

9<sup>th</sup> EBF Young Scientist Symposium



# How to adapt an ADA method to a vaccine efficacy study

**Swiss**  
**BioQuant**

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# What are biotherapeutics?

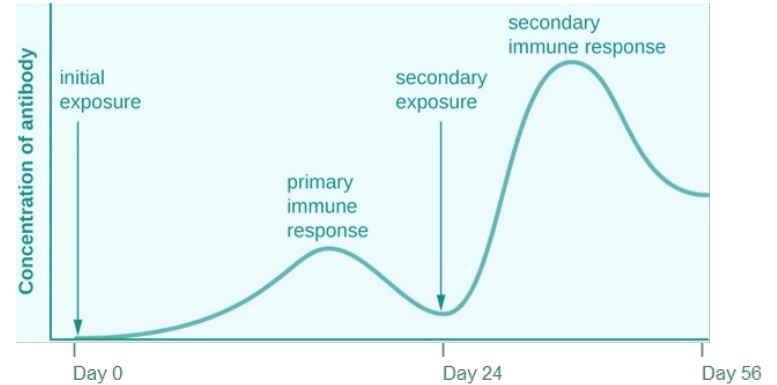
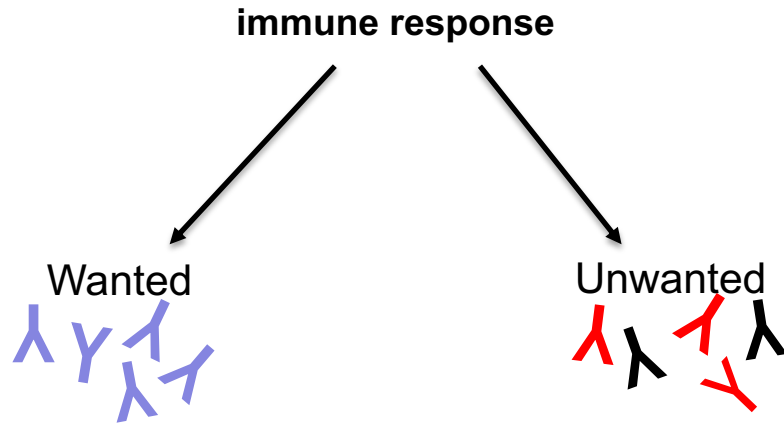
According to the **NLM-National Institutes of Health**, biotherapeutics are “antibody-drug cell therapy products where the active substance is extracted or produced from a biological source by a biotherapeutic technology.”

Biotherapeutic products include proteins and hormones, monoclonal antibodies, cytokines, antibodies, gene cell therapy products, vaccines, stem cell therapies, and more.



# Immunogenicity and immune response

Immunogenicity is the ability of a “non-self” substance to stimulate the immune system by provoking an



# Immunogenicity Assays

Nowadays immunogenicity assays are mostly used to monitor and detect the unwanted response of the immune system, but the same type of assay can be used for a different purpose or rather to measure the wanted reaction of the immune system after a vaccination

## ADA Investigation criteria

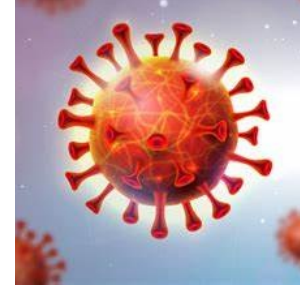
1. Screening cut point (SCP) and Confirmatory cut point (CCP)
2. Sensitivity and Precision
3. Selectivity
4. Drug tolerance
5. High dose hook effect
6. Stability

# Corona Virus

## COVID-19 pandemic:

The disease is mainly transmitted via respiratory route and the infection can occur over longer distances

