



# **WS 4: Supporting Cell & Gene Therapies in the Bioanalytical Laboratory**

**12<sup>th</sup> EBF Open Symposium**  
**Imagine! A new bioanalytical Earthrise**

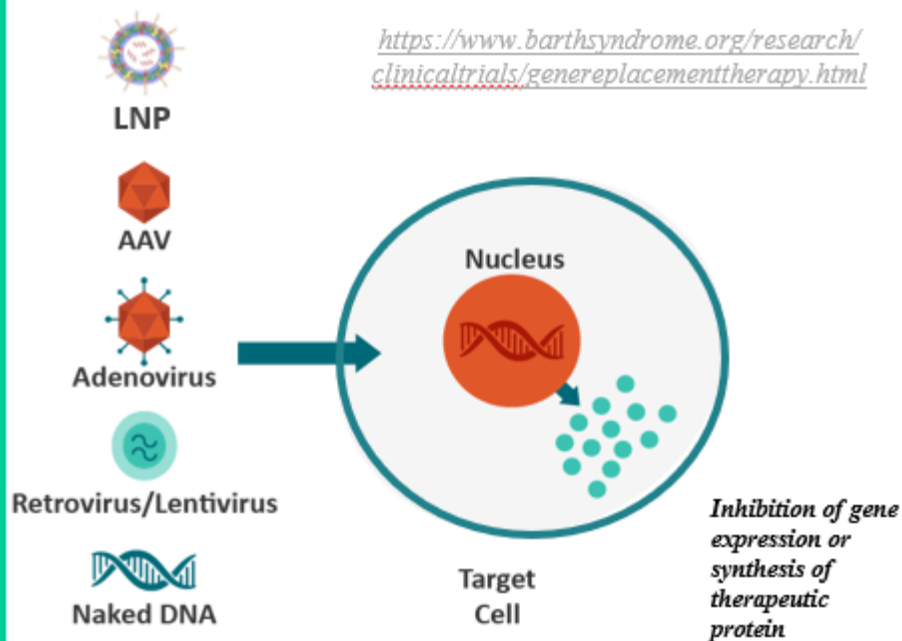
# Aim of session and discussion points

- Topic Introduction
- Survey results
- Immune response to cell and gene therapy
- Gene therapy – Exposure and integration
- Gene therapy – Transgene product assessment
- Cell Therapies – Pre-clinical development and animal models
- Cell therapies – Exposure assessment

