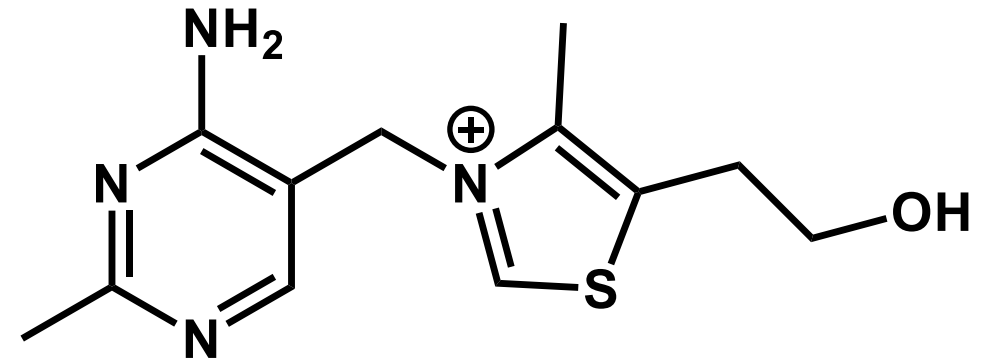


THE SIMULTANEOUS QUANTIFICATION OF THIAMINE, ITS PHOSPHATE DERIVATIVES AND PRECURSORS IN *ARABIDOPSIS*

Jana Verstraete, Simon Strobbe, Dominique Van Der Straeten and Christophe Stove

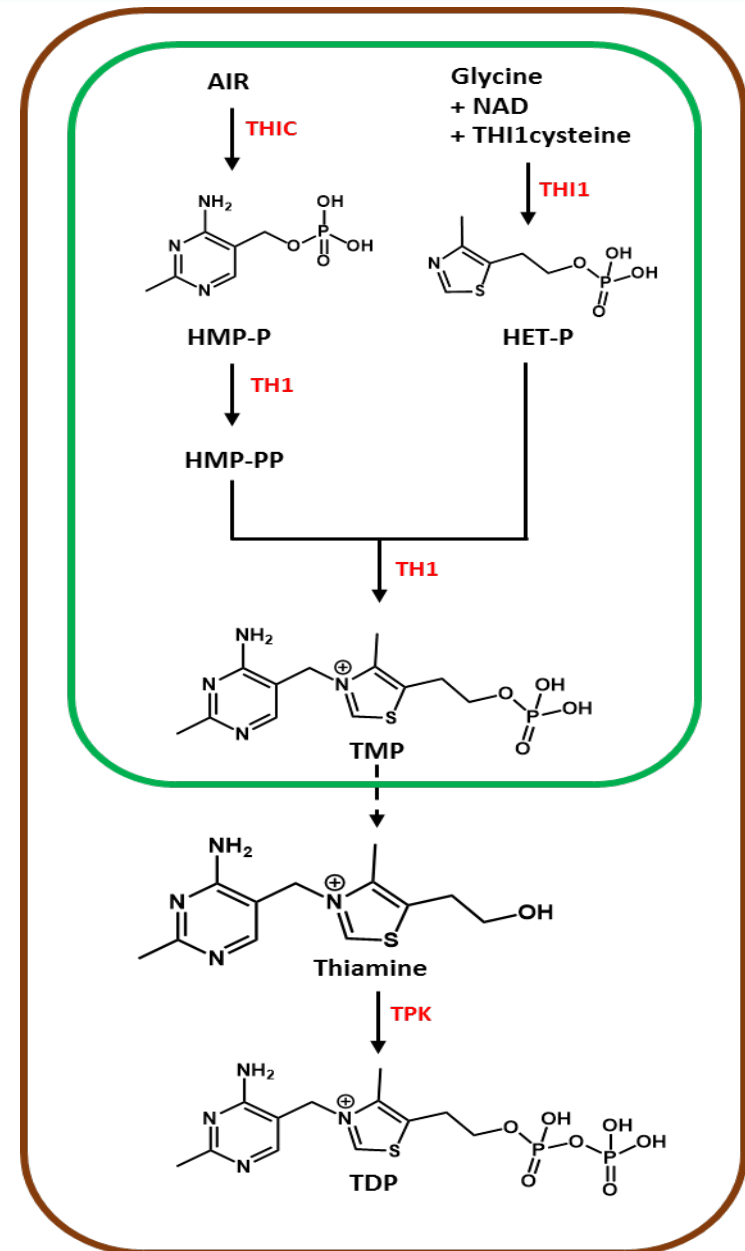
INTRODUCTION

- Vitamin B1
- Energy metabolism
- Deficiency > Beri-Beri
- Low content in major food crops, e.g. rice
- Mainly problem in developing countries



