific Antibodies

# ADA Assay: Key Validation Parameters

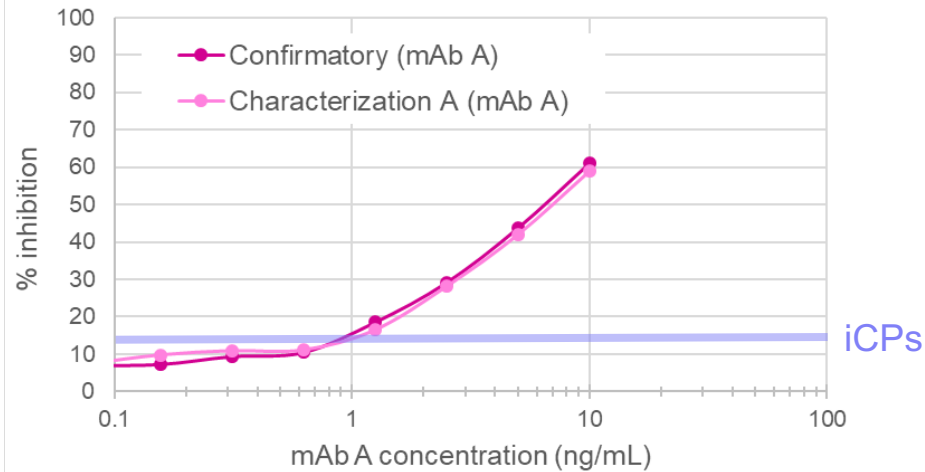
	Drug-AB		Drug-AC
	ADA-A	ADA-B	mAb mix
Positive control evaluated			
Screening cut point	x		x
Confirmatory cut point	x		x
Characterization cut points (2 assays)	x		x
Robustness	x		x
➔ Sensitivity and titer precision	x	x	x
Selectivity	x	x	x
Hemolytic / lipemic interference	x	x	x
Precision	x	x	x
Hook effect	x	x	x
Free drug interference	x	x	x
Stability	x	x	x

➤ With mAb mix – simplified validation as most parameters analyzed with only 1 PC

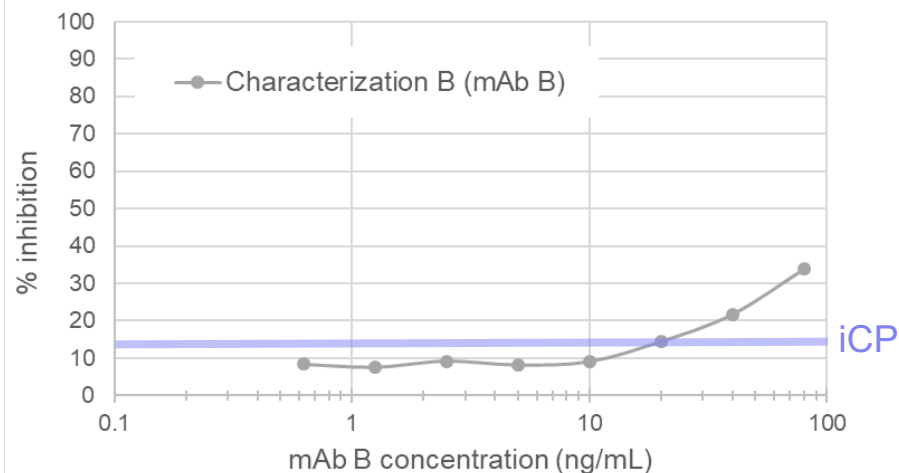


# Assay Sensitivity in Confirmatory and Characterization Assays

## Drug-AB



	Confi	Char A
PC	Titration of ADA-A	
Inhibitor	Drug-AB	Anti-A arm
Sensitivity	~ 1 ng/mL	



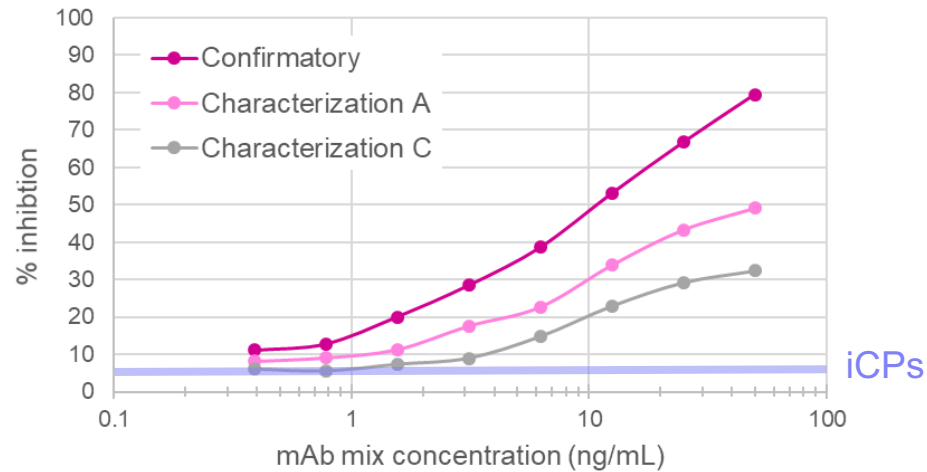
	Char B
PC	Titration of ADA-B
Inhibitor	Anti-B arm
Sensitivity	> 20 ng/mL

