

# The pursuit of robust quantification of oligonucleotides by hybridization assays

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**labcorp**

# Assay reagent quality

## Assay development case study

- Developing a dual probe hybridization assay for an aptamer therapeutic
- Assay framework was provided by the client
- Biotin- and Digoxigenin-labeled probes were obtained from a commercial vendor



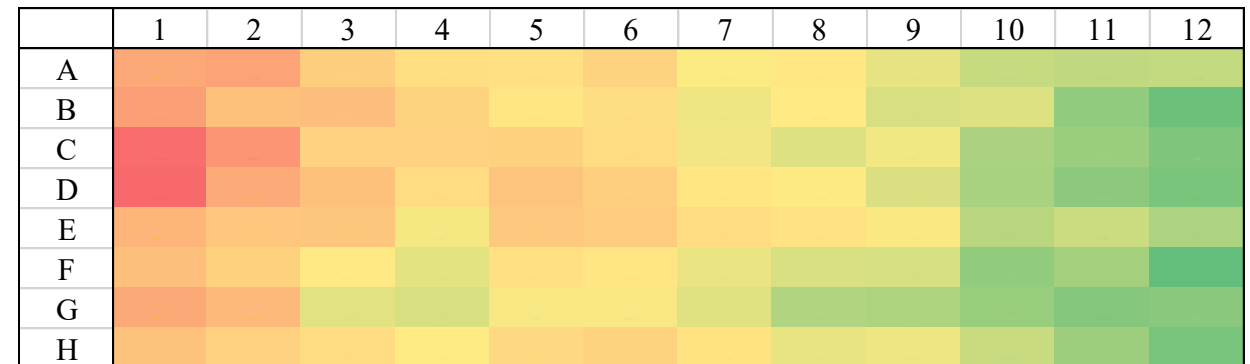
# Assay reagent quality – impact on assay performance

## Assay development summary

### Phase 1 – Fine tune from provided method

- Commercial probes
- 7 runs over 6 days for optimization
- 2 runs on Qualification Day 1
  - Fail for QC bias

Intra-Run A&P – 6 QCs at each level				
QC bias (# out of 6)				
	0-10%	10-20%	>20%	Fail
ULOQ	0	1	5	5
HQC	1	4	1	1
MQC	6	0	0	0
LQC	6	0	0	0
LLOQ	6	0	0	0



Whole plate precision (% CV) approximately 5%

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  - Fail for QC bias

### Phase 2 – Re-optimization

- Commercial probes
- 14 runs over 6 days to adjust assay parameters
  - Probe concentrations increased
- 4 runs over 2 Qualification days
  - Fail for QC bias, selectivity, stability

A&P – 3 QCs at each level				
QC bias (# out of 3)				
	0-10%	10-20%	>20%	Fail
ULOQ	0	2	1	1
HQC	0	2	1	1
MQC	2	1	0	0
LQC	0	2	1	1
LLOQ 1	0	2	1	0
LLOQ 2	2	0	1	0

Dilution linearity				
Bias (# out of 5)				
	0-10%	10-20%	>20%	Fail
In Range	0	4	1	1

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  - Fail for QC bias, selectivity, stability

### Phase 3 – Investigation

- **Fresh vs. frozen samples, location effects, incubation times**
- **38 total runs over 3 phases without transitioning to validation**

# Assay reagent quality – impact on assay performance

## Assay development summary

### Phase 4 – New probes produced by client

- High probe concentration → vendor supply issue and quality concerns
- 2 runs over 2 days for optimization
  - Range remains 50-fold, shifted up by factor of 2
- 8 runs to successfully qualify assay for validation
- 10 total runs to re-optimize and complete the development