INTRODUCTION

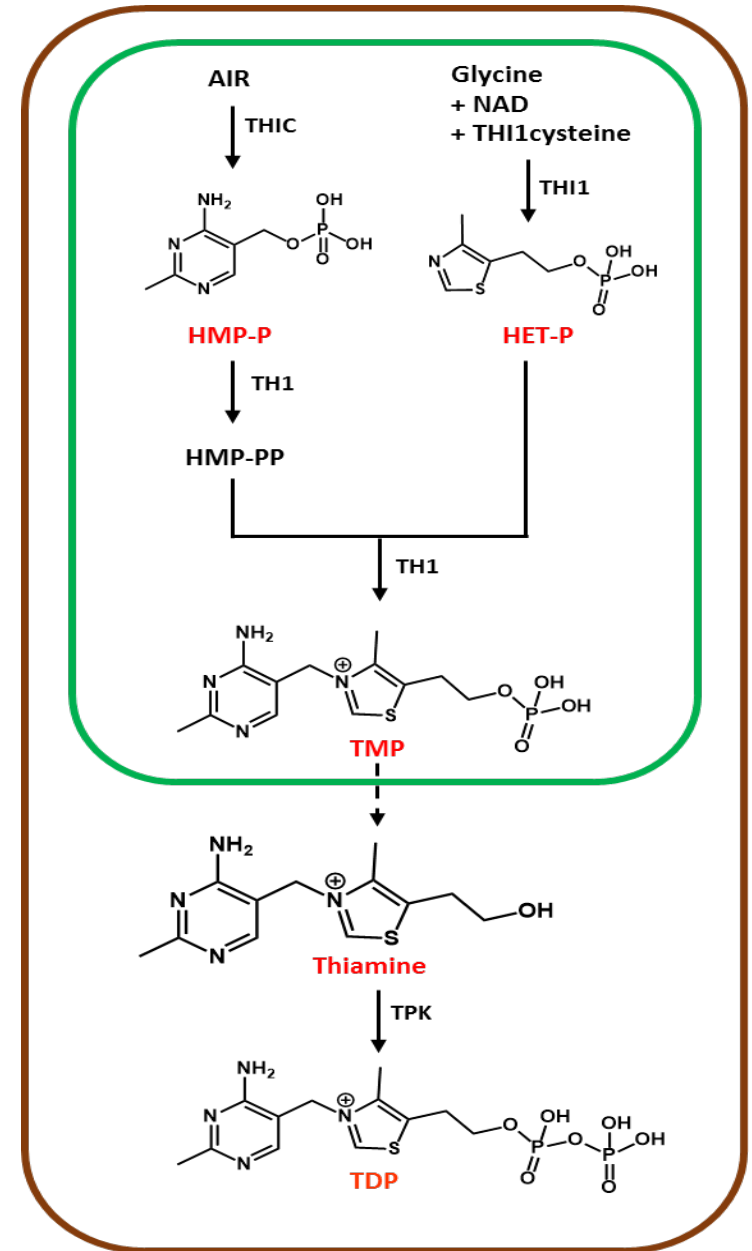
- ↑ thiamine levels via genetic modification of **key enzymes** in thiamine bio-synthesis



INTRODUCTION

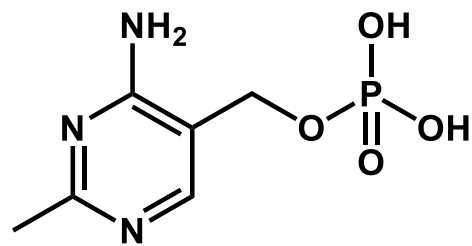
HOW?

