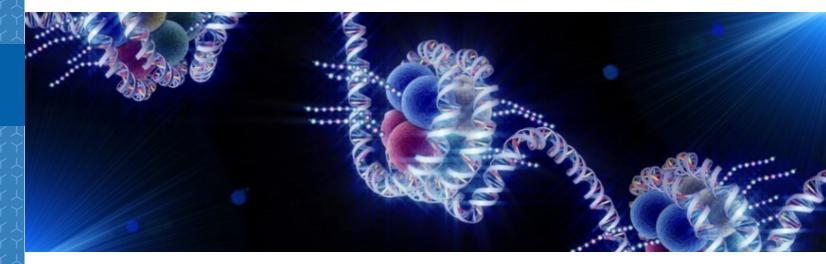
Novartis NIBR TM PK Sciences Bioanalytics



Flow cytometry based immunogenicity assays for CART

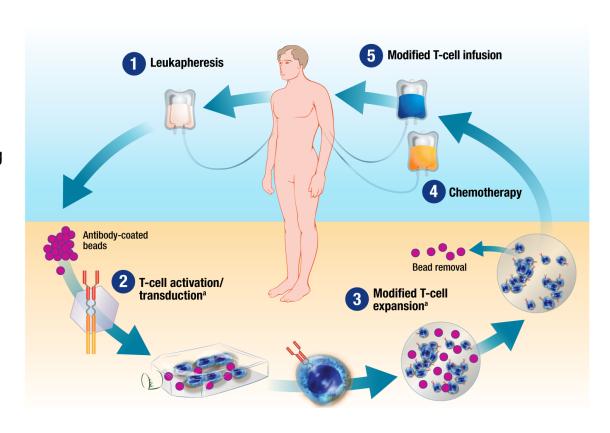
EBF Immunogenicity Training Day 23 - 24 March 2021

Christian Joffroy Novartis Institutes for BioMedical Research Translational Medicine PK Sciences-Bioanalytics



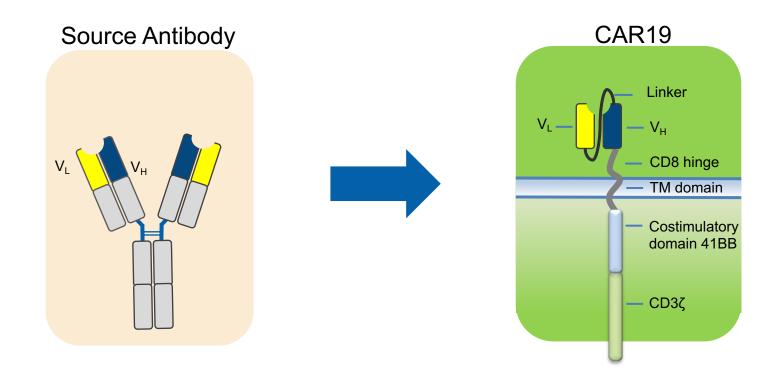
Chimeric antigen receptor T cells (CART)

- Leukapheresis: patient's T cells are harvested
- 2. T cells are activated on antibody-coated beads and genetically transduced ex vivo with a construct encoding the anti-CD19 CAR
- 3. CTL019 cells undergo ex vivo expansion on antibody-coated beads
- **4.** Chemotherapy: patient receives a preparative lymphodepleting regimen before T-cell infusion
- 5. CTL019 cells are re-infused into the patient, where they undergo in vivo expansion and target CD19+ cells for destruction





Composition of the CAR



The scFv (single chain variable fragment) targeting CD19 is based on a mouse hybridoma and has been characterized for its specificity to CD19 in several preclinical CAR T cell systems.