# Assay reagent quality – impact on assay performance

## Assay development summary

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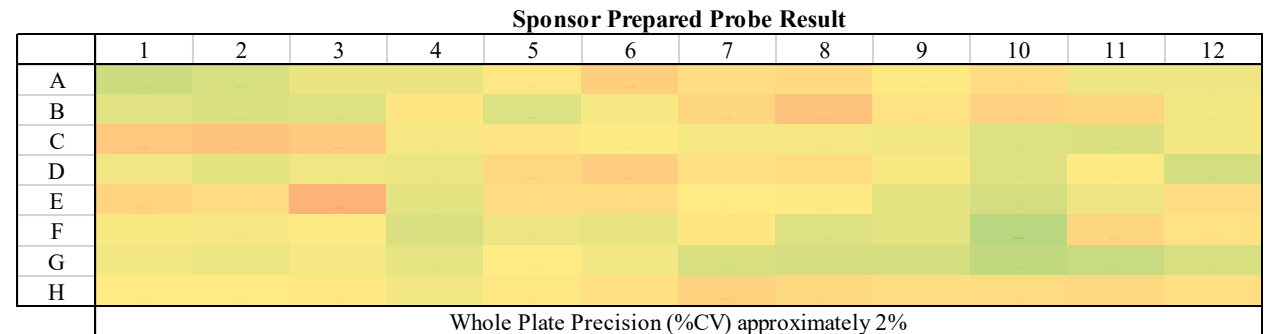
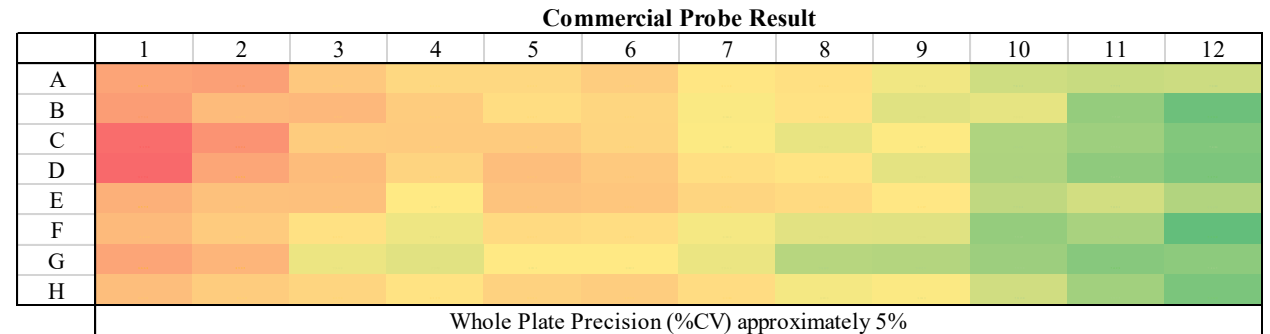
Dilution linearity				
Bias (# out of 6)				
	0-10%	10-20%	>20%	Fail
In Range	5	1	0	0

QC bias results					
		0-10%	10-20%	>20%	Fail
Run A: A&P 6 QCs at each level	ULOQ	5	1	0	0
	HQC	3	3	0	0
	MQC	5	1	0	0
	LQC	1	5	0	0
	LLOQ	4	2	0	0
Run B: A&P 3 QCs at each level	ULOQ	2	1	0	0
	HQC	3	0	0	0
	MQC	1	2	0	0
	LQC	3	0	0	0
Run C: 2 QCs at H/M/L	LLOQ	2	1	0	0
	HQC	2	0	0	0
	MQC	1	1	0	0
	LQC	1	1	0	0

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# Assay procedure control

## Assay development case study

- Dual probe hybridization assay for an ASO therapeutic
- Biotin- and Digoxigenin-labeled probes were obtained from a commercial vendor
- Attempt to transfer a fully validated assay
  - Streptavidin plate and probe lots previously noted as consequential to assay performance
  - Hybridization temperature was changed from 37°C to ambient temperature for better sensitivity



# Assay procedure control – impact on assay performance

## Assay development summary

- Many assay parameters verified/unchanged
  - MRD, probe concentrations
- Could not achieve the same LLOQ
  - Quantitative range shifted and shrunk
- Hybridization temperature changed from 37°C to ambient room temperature to achieve a lower LLOQ

Parameter	Transferred assay	In our lab	Adjusted assay
Quantitative Range	1.5 – 200 nM	4 – 150 nM	1 – 100 nM
Sample Incubation Temp.	37°C	37°C	ART