- Need for knowledge about regulation of thiamine bio-synthesis in plants
- determination direct **products** of key enzymes
- *Arabidopsis thaliana* = reference plant

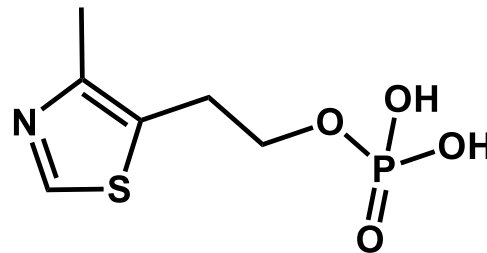


AIM OF THE STUDY

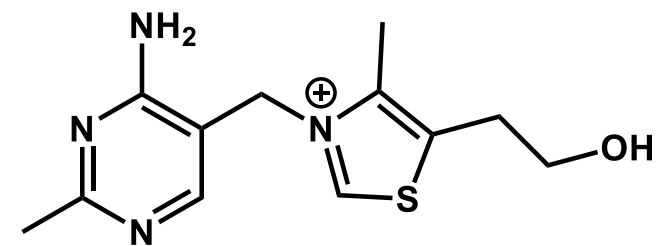
DEVELOPMENT OF LC-MS/MS METHOD FOR THE DETERMINATION OF THIAMINE, ITS PHOSPHATE DERIVATIVES AND PRECURSORS IN *ARABIDOPSIS THALIANA*



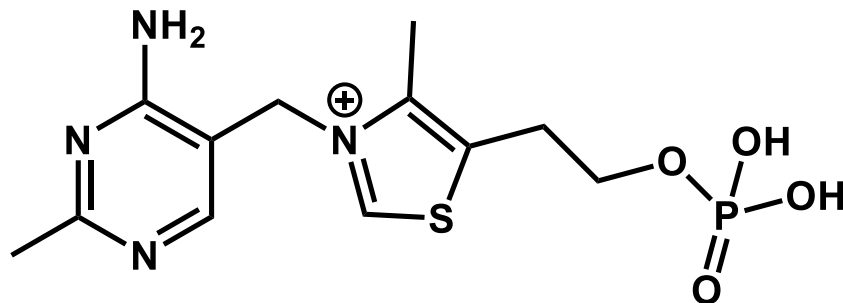
HMP-P



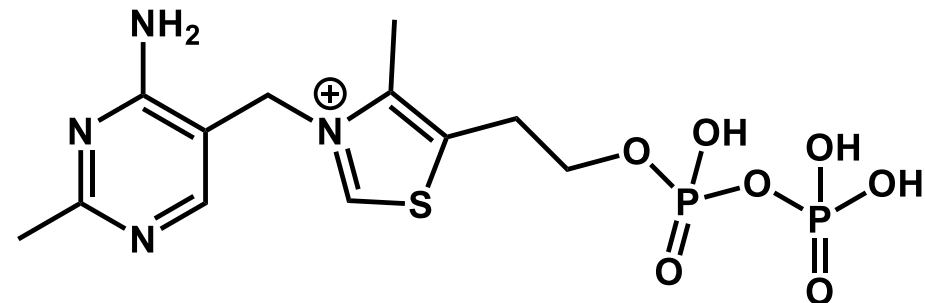
HET-P



Thiamine



Thiamine monophosphate (TMP)



Thiamine diphosphate (TDP)

CHALLENGES

- Polar compounds
- Ionogenic compounds
- Chelating compounds



Chromatography

- Labile compounds



Sample preparation

- Endogenous compounds



Validation

- No reference standards of HMP-P and HET-P
- No labeled internal standards for all compounds



