



High-throughput LC-MS/MS method for therapeutic oligonucleotides

Method for supporting pre-clinical studies

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11/29/23



OLIGONUCLEOTIDE DRUGS APPROVED

EXONDYS 51
(eteplirsen) Injection

Duchenne muscular dystrophy



CMV infection



Familial hypercholesterolemia



TTR Polyneuropathy

VYONDYS 53
(golodirsen) Injection

Duchenne muscular dystrophy



Familial chylomicronemia



Spinal muscular atrophy



Batten disease

Viltepso
(viltolarsen) injection

Duchenne muscular dystrophy



TTR polyneuropathy



Acute hepatic porphyria



Primary hyperoxaluria type 1



HEPLISAV-B®
[Hepatitis B Vaccine
(Recombinant) Adjuvanted]

CpG 1018 Hepatitis B vaccine

ANALYTICAL METHODOLOGY AT CRL

Consistent and global experience across multiple modalities

Mass Spectrometry

Oligonucleotides,
Modified RNA

Ligand Binding Assays

Oligonucleotides
(siRNA, ASO,
miRNA, etc.)

Immunogenicity Assays

Anti-drug
Antibody Assays
(Oligo-binding;
Clearance;
Neutralizing;
Endogenous nucleic
acids)

Molecular Biology

qPCR,
RT-qPCR
Branch DNA
RNA isolation
DNA isolation

HIGH-THROUGHPUT QUANTIFICATION OF OLIGONUCLEOTIDES FOR PRECLINICAL STUDIES

Transporter substrate assays

- Working Range requirement: 0.1-1,000 nM
- Matrices: HBSS, KH pH 7.4, cell extracts
- Sample number per study: <1000

Need for speed

- Low cost
- Rapid turnaround time
- Method development: Fit-For-Purpose
Method Validation: in 1-2 days
- Sample analysis and report: in 1-2 days

Fast, cheap and good



Can or cannot I have the best of them all?

OLIGO THERAPEUTICS - TWO PRIMARY ASSAY TYPES

Both Play a Role in Oligo Development Programs

Hybridization ELISA	LC-MS
Expensive custom probes required (capture, detector, cutting, etc. at \$2000 - \$10,000 each)	Internal standard required (off-the-shelf oligo may be used, \$500)
Moderate selectivity Assay reagents typically cross-react with catabolites / shortmers	Excellent selectivity Identify exact species measured and can monitor catabolites / shortmers
Throughput slightly less than average ELISA because of increased # of assay steps	Throughput average to slightly < average if multiple extractions required
No instrument issues	Harsh instrument conditions results in extra cleaning and downtime
Superior sensitivity 0.05 to 2 ng/mL LLOQ in plasma	Less sensitive 1 to 100 ng/mL LLOQ in plasma
Smaller dynamic range (2+ order of magnitude)	Larger dynamic range (3+ order of magnitude)
Minimal variation in assay procedure once format selected for a given drug platform, but skill of analysts very important	Minimal variation in sample extractions and LC conditions from analyte-to-analyte results in quick method development

Fast, cheap and good

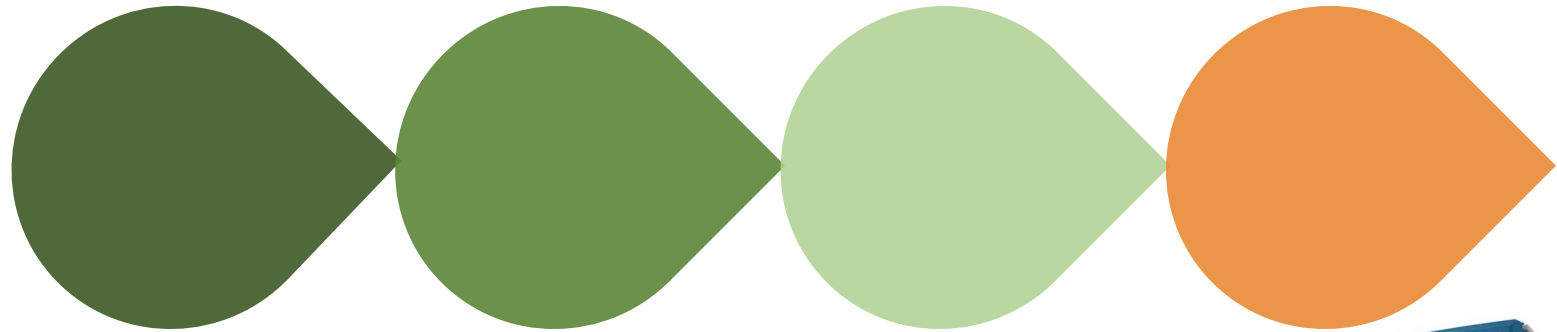


Can or cannot I have the best of them all?

TYPICAL SENSITIVITY OF HYBRIDIZATION ASSAYS

Biological Matrix	Format			
	Competitive Hybridization	Dual Hybridization	Hybridization Ligation	Nuclease-Dependent Cutting ELISA
Plasma (μL sample)	10 to 100 ng/mL (10 μL)	10 to 100 pg/mL (10-100 μL)	0.2 to 2 ng/mL (100-200 μL)	0.5 to 3 ng/mL (25-100 μL)
Tissues with Extraction	No Data	1 to 10 ng/g	5 to 30 ng/g	15 ng/g
Tissues without Extraction	No Data	10 ng/g	1 to 10 ng/g	15 ng/g

LC-MS of oligos



ION-PAIRING

- TEA-HFIP
- Apolar phases

ESI-MS

- SIM or MRM

RP

- NH_4OAc / $\text{NH}_4\text{O}_2\text{CH}$
- Apolar phases

HILIC

- NH_4OAc / $\text{NH}_4\text{O}_2\text{CH}$
- HILIC phases

HOW TO BOOST ANALYTICAL CAPACITY?

BY HAVING MORE LC-MS

KPI		Capacity increase
Throughput	(+)	2x
Uptime	(+)	2x
Maintenance	(-)	2x
Cost	(-)	2x
Mass Spectrometer Utilization	(-)	1x



BY HAVING HIGH THROUGHPUT LC-MS

KPI		Capacity increase
Throughput	(+)	2-16x
Mass Spectrometer Utilization	(+)	2-16x
Cost	(+)	2x
Maintenance	(+)	1x
Uptime	(-)	1x

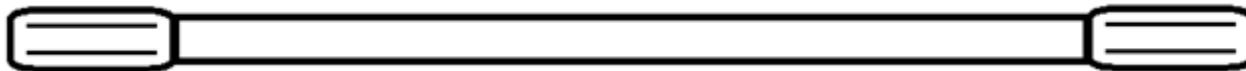


High Throughput Approach

Make a choice

- Resolution: Only for critical component(s)
- Plate Number: Less important for gradient separation.
- Peak capacity: Less important for MRM detection, matrix effect?
- Retention: Desalting, control of matrix effect
- Selectivity: Only for critical analytes

100 x 2.1 mm, 3um, 1 ml/min, t_g : 2.2 min

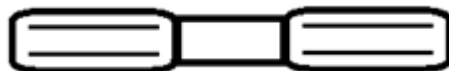


t_0 : 0.22 min

Cycle time: 3.96 min

Peak capacity ~ 126

10 x 2.1 mm, 3um, 1 ml/min, t_g : 0.2 min



t_0 : 0.02 min

Cycle time: 0.36 min

Peak capacity ~ 40

Cycle time: $t_g + 3 \times t_0 + 5 \times t_0$

Theoretical plate height small if:

1. Small particle size
2. Low flow rate
3. Low eluent viscosity
4. High temperature
5. Small molecule

$$N = \left(a \sqrt{\frac{1000 \eta \epsilon_T}{t_0 \Delta P}} + b \frac{1000 \eta \epsilon_T}{\delta^2 \Delta P} + c \frac{\delta^2}{t_0} \right)^{-1}$$

$$n_{C,G} \approx 1 + \frac{1}{4} \cdot \sqrt{N} \cdot \frac{10}{1 + 10 \frac{t_0}{t_G}}$$