→ Many novel candidate vaccines have been proposed

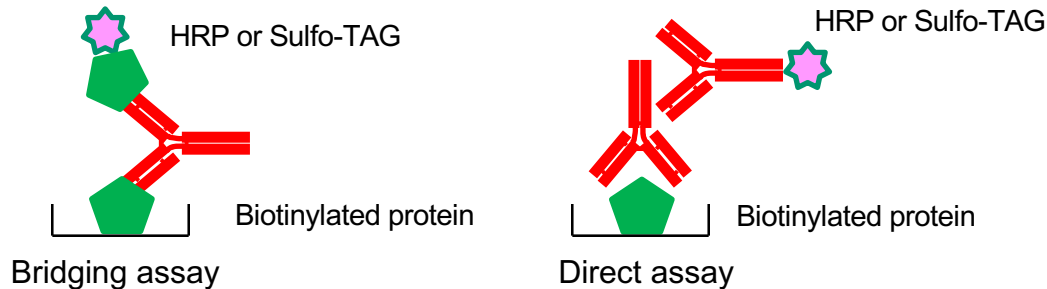


 **High request for new methods able to detect the immune response**

# Case Study

## Purpose of the study

- The purpose of this study was a validation of an analytical method for the determination of antibodies against SARS-COV-2 S protein (omicron variant) in rat serum by an electrochemi-luminescence immunoassay (ECLIA).



**Direct MSD immunoassay was used**

# Case Study

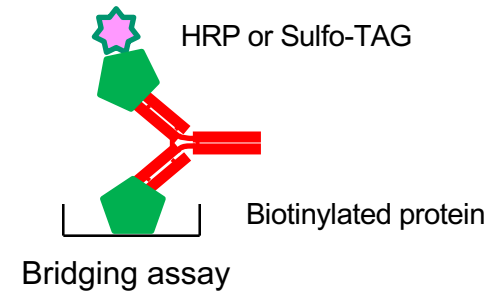
## Method development on MSD platform

*Fit for purpose* method development was needed

### 1. Set-up

- Coating: SARS-CoV-2 trimer protein
- Detection: SulfoTAG SARS-CoV-2 trimer protein

**No Signal**



➡ Coating and detection have been tested at different concentrations

# Case Study

## Direct assay on MSD platform

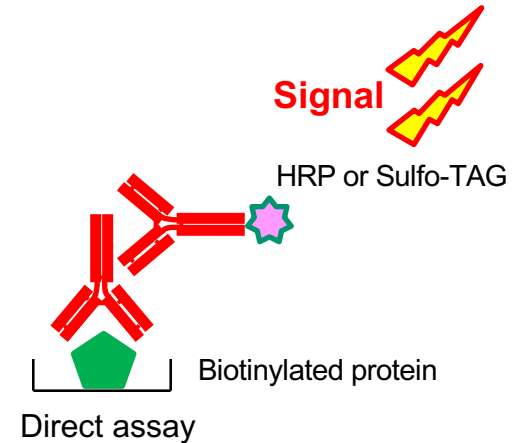
### 2. Set-up

Direct immunoassay on the MSD platform:

- Coating → SARS-CoV-2 (Omicron) S trimer Protein
- Detection → Goat anti Rat IgG HRP

Different conditions have been tested:

- Coating concentrations
- Different blockers
- Detection antibodies concentrations
- Increasing of washing steps
- Different MRDs



➡ High background and low S/N ratio



# Case Study

## Direct assay on MSD platform

- MSD standard plate
- Coating: SARS-CoV-2 (Omicron) S1+S2 trimer Protein (ECD, His Tag) in 1xPBS (from 2 different lot numbers)
- Blocking: 5% MSD Blocker A or 5% MSD Blocker A + 1% human serum
- MRD: 100 and 500
- Detection: SulfoTAG Goat anti-rat IgG (H+L)

Plate layout

ng/mL	1	2	3	4	5	6	7	8
<b>A</b>	0	5% MSD	0	5% MSD	0	5% MSD	0	5% MSD
<b>B</b>	100	Blocker A	100	Blocker A	100	Blocker A	100	Blocker A
<b>C</b>	500	+1%	500	+1%	500	+1%	500	+1%
<b>D</b>	25000	human	25000	human	25000	human	25000	human
<b>E</b>	0		0		0		0	
<b>F</b>	100	5% MSD	100	5% MSD	100	5% MSD	100	5% MSD
<b>G</b>	500	Blocker A	500	Blocker A	500	Blocker A	500	Blocker A
<b>H</b>	25000		25000		25000		25000	
MRD	100				500			

S/N ratio

	1	2	3	4	5	6	7	8
<b>A</b>	1.00	5% MSD	1.00	5% MSD	1.00	5% MSD	1.00	5% MSD
<b>B</b>	0.78	Blocker A	0.75	Blocker A	0.95	Blocker A	0.94	Blocker A
<b>C</b>	1.34	+1%	1.37	+1%	1.01	+1%	1.03	+1%
<b>D</b>	1.71	human	1.84	human	4.84	human	5.41	human
<b>E</b>	1.00		1.00		1.00		1.00	
<b>F</b>	0.66	5% MSD	0.58	5% MSD	0.96	5% MSD	0.95	5% MSD
<b>G</b>	1.51	Blocker A	1.45	Blocker A	1.11	Blocker A	1.10	Blocker A
<b>H</b>	2.31		2.49		5.04		5.57	
MRD	100				500			