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Quantitative Range	1.5 – 200 nM	4 – 150 nM	1 – 100 nM
Sample Incubation Temp.	37°C	37°C	ART

24 hr Room Temperature Stability				
	LQC (3 nM)		HQC (75 nM)	
Sample	Result	%Bias	Result	%Bias
1	4.93	64.3	111	48.0
2	4.78	59.3	101	34.7
3	4.98	66.0	99.9	33.2

Assay Development Summary

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ART Sample Incubation

37°C Sample Incubation

24 hr Room Temperature Stability					24 hr Room Temperature Stability				
	LQC (3 nM)		HQC (75 nM)			LQC (30 nM)		HQC (225 nM)	
Sample	Result	%Bias	Result	%Bias	Sample	Result	%Bias	Result	%Bias
1	4.93	64.3	111	48.0	1	33.7	12.3	243	8.0
2	4.78	59.3	101	34.7	2	31.5	5.0	231	2.7
3	4.98	66.0	99.9	33.2	3	28	-6.7	209	-7.1

Assay Development Summary

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- Many assay parameters verified/unchanged
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Parameter	Transferred assay	In our lab	Adjusted assay	Final assay
<b>Quantitative Range</b>	1.5 – 200 nM	4 – 150 nM	1 – 100 nM	10 – 300 nM
<b>Sample Incubation Temp.</b>	37°C	37°C	ART	37°C

ART Sample Incubation					37°C Sample Incubation				
24 hr Room Temperature Stability					24 hr Room Temperature Stability				
	LQC (3 nM)		HQC (75 nM)			LQC (30 nM)		HQC (225 nM)	
Sample	Result	%Bias	Result	%Bias	Sample	Result	%Bias	Result	%Bias
1	4.93	64.3	111	48.0	1	33.7	12.3	243	8.0
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# Enhanced assay sensitivity

## Assay development case study

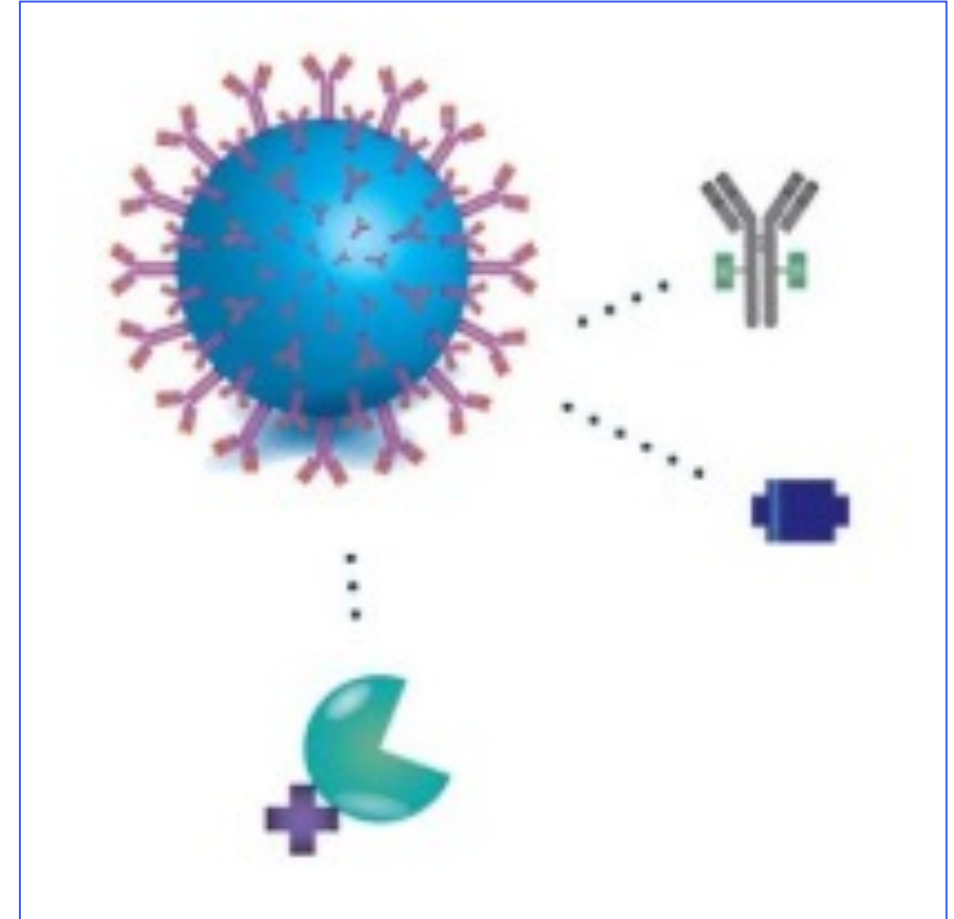
- Compare MSD ECL assay to Quanterix Simoa®
- Assays developed for 21-mer non-proprietary analyte
- Probe design
  - MSD ECL
    - 3' biotin for capture
    - 5' dig for detection
  - Quanterix Simoa®
    - 5' amine modification with spacer to couple to the beads
    - 3' biotin for detection with streptavidin-labeled enzyme