High Throughput Approach

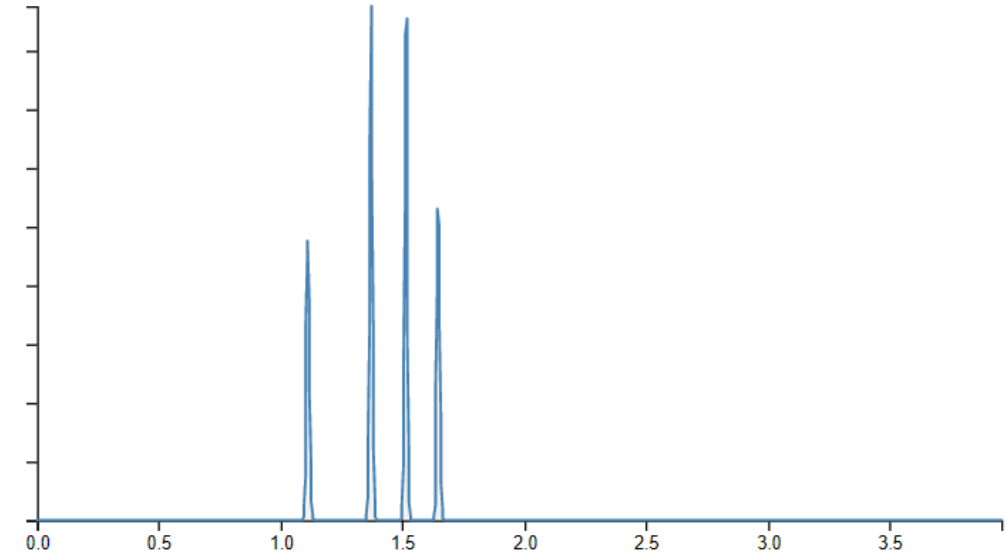
Make a choice

100 x 2.1 mm, 3 μ m, 1 ml/min, tg: 2.2 min

t₀: 0.22 min

Cycle time: 3.96 min

Peak capacity ~ 126

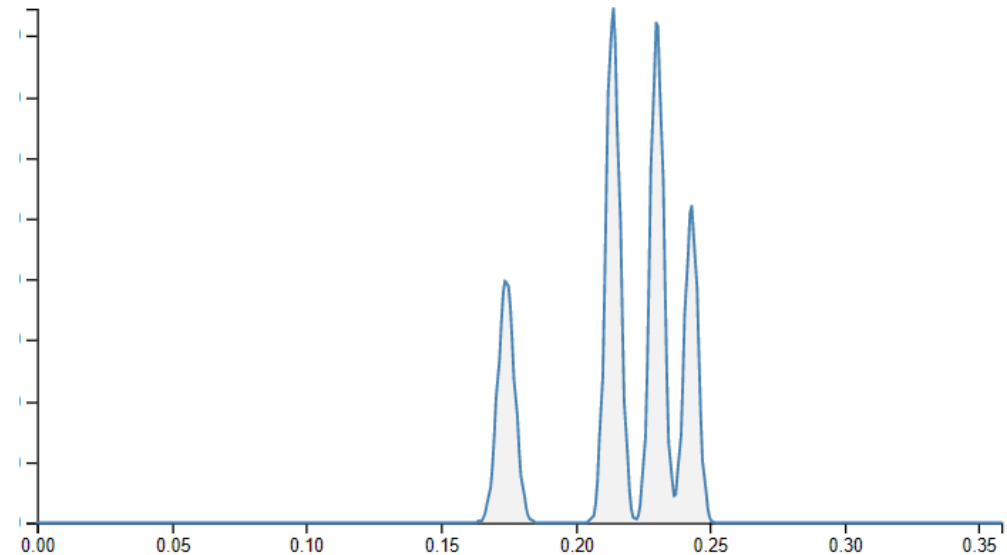


10 x 2.1 mm, 3 μ m, 1 ml/min, tg: 0.2 min

t₀: 0.02 min

Cycle time: 0.36 min

Peak capacity ~ 20 (40)

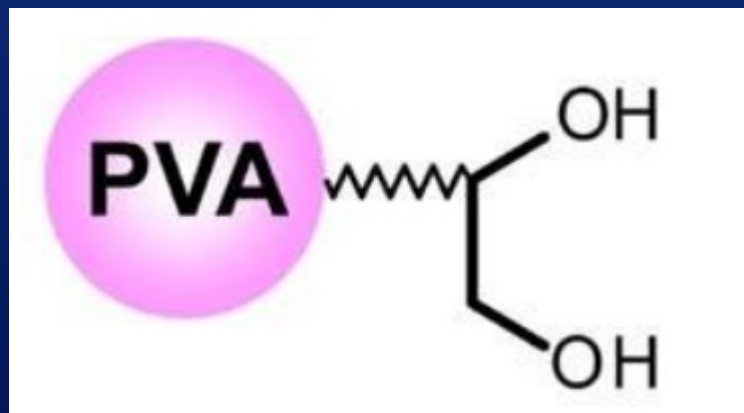


HT-LC- MS/MS OF OLIGOS

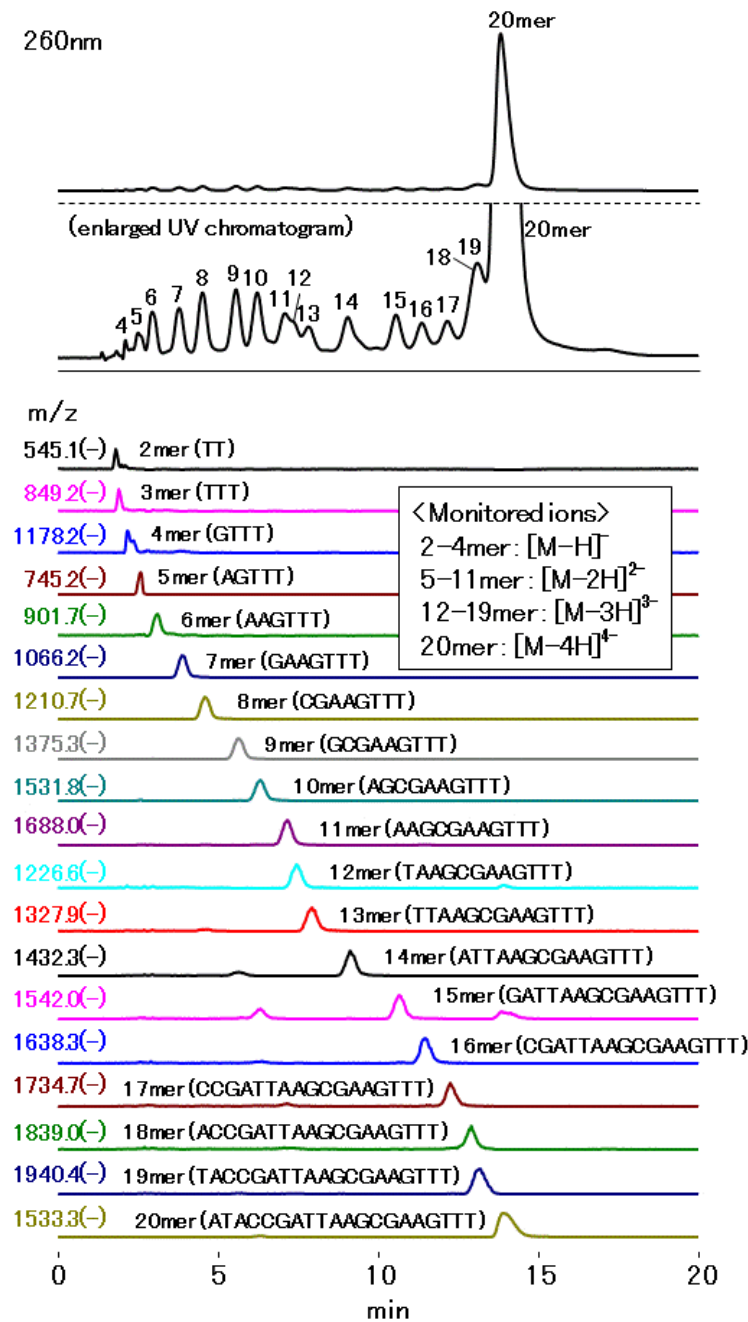
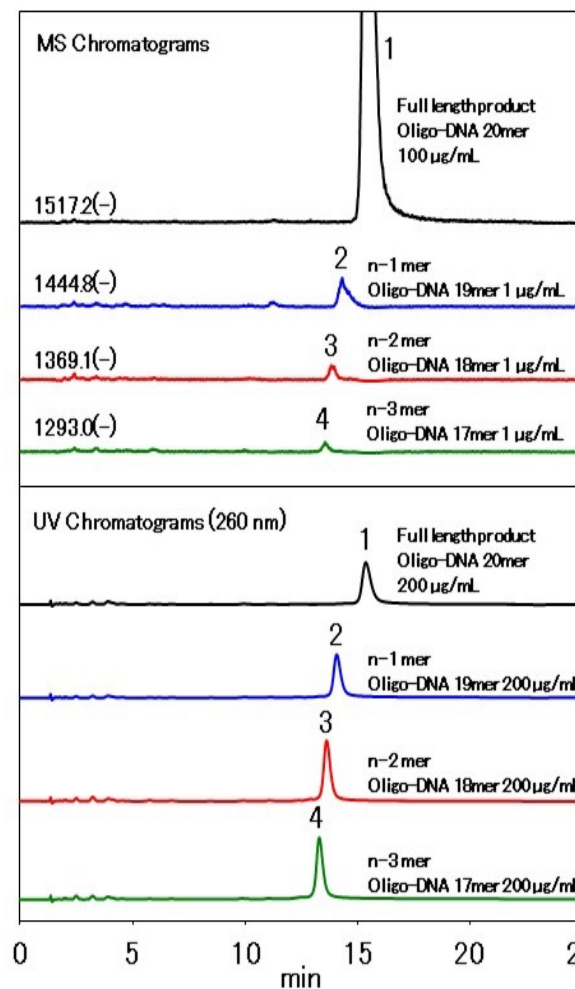
- 1 **HILIC-NH₄OAc**
- 2 RP-NH₄OAc
- 3 TRP BUFFERS