# Case Study

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F	100	5% MSD	100	5% MSD	100	5% MSD	100	5% MSD
G	500	Blocker A	500	Blocker A	500	Blocker A	500	Blocker A
H	25000		25000		25000		25000	
MRD	100				500			

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MRD	100				500			



**5% MSD Blocker A  
MRD 500**

# Case Study

## Direct assay on MSD platform

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B	100	Blocker A	100	Blocker A	100	Blocker A	100	Blocker A
C	500	+ 1%	500	+ 1%	500	+ 1%	500	+ 1%
D	25000	human	25000	human	25000	human	25000	human
E	0		0		0		0	
F	100	5% MSD	100	5% MSD	100	5% MSD	100	5% MSD
G	500	Blocker A	500	Blocker A	500	Blocker A	500	Blocker A
H	25000		25000		25000		25000	
MRD	100				500			

S/N ratio

	1	2	3	4	5	6	7	8
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H	2.31		2.49		5.04		5.57	
MRD	100				500			



**Still high background due to the presence of cross-reacting antibodies in the individual serum, but after the vaccination the amount of antibodies will be much higher: not a typical ADA study !**

# Case Study

## Sample testing from the pre-tox study

- With the same method we analyzed 6 samples provided by the sponsor

	Mean ECLU signal
Pre-dose	3037
Pre-dose	2809
Pre-dose	2830
<b>22 days post-treatment</b>	<b>330701</b>
<b>22 days post-treatment</b>	<b>415478</b>
<b>22 days post-treatment</b>	<b>353094</b>



**Average fold increase: 127**

# Case Study

## 1-The Screening (SCP) and Confirmatory (CCP) assay cut point were determined in six batches

1. Analyze NC data, identify and exclude outliers
2. Analyse 25 individuals by 2 different analysts in 6 different runs
3. Normalize data as ration of response signal to noise. All further analysis is on Log (S/n) scale, if no transformation is needed, then S-N may be used
4. Use the box plot analysis to identify and exclude outliers and re-evaluate the distributions
5. Neither the signal nor the log-transformed data were normally distributed after Shapiro-Wilk analysis



**Nonparametric Method for SCP**  
95<sup>th</sup> percentile  
Alternatively, 90<sup>th</sup> percentile may be used, as it corresponds approximately to the parametric formula that assures 5% false positive error rate with 90% confidence (2).



CF=SCP-NC  
CF=3.68

# Case Study

## 1-The Screening (SCP) and Confirmatory (CCP) assay cut point were determined in six batches

- Proposed PCs (dilution curve prepared with surrogate positive control)

Analyte	PC1	PC2	PC3	PC4	PC5	PC6	PC7
	[ng/mL]	[ng/mL]	[ng/mL]	[ng/mL]	[ng/mL]	[ng/mL]	[ng/mL]
surrogate standard	1000	2000	4000	8000	16000	32000	64000

- Proposed preliminary QCs

Analyte	pLQC	pHQC
	[ng/mL]	[ng/mL]
surrogate standard	5000	50000



**The determination of the CCP was performed together with the determination of the SCP**  
**The same individual blank matrix samples used for the determination of the SCP, were incubated with excess of drug**

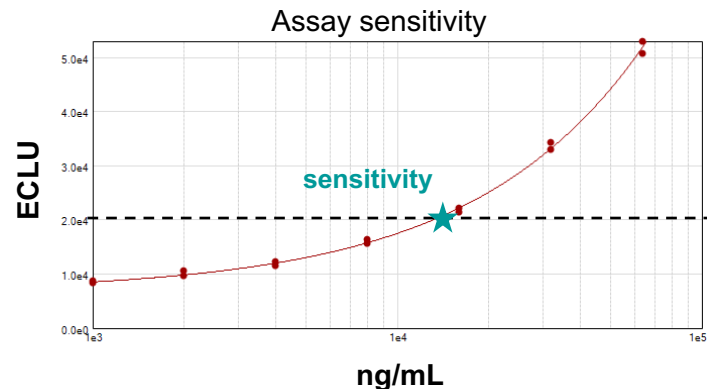
# Case Study

## 2-Sensitivity and precision of the screening assay

Sensitivity is determined interpolating the plate-specific SCP to the dilution curve