Quantification

OPTIMISATION OF THE CHROMATOGRAPHY

Phenomenex Gemini NX C18

Mobile phase A: 10 mM NH_4CO_3 pH 8.8

Mobile phase B: MeOH

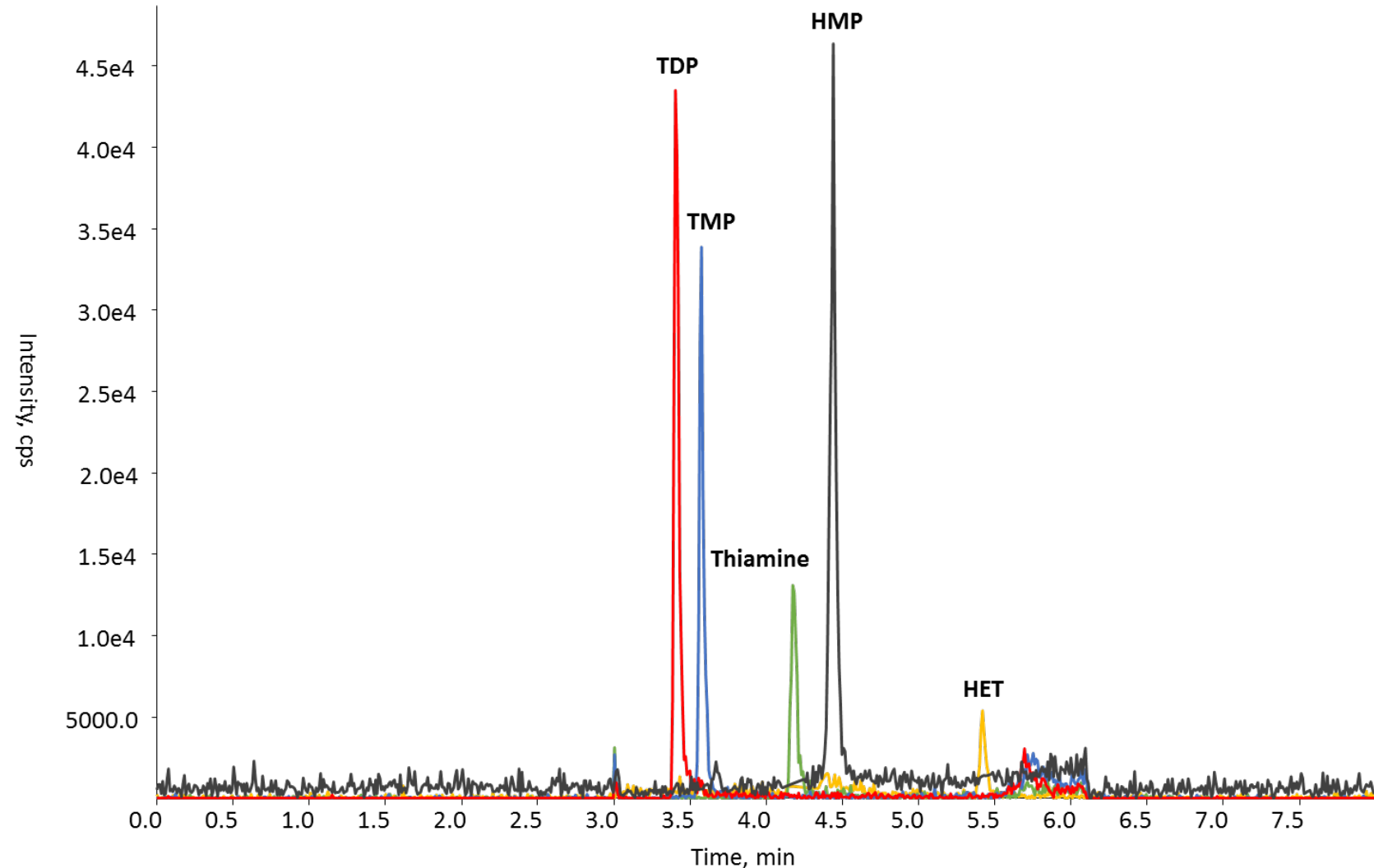
8-min gradient run

LLOQ level:

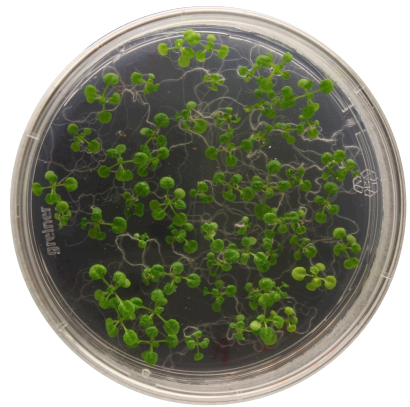
TDP, 7 ng/ml; TMP, 0.9 ng/ml;

thiamine, 3.6 $\mu\text{g/ml}$; HMP, 0.17 ng/ml;

HET, 0.09 ng/ml



OPTIMISATION OF THE SAMPLE PREPARATION PROCEDURE



200 mg

+ 1.5 ml 0.1 M HCl



30' at 74°C



10' at 30 Hz



+ 10U phosphatase



Total thiamine: thiamine + TDP + TMP

Total HET: HET + HET-P

Total HMP: HMP + HMP-P + HMP-PP



24h at 45°C

- phosphatase



TDP, TMP, thiamine, HET and HMP



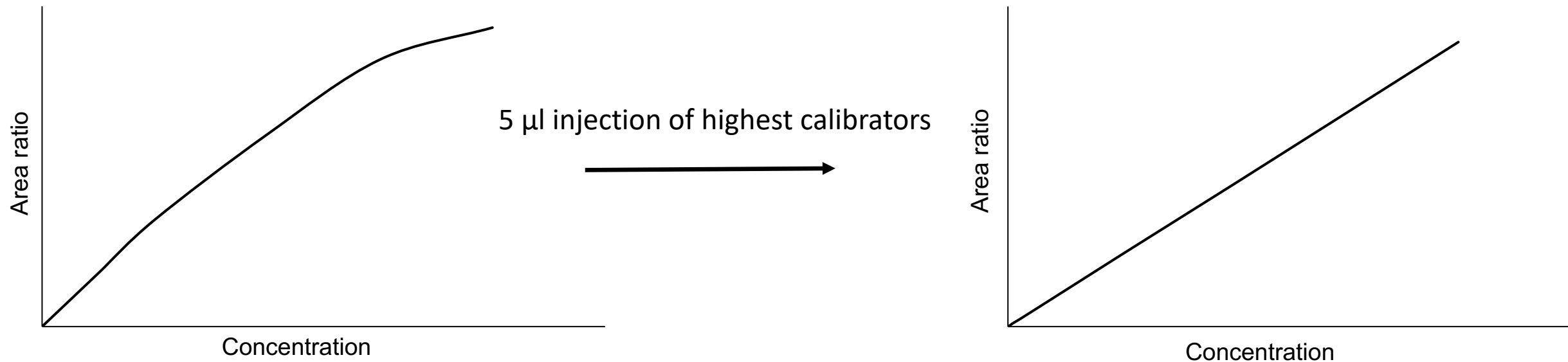
24h at 4°C

Parameters to evaluate:

- Selectivity
- Carry-over
- Calibration model
- Accuracy and precision
- Dilution integrity
- Matrix effect
- Recovery
- Stability

VALIDATION: CALIBRATION MODEL

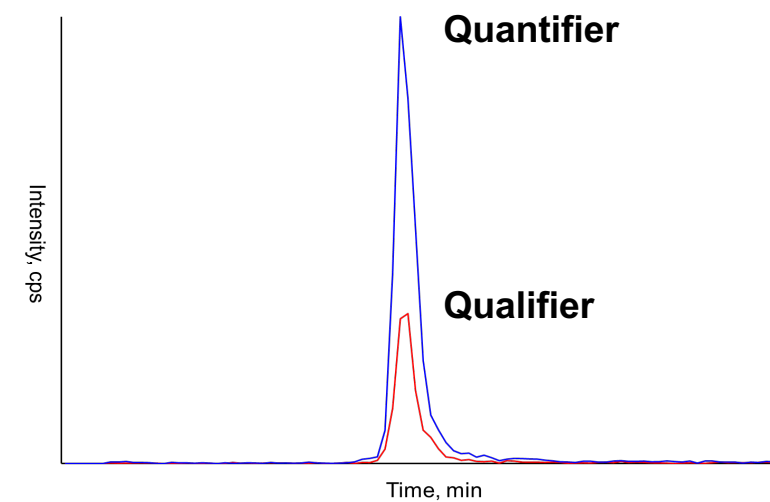
- Absence blank matrix → charcoal-treatment
- Analyte levels differed substantially between phosphatase-treated and non-treated samples
→ need for a very broad calibration range