HILIC WITH NH₄OAc

- HILIC-MS analysis of oligonucleotides (Easter 2010) without the use of ion-pair reagents (Loubé/MacNeil 2019)
- Applicable to oligodeoxy(ribo)nucleotides and phosphorothioates (PS)
- Improved LC-MS sensitivity over ion-pair reverse phase methods
- Additional chromatographic developments required to match UV-based PS methods

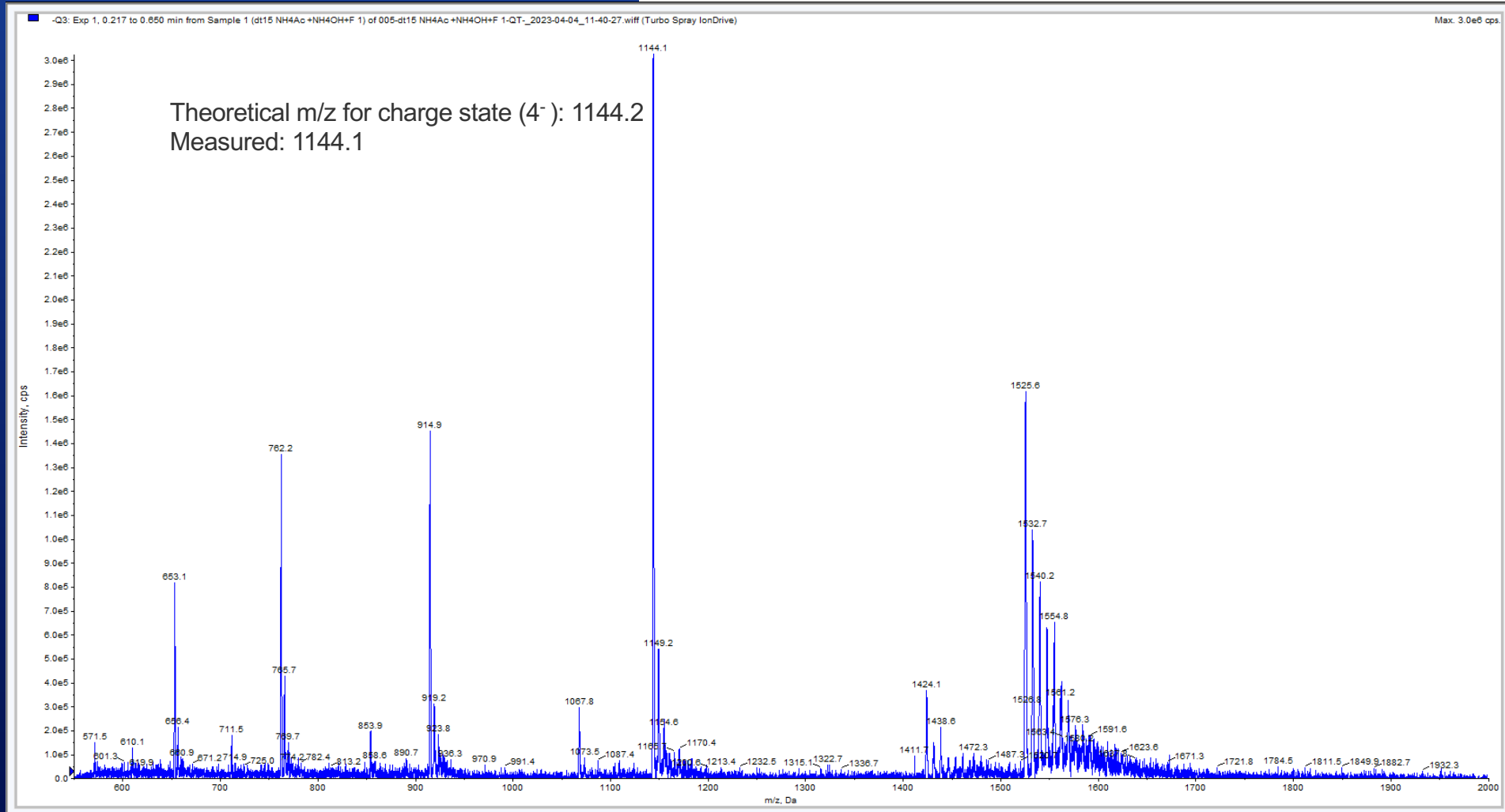


Shodex™ Technical Article No.4;
 LC/MS Analysis of Oligonucleotides
 Using a Polymer-Based Diol Column - Shodex™ HILICpak™ VN-50 2D



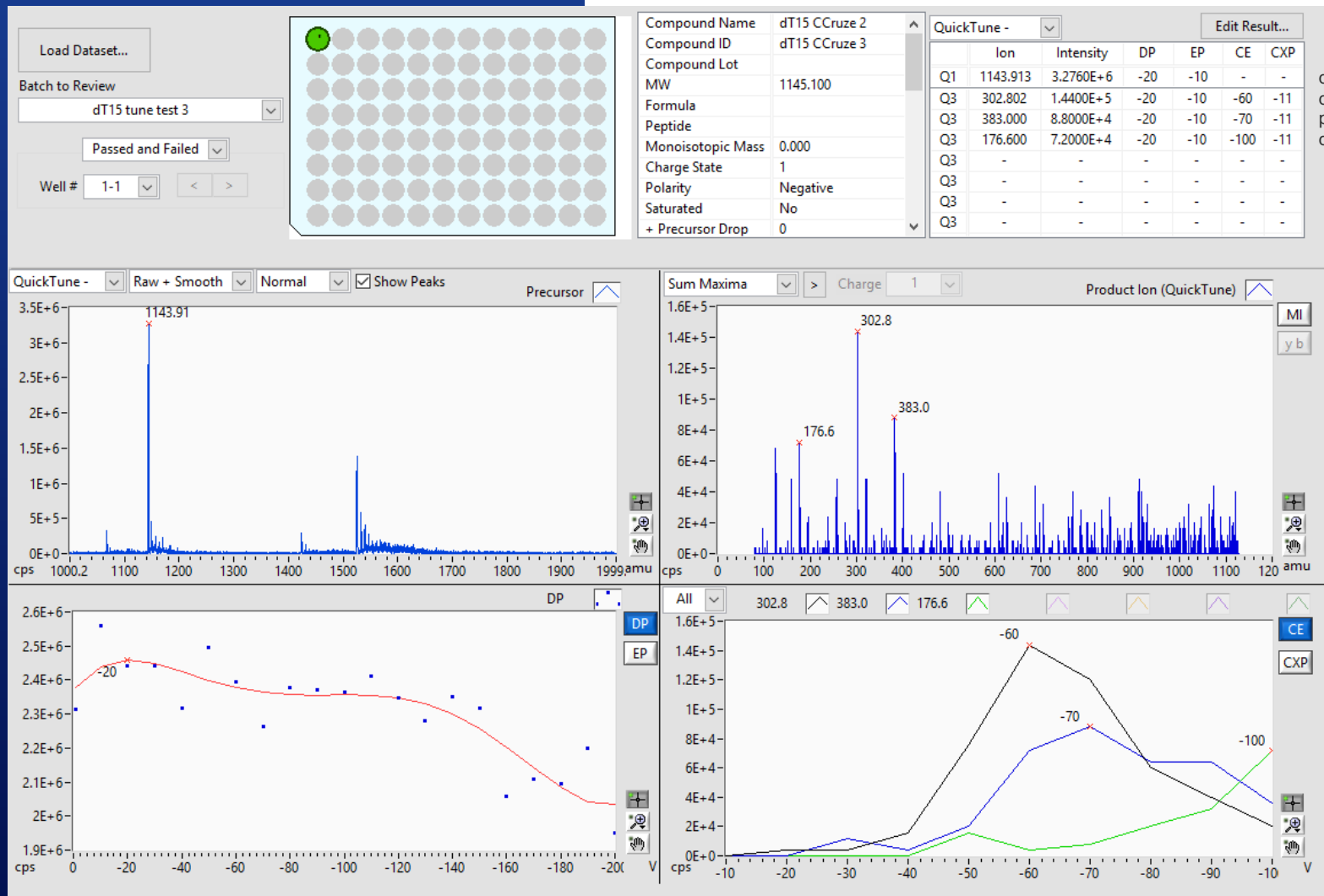
Mass Spectrum (after HILIC)

(pdT)₁₅, NH₄OAc in MeCN/H₂O, Shodex HILIC



MRM Optimization

(pdT)₁₅ in H₂O



dT15 (4-)
dTp-H2O (-)
pdTp-H2O (-)
dTp-T-H2O (-)

HPLC GRADIENT

(pdT)₁₅ in H₂O

Configure Pump

File Edit Table Help

Pump: Agilent - Stream 1 ext

Max Pressure: 2176 psi
Min Pressure: 0 psi
Flow Rate: 1.000 ml/min
Flow Composition: 100.000 % B
Solvent Valve A: A2
Solvent Valve B: B2
Acq Window Start: 0 min
Acq Window Duration: 0 min