Assay sensitivity=19700 ng/mL

One QC level has been selected: **50000 ng/mL**



**This concentration has been used to determine additional parameters:  
drug tolerance, stability, sensitivity, HDH**

# Case Study

## Sample testing in the tox study - Confirmatory assay results

SBQ-no	Group	Gender	Animal	Day	Batch	CSS	CID	% ID	Result
S0061	3	male	31	44	4	274000	76800	72.0	POSITIVE
S0062	3	male	32	44	4	259000	48400	81.3	POSITIVE
S0063	3	male	33	44	4	268000	59100	77.9	POSITIVE
S0064	3	male	34	44	4	335000	112000	66.6	POSITIVE
S0065	3	male	35	44	4	149000	41600	72.1	POSITIVE
S0066	3	male	36	44	4	334000	89800	73.1	POSITIVE
S0067	3	male	37	44	4	255000	76100	70.2	POSITIVE
S0068	3	male	38	44	4	269000	55400	79.4	POSITIVE
S0069	3	male	39	44	4	286000	80300	71.9	POSITIVE
S0070	3	male	40	44	4	390000	147000	62.3	POSITIVE
S0071	3	female	76	44	5	175000	39900	77.2	POSITIVE
S0072	3	female	77	44	4	222000	51400	76.8	POSITIVE
S0073	3	female	78	44	4	276000	72900	73.6	POSITIVE
S0074	3	female	79	44	5	179000	25300	85.9	POSITIVE
S0075	3	female	80	44	4	325000	105000	67.7	POSITIVE
S0076	3	female	81	44	4	244000	84500	65.4	POSITIVE
S0077	3	female	82	44	4	166000	47400	71.4	POSITIVE
S0078	3	female	83	44	4	325000	96200	70.4	POSITIVE
S0079	3	female	84	44	4	286000	97300	66.0	POSITIVE
S0080	3	female	85	44	5	127000	22800	82.0	POSITIVE
S0081	3R	male	41	57	5	61800	7440	88.0	POSITIVE
S0082	3R	male	42	57	5	80600	7760	90.4	POSITIVE
S0083	3R	male	43	57	5	163000	18100	88.9	POSITIVE
S0084	3R	male	44	57	5	167000	19000	88.6	POSITIVE
S0085	3R	male	45	57	5	183000	17600	90.4	POSITIVE
S0086	3R	female	86	57	5	230000	36800	84.0	POSITIVE
S0087	3R	female	87	57	5	180000	27100	84.9	POSITIVE
S0088	3R	female	88	57	5	176000	30800	82.5	POSITIVE
S0089	3R	female	89	57	5	219000	29200	86.7	POSITIVE
S0090	3R	female	90	57	5	195000	30000	84.6	POSITIVE



**All treated animals have been confirmed positive**

Screening assay	ECLU signal (mean)
Pre-dose	608
44 days post-treatment	893600



**Average fold increase 1470**

CSS: confirmatory screening signal  
 CID: conf ID signal  
 %ID: % Immunodepletion



# Conclusions

- ➡ Covid-19 pandemic: high request for new methods able to detect the immune response
- ➡ Direct immunoassay was used
- ➡ Coating and detection have been tested at different concentrations
- ➡ Still high background due to the presence of cross-reactive antibodies in the individual serum however this has not impacted the suitability of the method since vaccinated animals showed very high signals
- ➡ After the analysis of the 6 cut point runs, the sensitivity of the assay (and as well the calculated LQC) was much higher than the initial planned, for this reason only a single QC level was used in 4 sets.

It's possible to use an ADA method for a vaccine efficacy study!

# Acknowledgment

LBA analysis group and all SBQ members

**Salvatore Calogero (Department head)**

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Julie Zurbuch

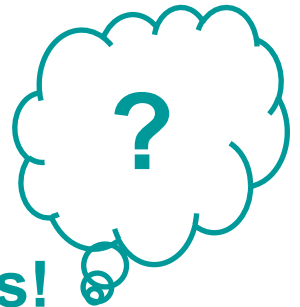
Katrin Leuenberger

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**Dr. Christoph Siethoff (CEO)**

**Mark Enzler (CSO)**



**Time for questions!**