CAR-T immunogenicity

Humoral - ADA (antidrug antibody)

Potential risks of anti-CAR antibodies:

- Neutralization, i.e.
 functional inhibition of
 CAR binding to its target
- Induction of CAR-T cell death by CDC, ADCC, uncontrolled receptor triggering

Cellular immunogenicity

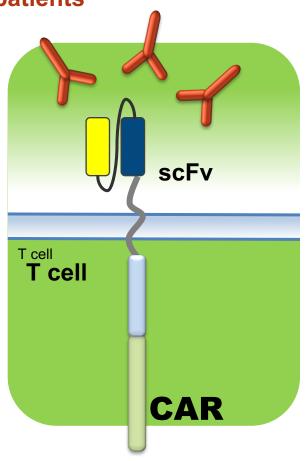
Potential risks:

- Activation of cytotoxic CD8 T cells by CAR proteins
- Elimination of CAR T cells



Humoral immunogenicity assay (ADA)

Anti-CAR antibodies (ADA) in serum samples from patients



anti-CAR ADA assay formats

- Cell-based
 - Capture ADA with CARexpressing T-cell line
 - Mimics membrane CAR on CAR-T cell
 - Flow cytometry output
- LBA (Ligand Binding Assay)
 - Soluble CAR or CAR fragments (can be difficult)
 - Parent mAb used to derive CAR-scFv
 - No membrane context
 - ELISA type assay

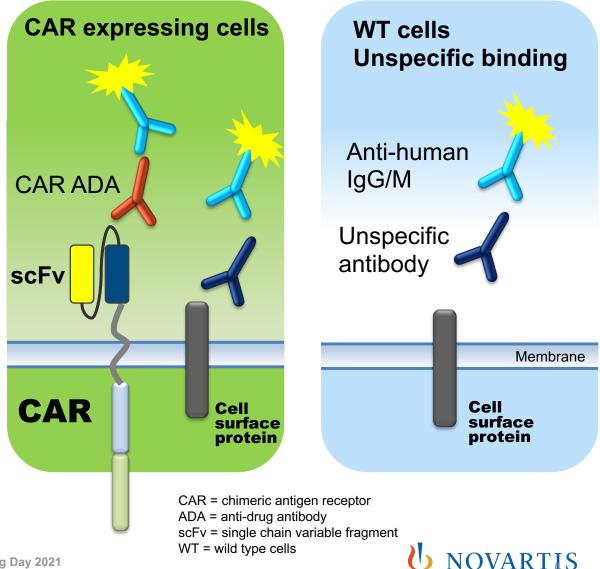


Cell based humoral IG assay

Jurkat cell lines (CAR+ vs. wild type) are used to detect ADAs in clinical samples

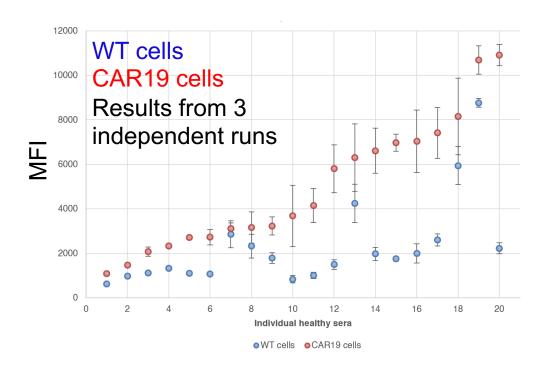
Calculation of CART ADA signal: Signal on CAR+ cells minus signal on WT cells (background)

Comparable cell numbers?
Staining of Notch3 receptor



Pre-existing ADA & Cut point

Pre-existing antibodies in >80% of individual sera (validation) and patients (clinical studies)



Potthoff B et al- (2020) "A cell-based immunogenicity assay to detect antibodies against chimeric antigen receptor expressed by tisagenlecleucel,