Time Program

	Time [min]	Flow Rate	% A	% B
1	0.000	1.000	0.000	100.000
2	0.150	1.000	0.000	100.000
3	0.610	1.000	70.000	30.000
4	0.640	1.000	70.000	30.000
5	0.700	1.000	70.000	30.000
6	0.701	1.500	0.000	100.000
7	0.810	1.500	0.000	100.000
8				

Concentration, %

Time min

OK Cancel

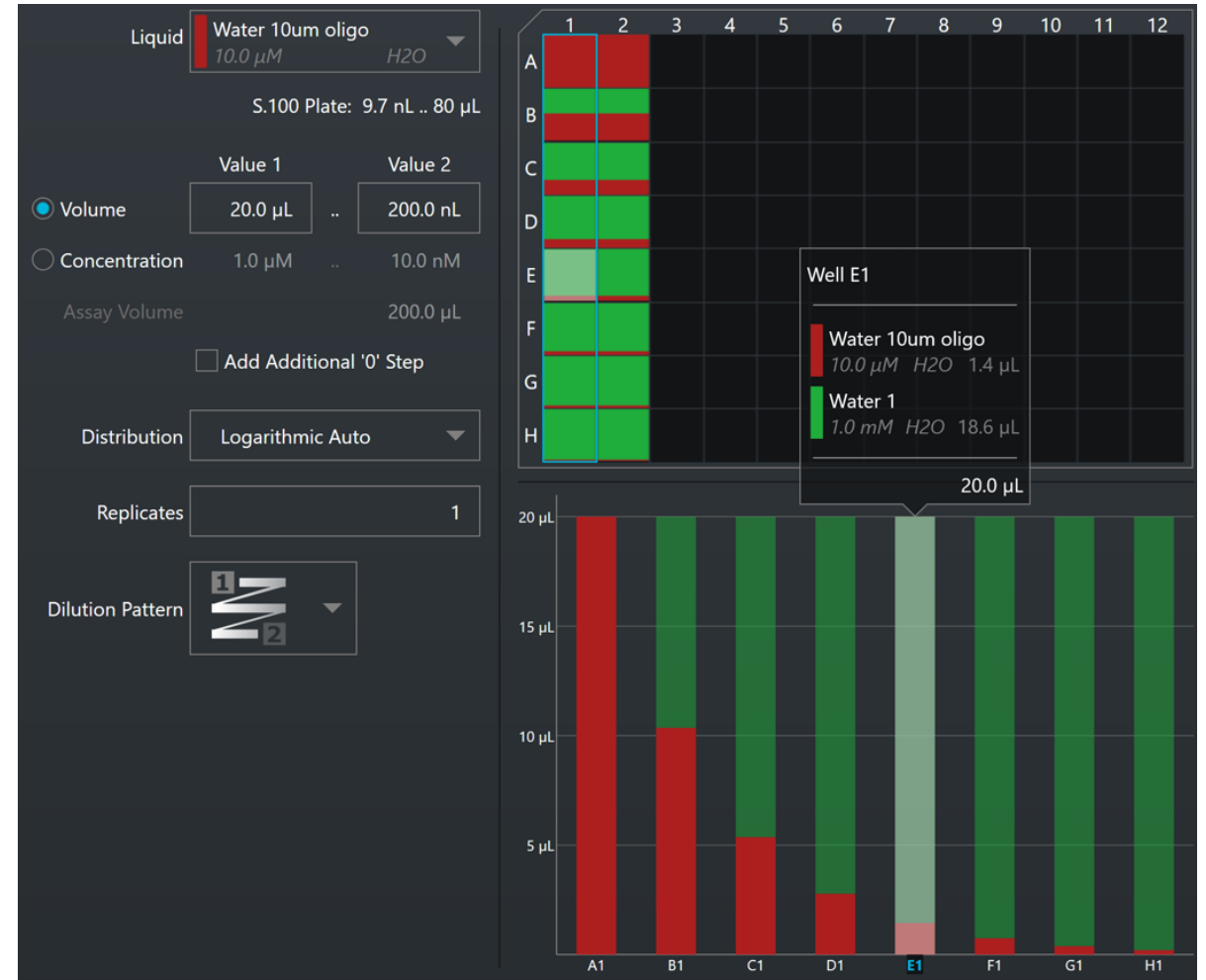
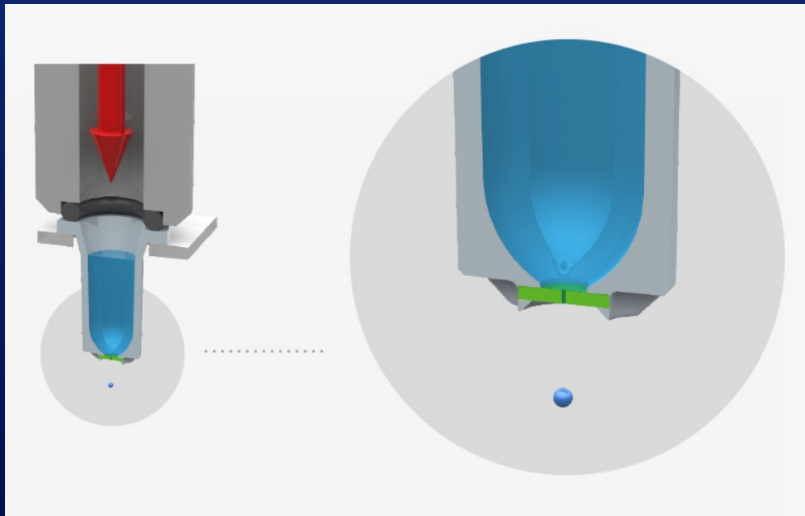
CALIBRATION SAMPLE PREPARATION

(pdT)₁₅ in H₂O or buffer

Instrument: iDOT non-contact liquid handler

QC preparation separately

Time needed: <1min/96 well

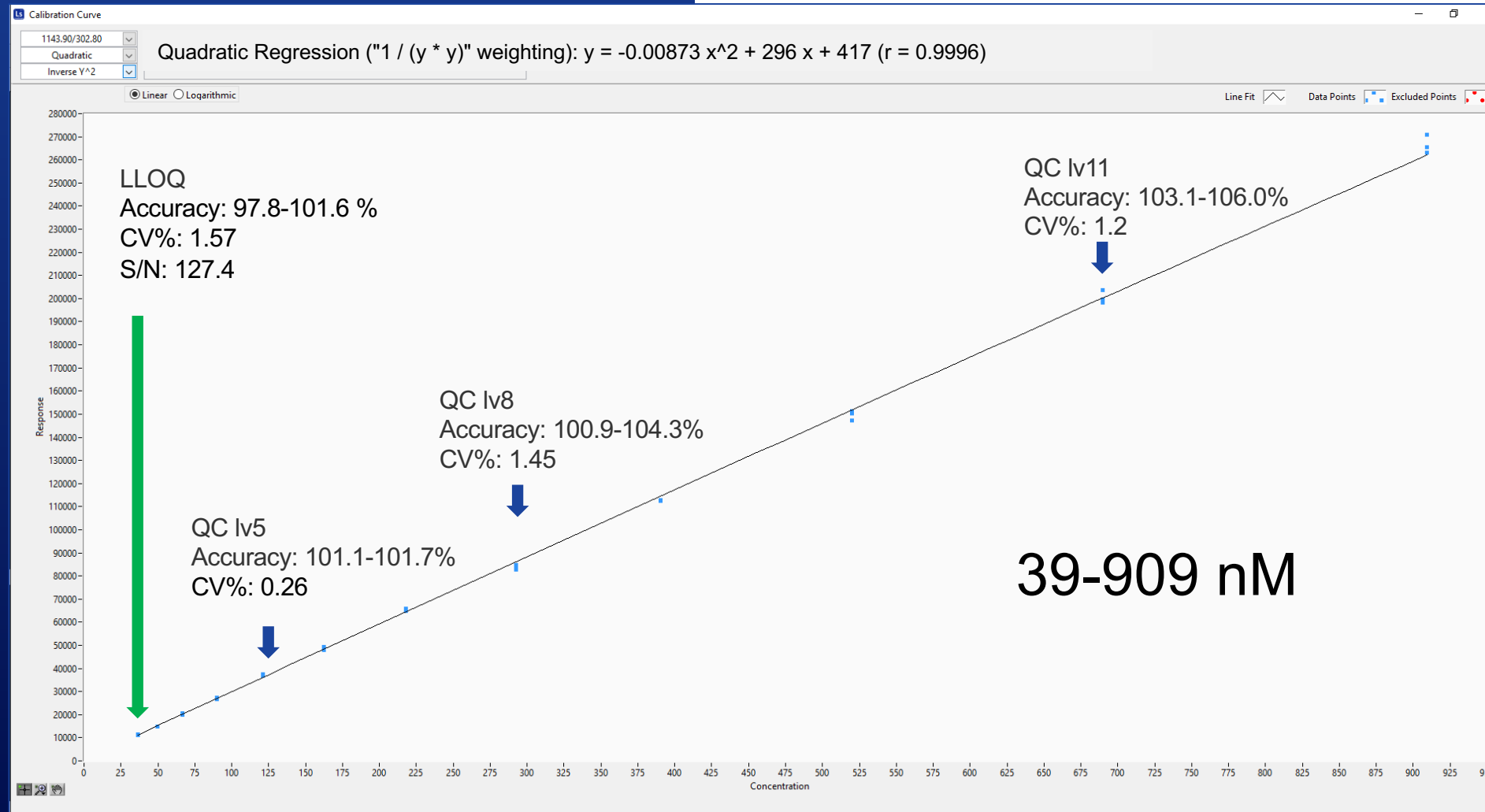


CALIBRATION

(pdT)₁₅ in H₂O, NH₄OAc in MeCN/H₂O, Shodex HILIC

The benefits of HILIC for oligos (R. MacNeil et al., Bioanalysis (2019) 11(12), 1155–1167):