Two LPCs:

- LPC (ADA-A): screening / confirmatory / Char A
- LPC (ADA-B): Char B

# Assay Sensitivity in Confirmatory and Characterization Assays

## Drug-AC



	Confi	Char A	Char C
PC	Titration of Ab mix		
Inhibitor	Drug-AC	Anti-A arm	Anti-C arm
Sensitivity	2-8 ng/mL		

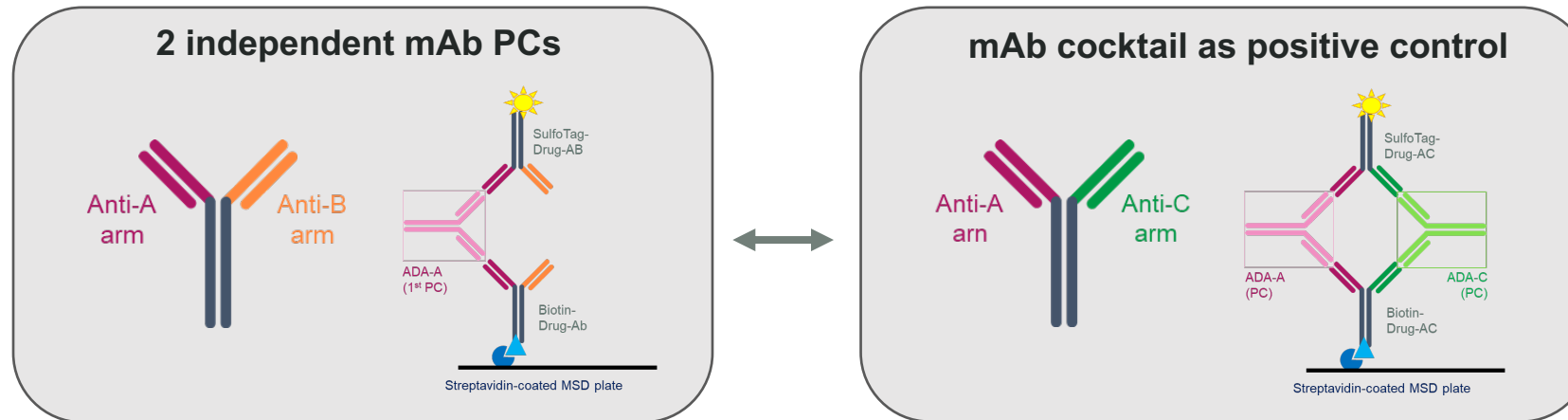
Only 1 LPC

- Despite the partial % inhibition in the Characterization assays the positive control could be diluted below the inhibition cut points.
- If sensitivities in confirmatory vs. characterization assays are too different, an additional LPC level ( $LPC_{Char}$ ) may be introduced in the characterization assays

**03**

**Conclusion**

# Pros and Cons of Using a Reference Antibody Mix to Detect ADA Against Bispecific Biotherapeutics



## Pros

- Antibody not selected solely based on performance (Ab with highest affinity)
- A cocktail of reference Ab will target both arms of the bispecific Ab thereby mimicking a polyclonal immune response
- Life-cycle management of mAb (vs pAb): easier to control batch consistency, sufficient quantity for long-term supply
- Most validation parameters (e.g. sensitivity, drug tolerance, selectivity) tested only with one positive control → reduce validation experiments, minimize spiking
- Fulfills the requirements of positive control in all tiers of ADA assays.

## Cons

- If the affinities of the antibodies are too different combined with a high inhibition cut point – false negative results may occur

# Conclusion

- Traditional pAb can be successfully replaced by a mAb cocktail (pseudo-pAb)
- mAb cocktail (pseudo-pAb) was also used as PC for NAb assay (case study 2 – Drug-AC)

Can be applied more broadly to other multi-domain drugs (e.g. ADC, fusion proteins), provided:

- - The affinities of each domain-specific mAb are not too different (peggylated drug)
  - Not too many domains → partial inhibition with narrow dynamic range (risk of false-negative)

- Positive control strategy should be considered early on as this can influence the generation of reference antibodies



Thanks to Florian Bernet, Marita Zoma, Harley Williams, Petra Struwe

**THANK YOU**