Detector saturation

VALIDATION: SELECTIVITY

- Quantifier/Qualifier ratio
- Ion ratio's were all within the tolerated window of ion ratio neat standards solutions



		HET	HMP	Thiamine	TMP	TDP
Neats	mean	0.20	0.19	1.13	0.27	0.38
	tolerated window	[0.15-0.25]	[0.14-0.24]	[1.03-1.23]	[0.22-0.32]	[0.3-0.46]
	CV (%)	14%	10%	8%	12%	9%
Charcoal treated Arabidopsis	mean	0.20	0.19	1.14	0.26	0.37
	CV (%)	9%	10%	9%	12%	7%
Spiked Arabidopsis	mean	0.20	0.19	1.15	0.26	0.38
	CV (%)	19%	8%	8%	10%	8%
Arabidopsis	mean	0.25	0.18	1.14	0.26	0.37
	CV (%)	32%	9%	9%	9%	6%

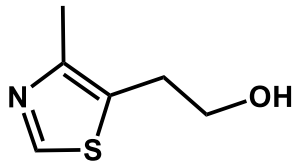
VALIDATION: MATRIX EFFECT (ME)

- Based on Matuszewski *et al.* (2003)
- Absolute ME = $\text{analyte signal matrix} / \text{analyte signal neat solvent}$
→ correct for endogenous signal matrix
- Relative ME = $\text{ME analyte} / \text{ME internal standard}$

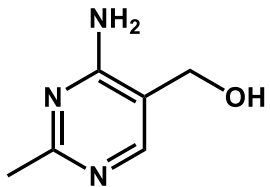
We demonstrated that:

- The applied internal standard compensated for matrix effect

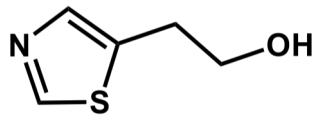
VALIDATION: MATRIX EFFECT (ME)



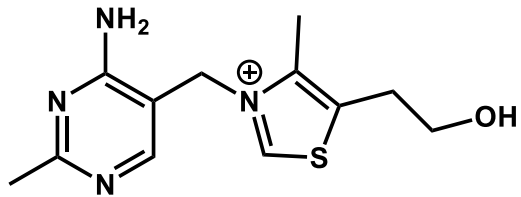
HET



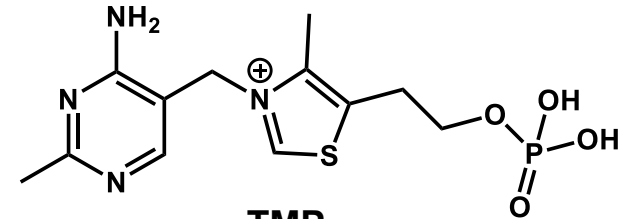
HMP



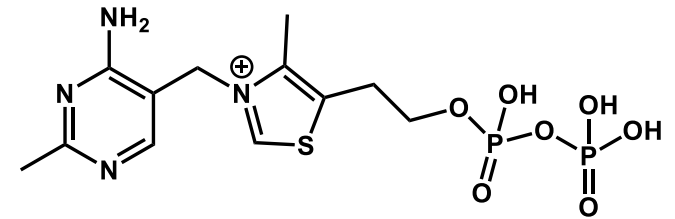
HET analogue



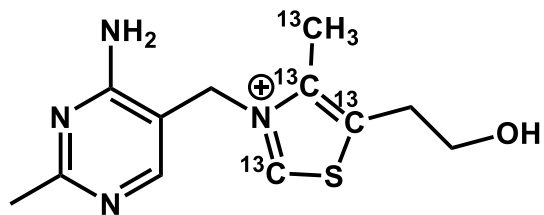
Thiamine



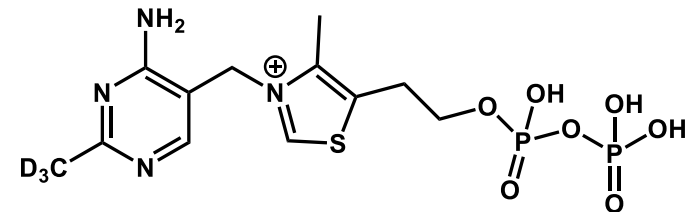
TMP



TDP



^{13}C Thiamine



D_3 TDP

VALIDATION: MATRIX EFFECT (ME)