- less signal suppression and variability
- less instrumental downtime for maintenance and cleaning
- less frequent analytical column changes
- no need for exotic flushing processes to switch over of instrumentation for other methods
- smoother workflows and better efficiency



HILIC CALIBRATIONS OF 12 DNA PRIMERS

18-25mers in H₂O, NH₄OAc in MeCN/H₂O, Shodex HILIC

Target	R/F	Sequence	Mass	Charge	Transition	S/N	LLOQ nM	ULOQ nM	QC 1 (99nM)	QC 1 (99nM)	QC 2 (244nM)	QC 2 (244nM)	QC 3 (593nM)	QC 3 (593nM)
1 hOATP4C1 R	R	CTACAATACCTTGCGTGACGGC	6695	-5	1337.65/1307.40	11.9	39	909	105.30%	104.10%	102.60%	104.70%	103.30%	102.70%
2 SVCT2 (SLC23A2) R	R	AAGCTAGGAGCCCAGGATCA	6160	-4	1538.40/1501.10	54.2	39	909	96.00%	92.90%	97.40%	95.10%	104.60%	98.30%
3 SVCT1 (SLC23A1) R	R	TAACCATGTGCTGGTCTGG	6164	-4	1539.54/1501.31	56.8	39	909	96.00%	100.50%	98.60%	102.40%	103.40%	102.60%
4 pig Bcrp (Abcg2) R	R	CTGCTGAAACACTGGTTGGTT	6452	-5	1289.13/624.73	34.2	39	909	101.60%	98.20%	103.80%	103.00%	101.80%	105.00%
5 cyMDR1 R	R	ACAGTGTCAAGTTGCCAACCA	6086	-4	1520.02/1486.30	42.9	39	909	109.10%	95.50%	99.20%	100.70%	101.70%	99.00%
6 hOATP4C1 F	F	AAATCGAAGTCTCTGCCTTGTCTC	7568	-5	1512.13/1074.15	31.8	39	909	103.50%	109.30%	101.10%	104.30%	101.00%	96.00%
7 SVCT2 (SLC23A2) F	F	GCTGCAGCCAGCTAGGTC	5501	-4	1373.59/1336.00	79.0	39	909	104.40%	96.90%	103.00%	103.50%	100.10%	99.80%
8 SVCT1 (SLC23A1) F	F	GGCCTTTGTCAAGTCATCCC	6044	-4	1509.53/1472.06	32.1	39	909	96.30%	100.20%	99.30%	99.70%	98.30%	96.40%
9 SNAT1 (SLC38A1) F	F	GCTTTGGTTAAAGAGCGGGC	6213	-5	1241.25/1211.17	15.1	39	909	101.00%	97.10%	107.40%	108.30%	103.00%	100.80%
10 SNAT2 (SLC38A2) F	F	AATAGAGACCACCGAGGCCG	6145	-5	1227.71/1200.69	18.8	39	909	108.70%	91.70%	100.40%	93.80%	101.40%	103.60%
11 ATB0,+ (SLC6A14) F	F	TTGGGGTGGCTTAGTTGCTC	6186	-5	1235.99/1205.56	23.7	39	909	97.00%	101.30%	106.60%	103.10%	95.20%	98.20%
12 ASCT1 (SLC1A4) F	F	TCTCCTCGCCTTCTCGCAC	5931	-4	1481.29/769.90	74.2	39	909	101.80%	97.50%	106.20%	105.80%	97.20%	95.10%

WORKS FROM WATER

- Matrix: H₂O → SPE for any other matrices
- ADDA – Sciex 6500+ Triple Quadrupole MS
- Negative MRM charge state of parent: -4 or -5
- Working Range 39-909 nM
- Signal-to-noise at LLOQ ≥ 11.9
- Accuracy of low QC (99 nM) 91.7-109.3%

Quant

- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.0119 x^2 + 45.7 x - 36$ (r = 0.9982)
- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.00876 x^2 + 130 x - 744$ (r = 0.9984)
- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.0154 x^2 + 189 x - 823$ (r = 0.9988)
- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.00686 x^2 + 29.8 x + 136$ (r = 0.9973)
- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.0236 x^2 + 193 x - 443$ (r = 0.9987)
- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.00919 x^2 + 68.5 x + 86$ (r = 0.9995)
- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.0202 x^2 + 173 x - 791$ (r = 0.9992)
- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.0102 x^2 + 131 x - 63.1$ (r = 0.9990)
- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.0262 x^2 + 91.9 x + 400$ (r = 0.9972)
- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.00991 x^2 + 33 x + 31.1$ (r = 0.9952)
- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.00396 x^2 + 81.3 x - 259$ (r = 0.9932)
- Quadratic Regression ("1 / (y * y)" weighting): $y = 0.0866 x^2 + 218 x + 1.46e+003$ (r = 0.9987)

- ASO, ssDNA, ssRNA, thioates up to ~25mers, longer run time for siRNA, dsDNA
- MS limitations – mass range and CID optimization

SPE OF OLIGOS

High-Throughput

Clarity® OTX™ was designed with the fast-paced DMPK/ADME environment in mind. It is a simple, rapid, and reproducible solution that efficiently extracts oligos from biological matrices and can be easily automated to eliminate sample backlogs and meet critical deadlines. By eliminating the need for LLE (liquid-liquid extraction), providing a 96-well plate format for liquid handler compatibility, and specifically targeting synthetic oligo therapeutics chemistries, Clarity OTX delivers a 15-minute extraction procedure.

Suitable for Most Oligo Therapeutics & Samples

Oligo Types:	Sample Types:
DNA	Plasma
Aptamers	Serum
RNAi/siRNA	Urine
Thioates	Tears
Lipid-conjugates	Saliva
Liposome encapsulated	Tissue

SPE OF OLIGOS

Oasis Cartridges and 96-Well Plates

Product Description

The Oasis WAX 96-well plate contains the Oasis WAX sorbent, which is a polymeric reversed-phase, weak anion exchange mixed-mode sorbent that has been optimized for fast, simple, and highly selective sample preparation of strong acidic compounds. The unique balance of hydrophobicity and water-wettability of the Oasis WAX sorbent means you will never have to worry about poor results if individual wells of the 96-well plate dry out during the critical steps prior to sample loading. The Oasis WAX 96-well plate is designed to be used on many manifold configurations and most robotic liquid handling systems.

Optimized sample preparation

The solid-phase extraction (SPE) of RM1 and the analog internal standard VA1 from human plasma was performed as follows. The SPE sorbent was Waters Oasis[®] WAX, 10 mg, a mixed-mode phase with cation exchange and reversed-phase moieties, in 96-well format. The 96-well 1 ml collection plates were regular inert polypropylene from Porvair (Wrexham, UK). Each step where liquid was applied was performed with the minimal aid of positive pressure to help percolation and passage of the liquid sample through the sorbent bed. The positive pressure manifold was from Agilent Technologies (DE, USA)

The analog internal standard VA1, in 1:1, v:v, acetonitrile:water, at 2500 nM, was added in 20 µl aliquots to 100 µl plasma within 1.5 ml regular polypropylene tubes. This resulted in an internal standard concentration of 500 nM in matrix. Then, a two-second vortex of each tube took place. This was followed by the addition of 225 µl 4.5% H₃PO₄ (aq) to each sample and another vortex step.