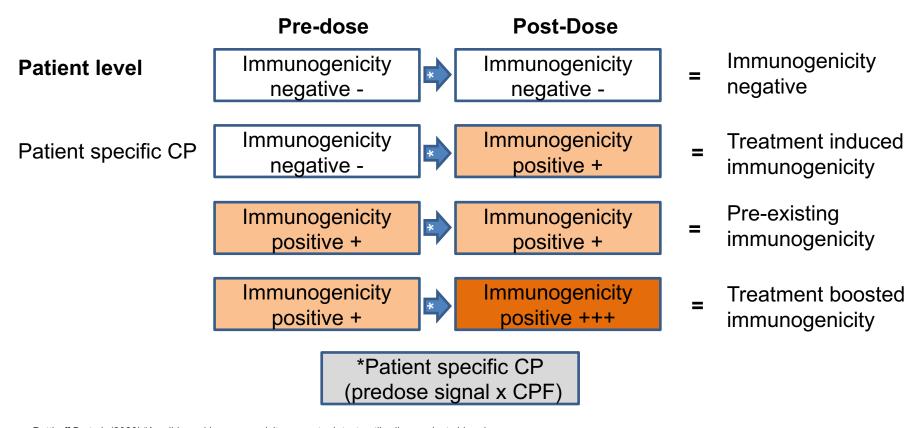
JIM, Vol. 476,112692, https://doi.org/10.1016/j.jim.2019.112692

- high signals of healthy donor sera didn`t allow for meaningful cut-point evaluation and outlier removal
- CAR19 specific signals (inhibition with CAR19 protein)

=> Immunoglobulin depleted sera were used for CP evaluation



Data interpretation - Patient



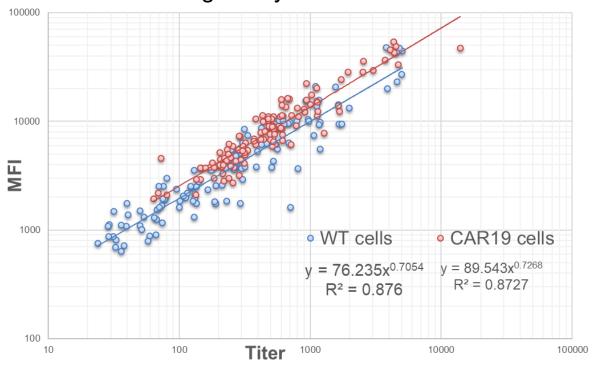
Potthoff B et al- (2020) "A cell-based immunogenicity assay to detect antibodies against chimeric antigen receptor expressed by tisagenlecleucel, JIM, Vol. 476,112692, https://doi.org/10.1016/j.jim.2019.112692



Correlation of screening and titration results

- Good correlation between screening (MFI) and titer results shown during validation and clinical sample analysis
- Possible since screening signal is not "capped" but correlates well with the analyte even at higher titers
- Screening step is reported since it does not only indicate whether samples are positive or negative but already reflect the different amounts of ADA

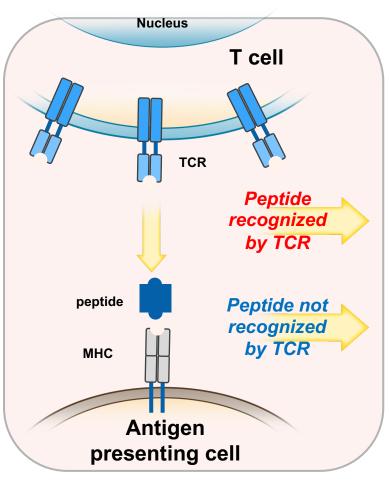




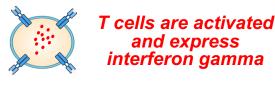
Potthoff B et al- (2020) "A cell-based immunogenicity assay to detect antibodies against chimeric antigen receptor expressed by tisagenlecleucel, JIM, Vol. 476,112692, https://doi.org/10.1016/j.jim.2019.112692



Cellular immunogenicity - T cell activation assay



- Patient`s PBMCs are collected and incubated with CAR peptides
- T cell activation is measured by Flow cytometry (intracellular IFNg staining)