# Enhanced assay sensitivity with non-traditional LBA platform

## Quanterix Simoa® summary

- Traditional sandwich assay format in solution on magnetic beads
  - Streptavidin-labeled enzyme produces fluorescence
- Sample is loaded to a disk and travels across an array of small wells
- Fluorescence image is captured, fluorescent beads are counted
  - At higher sample concentrations, number of binding events per bead can be determined

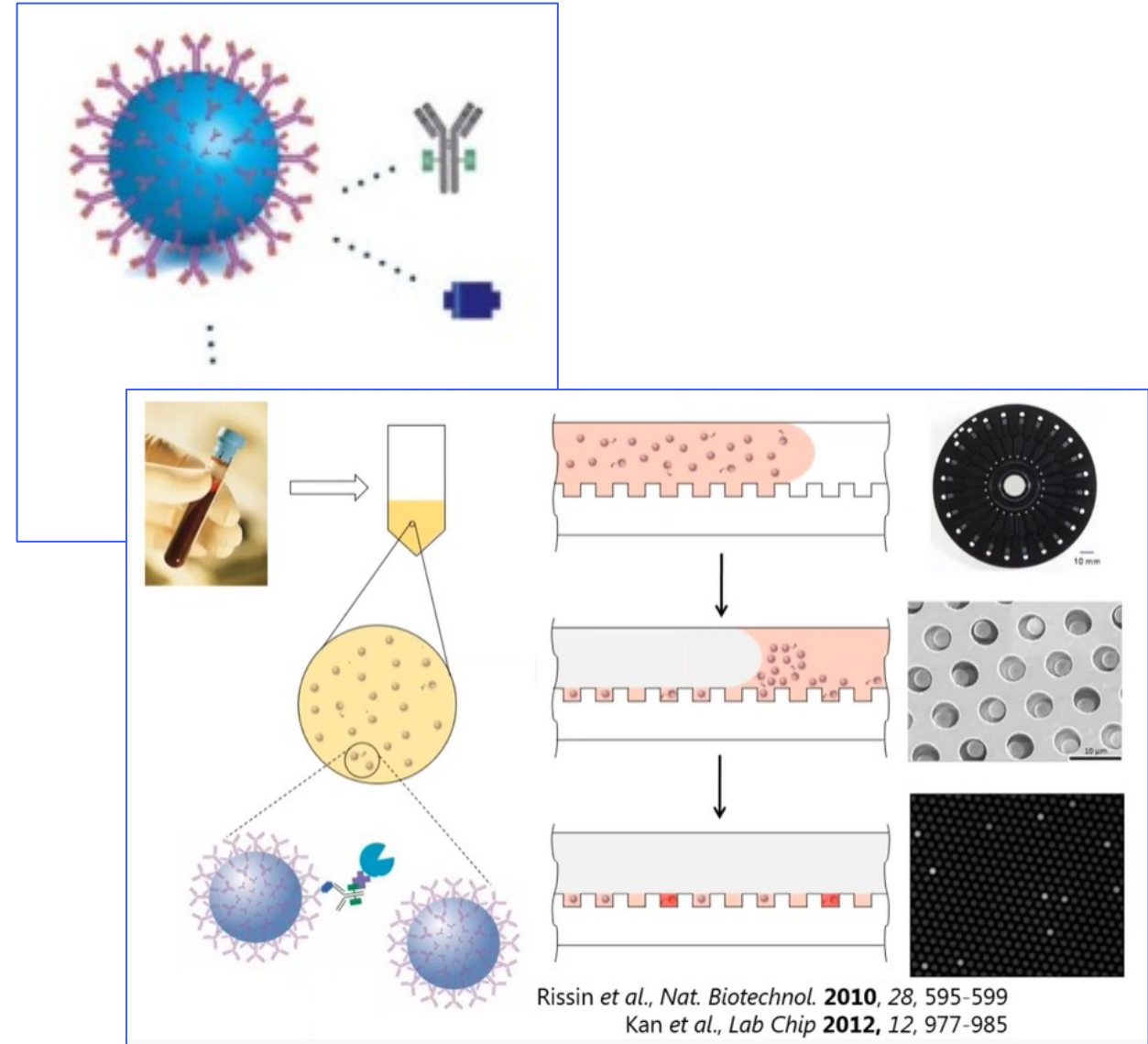




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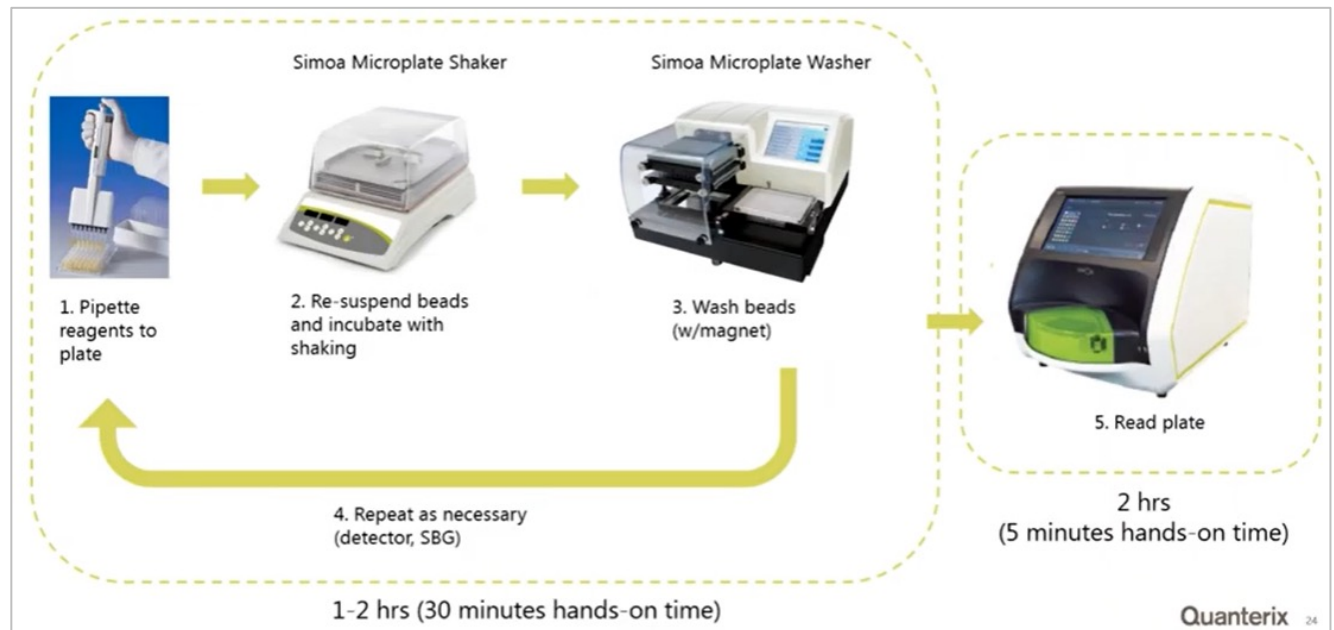