An oligonucleotide bioanalytical LC–SRM methodology entirely liberated from ion-pairing

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Note: At time of writing: Envigo CRS, PO Box 2360 Mettlers Road, East Millstone, NJ 08875-2360, USA

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HT-LC- MS/MS OF OLIGOS

- 1 HILIC-NH₄OAc
- 2 **RP-NH₄OAc**
- 3 TRP BUFFERS

RP WITH NH₄OAc

NH₄OAc vs. Ion Pair agents

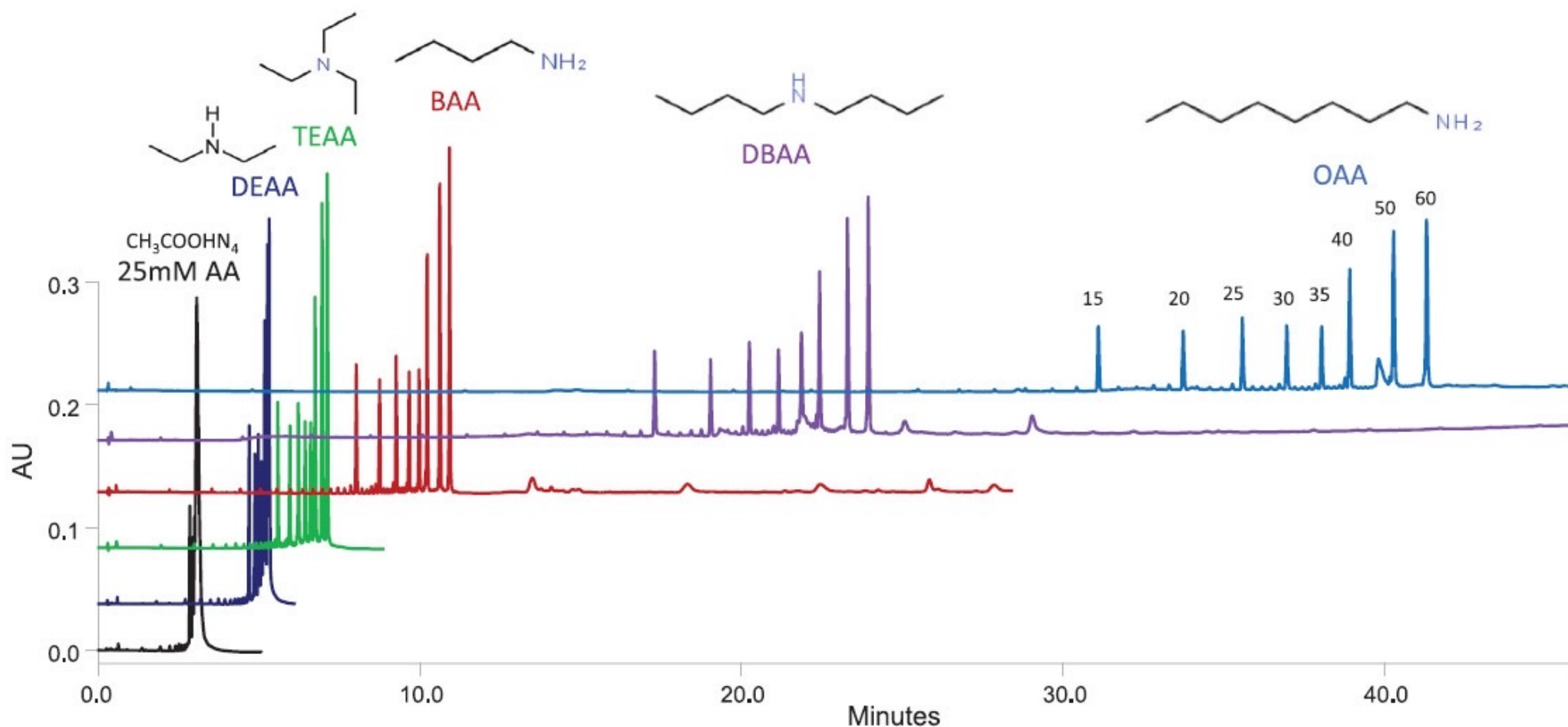
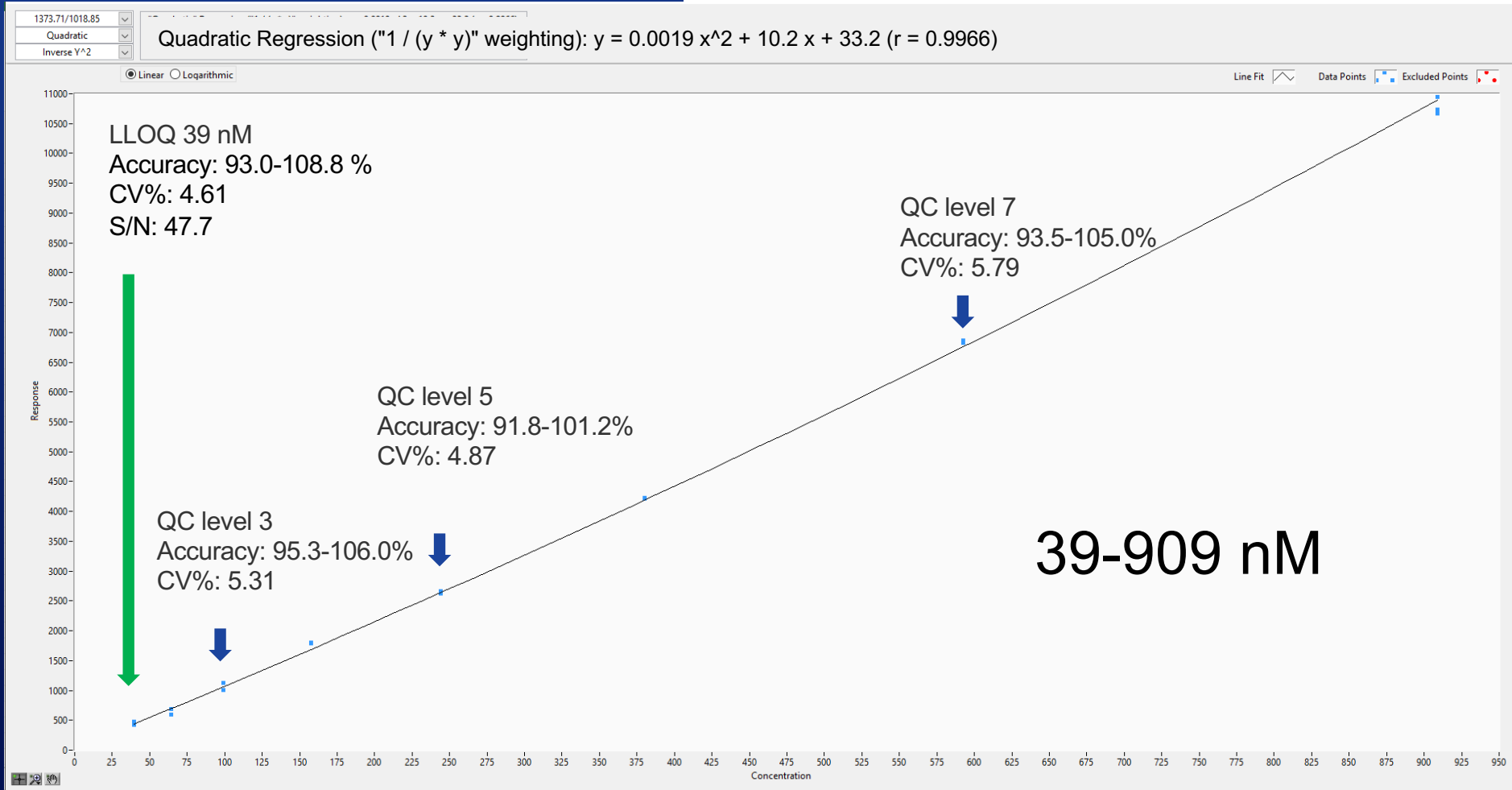


Fig. 2. Separation of 15, 20, 25, 30, 35, 40, 50, and 60 mer oligodeoxythymidines using selected 100 mM ion-pairing buffers. The data are compared to non-ion-pairing 25 mM ammonium acetate buffer. Retention and peak capacity data are listed in Table 4. For separation conditions see Experiment 2 in Table 3.