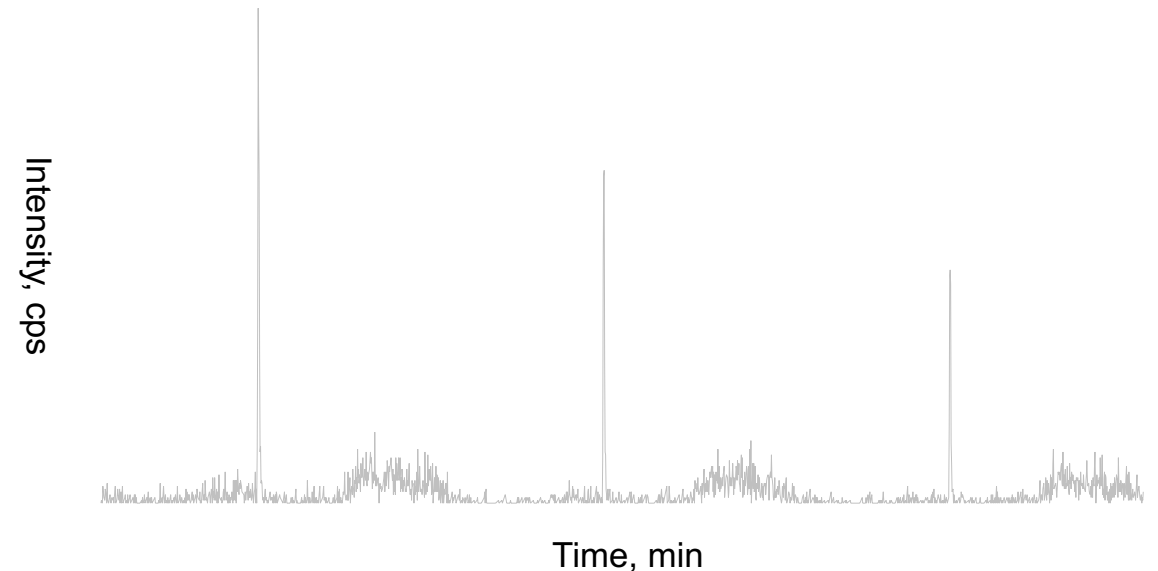
- Based on Matuszewski *et al.* (2003)
- Absolute ME = analyte signal matrix / analyte signal neat solvent
→ correct for endogenous signal matrix
- Relative ME= ME analyte/ ME internal standard

We demonstrated that:

- The applied internal standards compensated for matrix effect
- The relative matrix effect in charcoal-treated matrix was equal to the relative matrix effect in *Arabidopsis* matrix

VALIDATION: CARRY-OVER

- Relevant carry-over for TMP and TDP
- Could not be improved by optimising needle wash
- Even without intervention of needle, carry-over is still an issue
- Specific measures to minimize carry-over should be taken



VALIDATION

Parameters to evaluate:

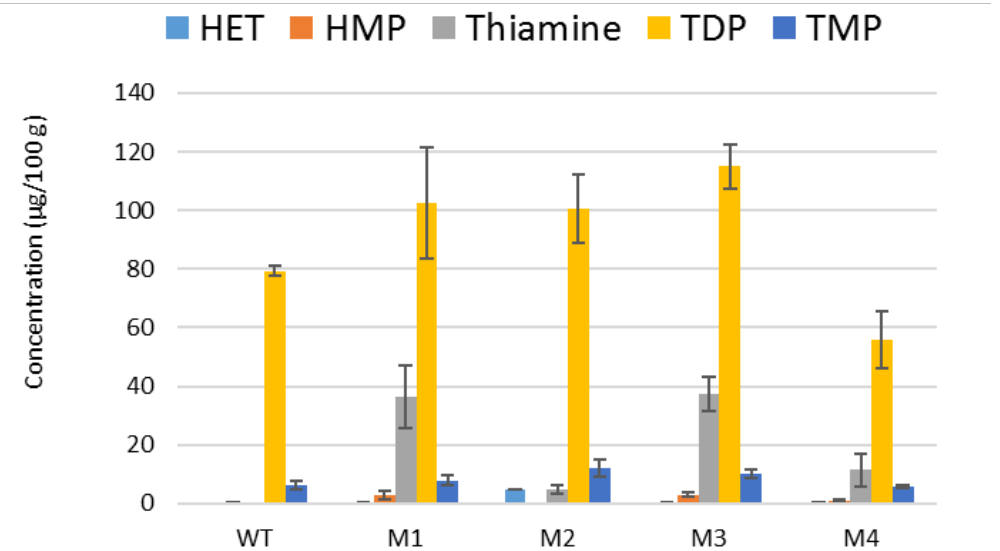
- Selectivity
- Carry-over
- Calibration model
- Accuracy and precision
- Dilution integrity
- Matrix effect
- Recovery
- Stability



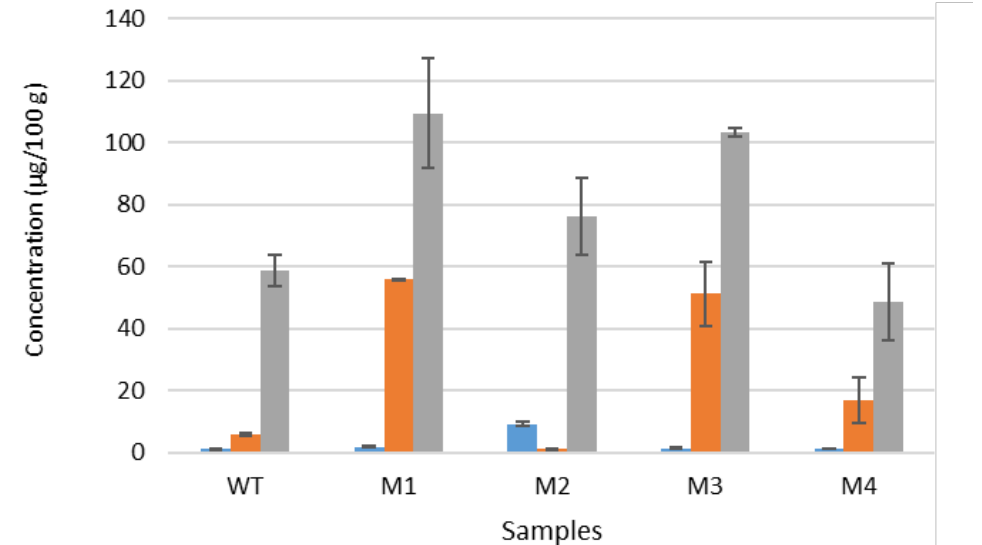
WT = non-genetically modified = 'control'

M = genetically modified lines

Non-phosphatase treated



Phosphatase treated



CONCLUSION

- Development of LC-MS/MS method for the quantification of thiamine, its precursors and phosphate derivatives in *Arabidopsis thaliana*
- Successfully tackled several challenges: high polarity analytes, lability analytes, endogenous presence analytes...
- Successful validation of the method based on international guidelines
- The method was successfully applied on genetically modified *Arabidopsis* lines

Jana Verstraete - Prof. dr. Christophe Stove
Simon Strobbe– Prof. dr. Dominique Van Der Straeten

Faculty of Pharmaceutical Sciences
Department of Bioanalysis
Laboratory of Toxicology

E jrverstr.Verstraete@ugent.be / Christophe.Stove@ugent.be
T +32 9 264 81 36

www.ugent.be

 Ghent University
 @ugent / @christophestove
 Ghent University