# Enhanced assay sensitivity with non-traditional LBA platform

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# Enhanced assay sensitivity with non-traditional LBA platform

## Simoa® Technology and ECL Comparison

### Simoa® Method outline (in human serum)

1. Dilute all samples in buffer
  2. Hybridization: Combine capture beads + diluted samples + detector probe and incubate (2 hour incubation)
  3. Wash beads and incubate with SBG enzyme (1 hour incubation)
  4. Wash beads and load onto instrument
  5. Plate reading (2-3 hours)
- Total Assay Time = 7-8 hours
- Readout signal: Fluorescence
- Reported raw data: AEB (Average number of Enzyme per Bead)

### ECL Method outline (in human serum)

1. Hybridization: Incubate capture probe + samples (1.5 hour incubation)
  2. Add hybridized probe/samples to streptavidin coated MSD plate and incubate (1-2 hour incubation)
  3. Wash plate and add digoxigenin-labeled detector probe (1 hour incubation)
  4. Wash plate and add Sulfo Tag-labeled anti-digoxigenin detection antibody (1 hour incubation)
  5. Wash plate and read
- Total Assay Time = 5-6 hours
- Readout signal: electrochemiluminescence
- Reported raw data: RLU

# Enhanced assay sensitivity with non-traditional LBA platform

## Simoa® Technology and ECL Comparison Accuracy & Precision

Simoa® Range I	Concentration (fM)	%Bias	%TE
ULOQ	10,000	-3.9	15.3
HQC	7500	-10.4	17.4
MQC	550	-9.8	16.9
LQC	60	-17.5	35.6
LLOQ	30	-30.6	54.2

Samples were tested in duplicates

Simoa® Range II	Concentration (fM)	%Bias	%TE
ULOQ	8000	7.5	19.6
HQC	6000	3.8	13.6
MQC	900	2.1	12.8
LQC	200	15.1	37.0
LLOQ	100	4.4	14.1

Samples were tested in triplicates

ECL	Concentration (fM)	%Bias	%TE
ULOQ	2,000,000	5.5	13.0
HQC	1,600,000	4.4	9.3
MQC	100,000	-2.3	7.0
LQC	10,000	-6.4	11.8
LLOQ	4000	-10.5	18.7

Samples were tested in duplicates

# Enhanced assay sensitivity with non-traditional LBA platform

## Simoa<sup>®</sup> Technology and ECL Comparison Final Results

Parameter	Simoa <sup>®</sup>	ECL
Lower Limit of Quantitation	100 fM	4000 fM
Upper Limit of Quantitation	8000 fM	2,000,000 fM
Sample Requirement per Test	25 µL	150 µL
MRD	1/14	1/3.3
Curve Fit; Weighting Factor	4-PL; 1/y <sup>2</sup>	4-PL; 1/y <sup>2</sup>
Method Selectivity	Pass	Pass
Dilution Linearity	1/500,000	1/40,000

# Enhanced assay sensitivity with non-traditional LBA platform

## Simoa<sup>®</sup> Technology and ECL Comparison Final Results

Parameter	Simoa <sup>®</sup>	ECL
Lower Limit of Quantitation	100 fM	4000 fM
Upper Limit of Quantitation	8000 fM	2,000,000 fM
Sample Requirement per Test	25 $\mu$ L	150 $\mu$ L
MRD	1/14	1/3.3
Curve Fit; Weighting Factor	4-PL; $1/y^2$	4-PL; $1/y^2$
Method Selectivity	Pass	Pass
Dilution Linearity	1/500,000	1/40,000

**40x improvement in sensitivity**

# Conclusions

- **Assay procedure details**
- **Reagent characteristics**
  - Probe quality
  - Probe design (LNA, other modifications)
- **Seeking new formats to meet assay demands**
  - Simoa<sup>®</sup> or other technologies



# Acknowledgments

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Thank you!  
Questions?