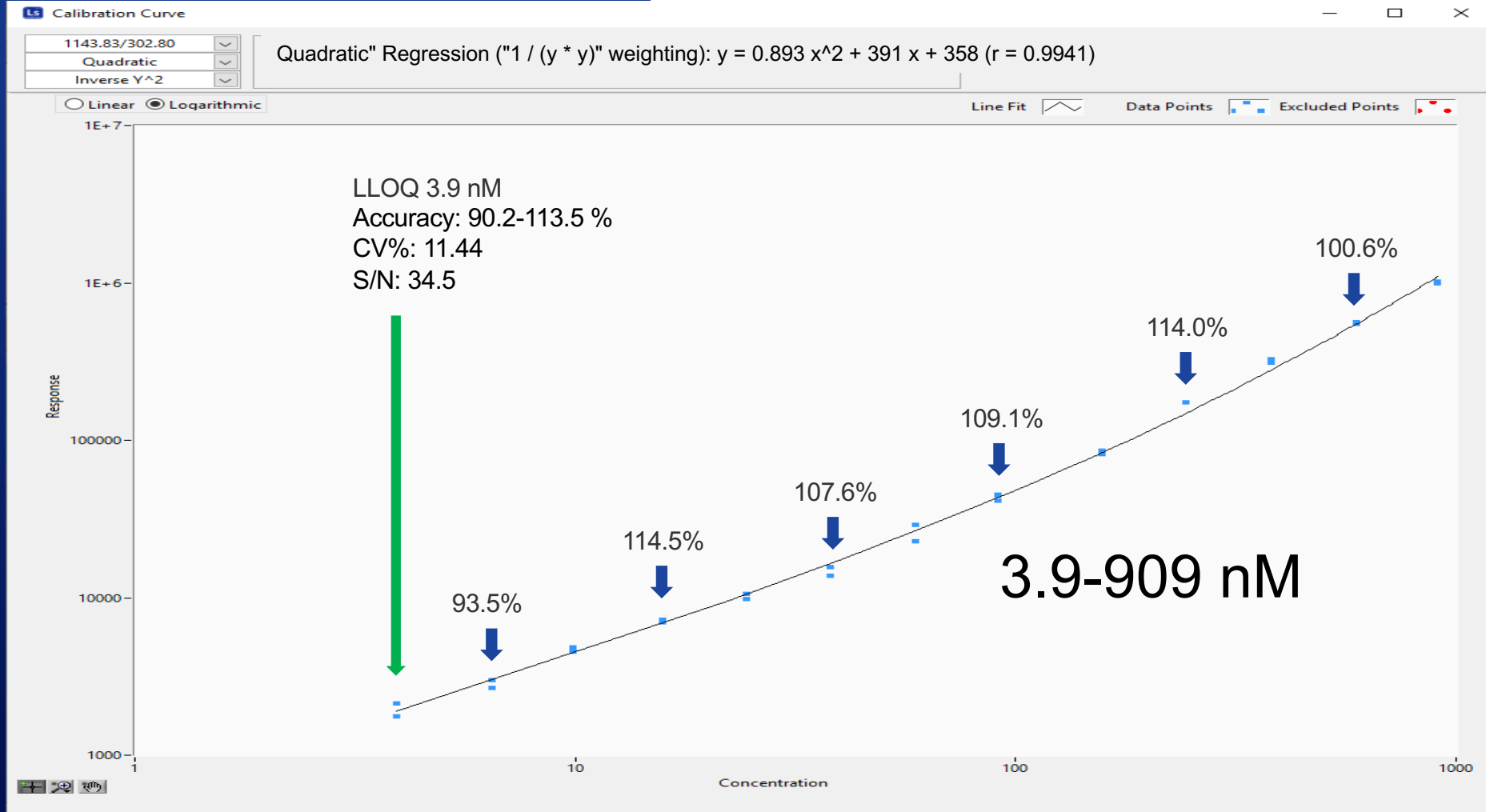
CALIBRATION

- GCTGCAGCCAGCTAGGTC (SVCT2 F) in HBSS, NH₄OAc in H₂O/MeCN, Hypersil Gold C18, 10*2.1mm, 3um
- ADDA-6500+
- MRM (4-)
- S/N 47.7



CALIBRATION (Inj.: 50 µL)

- (pdT)₁₅ in HBSS, NH₄OAc in H₂O/MeCN, Hypersil Gold C18, 10*2.1mm, 3 µm
- ADDA-6500+
- MRM (4-)
- S/N 34.5



HT-LC- MS/MS OF OLIGOS

- 1 HILIC
- 2 **NH₄OAc**
- 3 **TRP BUFFERS**

CALIBRATION (inj: 50 µL)

- GCTGCAGCCAGCTAGGTC (SVCT2 F) in BUFFER, NH₄OAc in H₂O/MeCN, Hypersil Gold C18, 10*2.1mm, 3 µm
- ADDA-6500+ MRM (4-)
- 10-1,000 nM

Buffer	QC1 (37.3 nM)		QC2 (139 nM)		QC3 (518 nM)		Analyte Quant
HBSS	99.70%	95.80%	91.80%	92.70%	94.10%	99.90%	Quadratic Regression ("1 / (y * y)" weighting): $y = 0.118 x^2 + 140 x + 96.2$ (r = 0.9964)
KH	88.00%	97.00%	89.70%	96.00%	100.80%	102.10%	Quadratic Regression ("1 / (y * y)" weighting): $y = 0.118 x^2 + 143 x + 904$ (r = 0.9969)
SM	111.50%	107.40%	100.50%	94.40%	96.10%	103.60%	Quadratic Regression ("1 / (y * y)" weighting): $y = 0.109 x^2 + 204 x + 559$ (r = 0.9974)
BSEP	108.30%	105.70%	107.80%	98.70%	98.00%	99.90%	Quadratic Regression ("1 / (y * y)" weighting): $y = 0.159 x^2 + 44.6 x + 145$ (r = 0.9928)

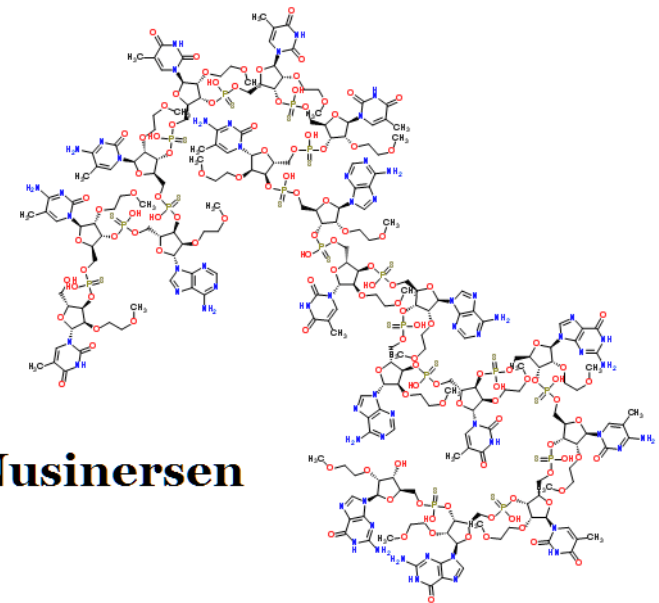
HBSS: Hank's balanced salt solution
 KH: Krebs-Henseleit Buffer pH 7.4
 SM: Start mix
 BSEP: Bile Salt Export Pump

Buffer TEST

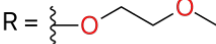
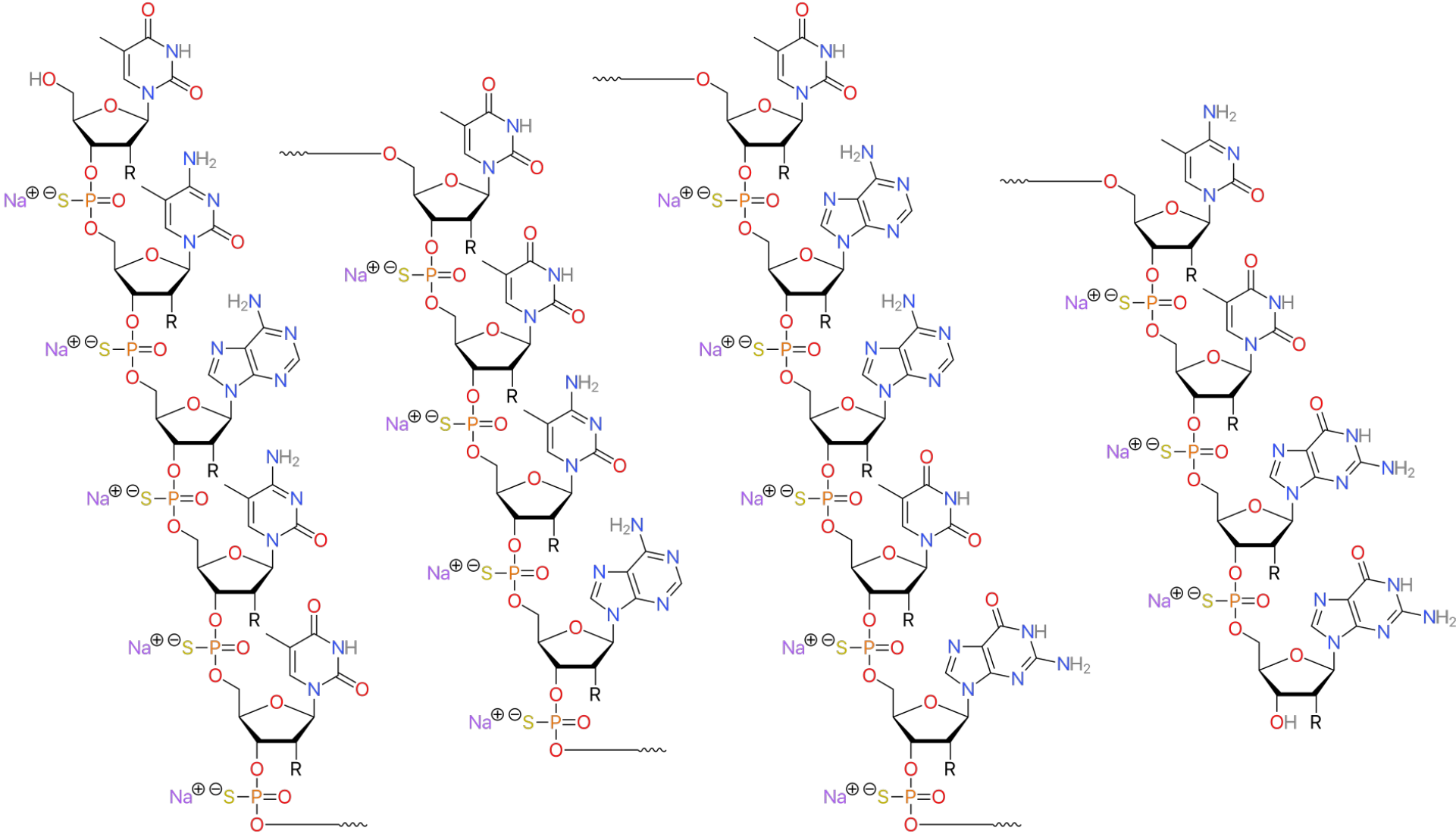
1 μ M 18mer (SVCT2), 100 nM (pdT)₁₅ IS