T cell



No activation



Cellular immunogenicity assay

Controls:

- **SEB** (staphylococcal enterotoxin B)
- **CEF**: pool of 27 peptides corresponding to viral and vaccine sequences frequently recognized by CD4+ and CD8+ T lymphocytes
- **FMO:** stimulation with SEB and incubating with all fluorescently labeled antibodies except IFNγ
- **DMSO** (Negative control, used to dissolve peptides)

Actual stimulation:

- 2 CAR peptide pools (15-mer overlapping peptides)

Read out: percentage of IFNγ+CD4+ T cells and IFNγ+CD8+ T cells with respect to their parent populations. No threshold/cut point is applied. Percentages of IFNg positive cells are reported.



Cellular immunogenicity assay

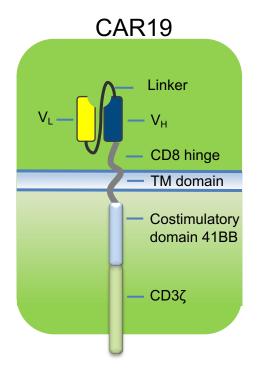
Ranges observed in exemplary clinical study

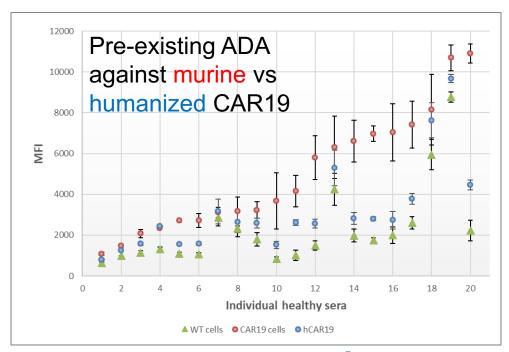
Population	Treatment	Values
IFNγ+ CD4 T cells	SEB treated	1.610-27.300%
	CEF treated	<lod-0.093%< td=""></lod-0.093%<>
	Pool 1 treated	<lod-0.091%< td=""></lod-0.091%<>
	Pool 2 treated	LOD-0.081%
IFNγ+ CD8 T cells	SEB treated	2.710-25.600%
	CEF treated	0.076-2.450%
	Pool 1 treated	<lod< td=""></lod<>
	Pool 2 treated	<lod-0.051%< td=""></lod-0.051%<>



Outlook – further characterization of humoral immunogenicity against CAR19

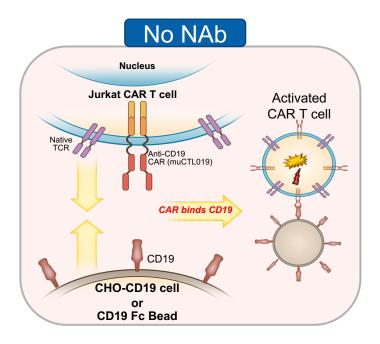
- No impact of detected ADAs on efficacy, safety or exposure
- Identification of the predominant CAR19 ADA binding sites
- Spiking of extracellular domains (murine and humanized version of CAR19) and peptides



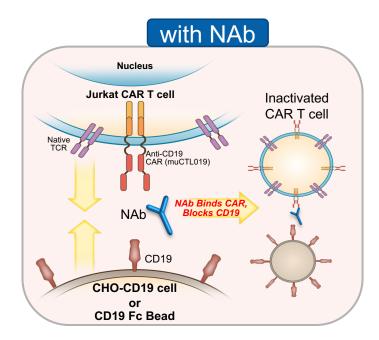




Outlook – potential cell-based CAR T NAb assay



- Jurkat-muCTL019 binds CD19 (CHO or Beads)
- Jurkat activated → produces measurable response (Luciferase or IFNy)



- NAbs present in sample, bind CAR and block CAR-CD19 interaction
- Jurkat remains inactivated → limits measurable response (Luciferase or IFNy)



Thank you



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