	TA/IS			Mean	Stdev	CV%
H2O	1.09	1.08	1.01	1.06	0.04	3.45%
HBSS	3.86	3.72	3.70	3.76	0.07	1.83%
KH	3.72	3.87	4.00	3.86	0.11	2.94%
SM	4.69	4.87	4.78	4.78	0.07	1.49%
BSEP	3.42	3.53	3.74	3.56	0.13	3.73%
	Peak Area			Mean	Stdev	RSD%
H2O	26337	32698	27822	28952	2717	9.38%
HBSS	25238	28471	28293	27334	1484	5.43%
KH	22378	24408	26135	24307	1535	6.32%
SM	20427	22305	24999	22577	1876	8.31%
BSEP	15352	21027	21544	19308	2805	14.53%

Uptake transporter assay with HT- LC-MS/MS



Nusinersen / Spinraza



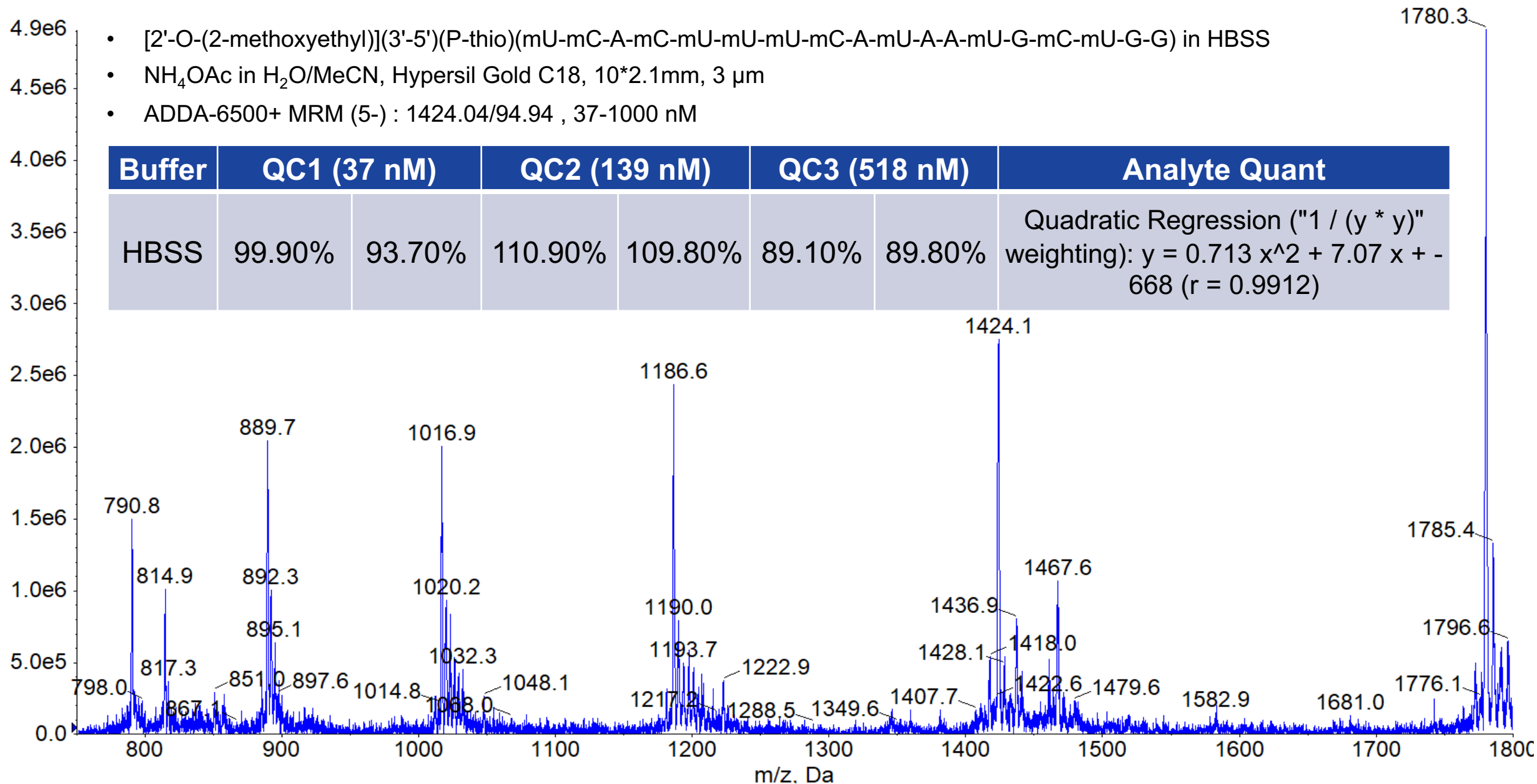
CALIBRATION

Max. 4.9e6 cps

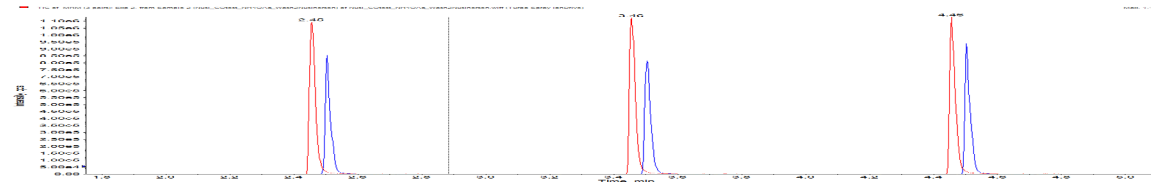
- [2'-O-(2-methoxyethyl)](3'-5')(P-thio)(mU-mC-A-mC-mU-mU-mU-mC-A-mU-A-A-mU-G-mC-mU-G-G) in HBSS
- NH₄OAc in H₂O/MeCN, Hypersil Gold C18, 10*2.1mm, 3 μm
- ADDA-6500+ MRM (5-) : 1424.04/94.94 , 37-1000 nM

Buffer	QC1 (37 nM)		QC2 (139 nM)		QC3 (518 nM)		Analyte Quant
HBSS	99.90%	93.70%	110.90%	109.80%	89.10%	89.80%	Quadratic Regression ("1 / (y * y)" weighting): $y = 0.713 x^2 + 7.07 x - 668$ (r = 0.9912)

Intensity, cps



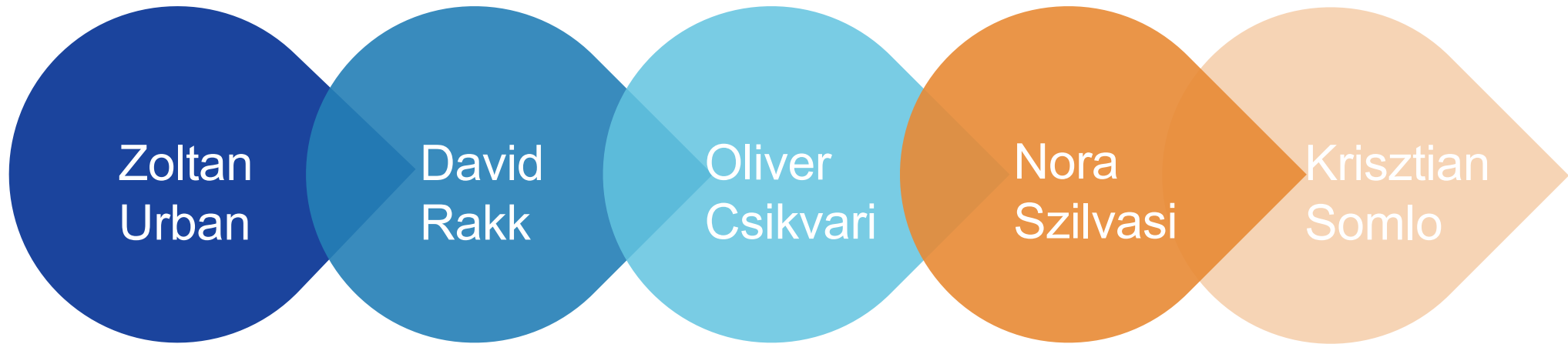
Summary



High-throughput quantitative measurement of 15-25 mer ssDNA from preclinical *in vitro* buffers with 1 minute cycle time without strong ion-pair reagents was performed applicable for small therapeutic ASOs.

Method	Matrix	Sample preparation	Benefits	Disadvantages
IP-RP-MS	MIC	OE	<ul style="list-style-type: none"> • Generic • Great separation 	<ul style="list-style-type: none"> • Signal suppression • Variability • Contamination related MS down-time, maintenance, cleaning, long switch procedure • Long acquisition • HPLC column degradation
HILIC-MS	Water	SPE	<ul style="list-style-type: none"> • Easy switch bw methods • Fast • Good separation 	<ul style="list-style-type: none"> • Matrix effect
RP-MS	MIC	Simple	<ul style="list-style-type: none"> • Low LLOQ • Easy switch bw methods • Fast 	<ul style="list-style-type: none"> • LC selectivity • ds oligos separation

Acknowledgement



- Screen group lead
- Senior scientist
- Scientist
- RnD Scientist
- Assistant