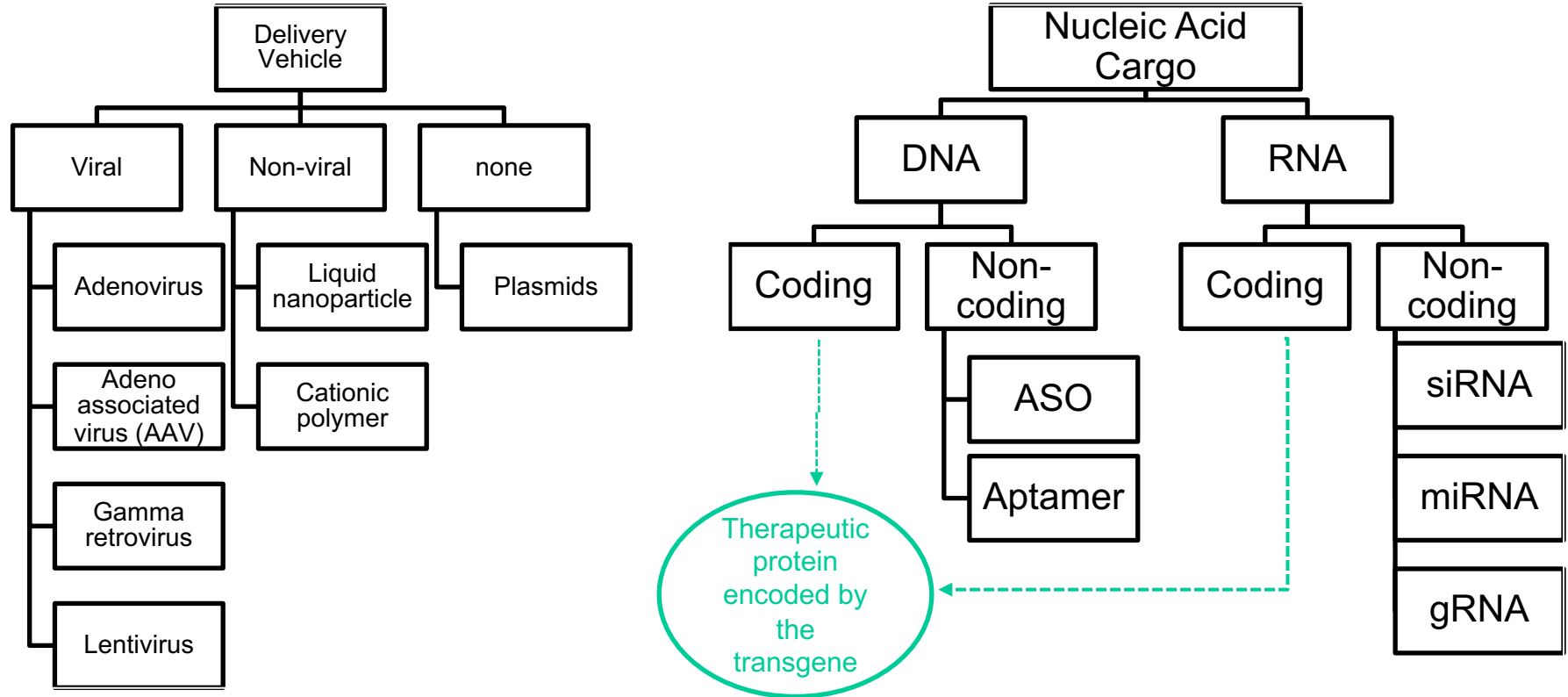
## Gene Therapies

All strategies that modify an individual's protein make-up by introducing exogenous nucleic acid or nucleic acid modifiers, regardless of delivery\*

Defined by the precision of the procedure and the intention of direct therapeutic effect.



# GT components define the Bioanalytical Support

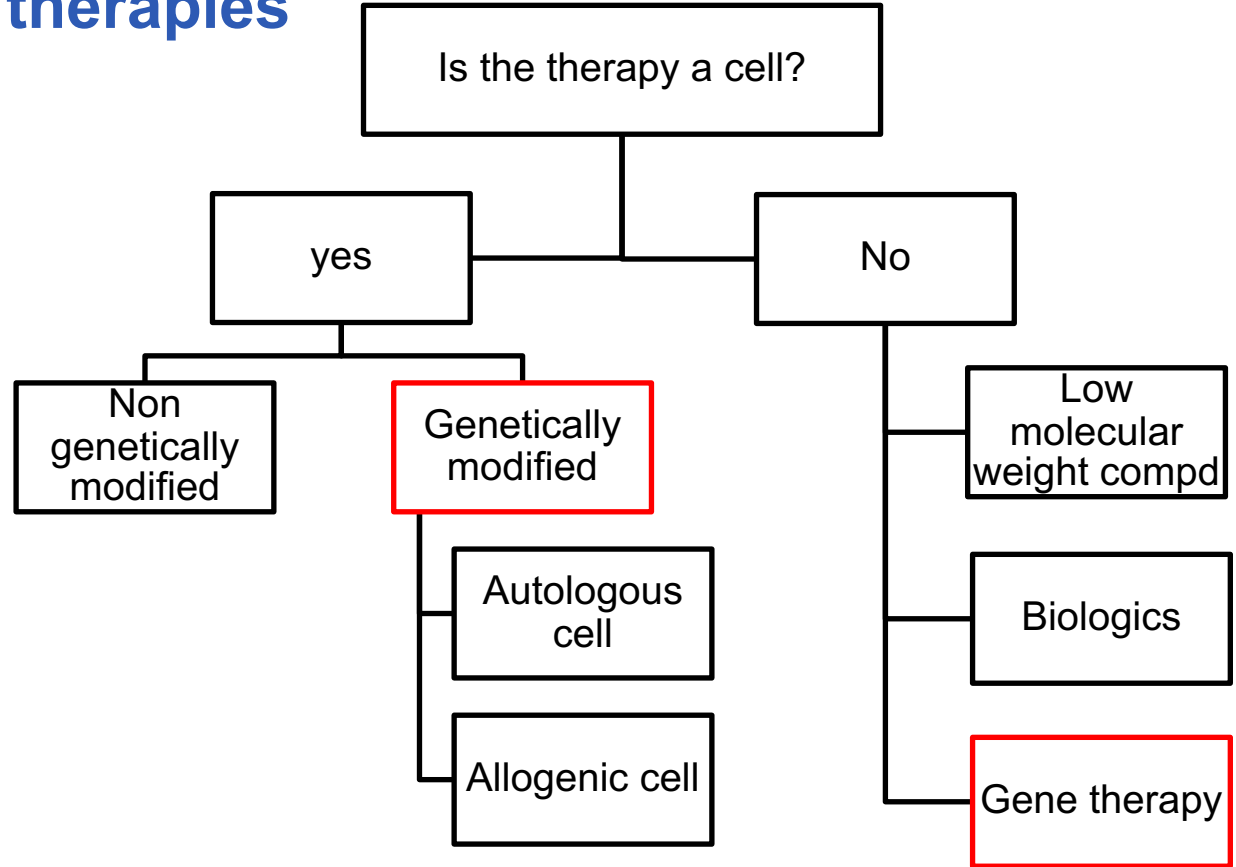


# Cell “vs” gene therapies

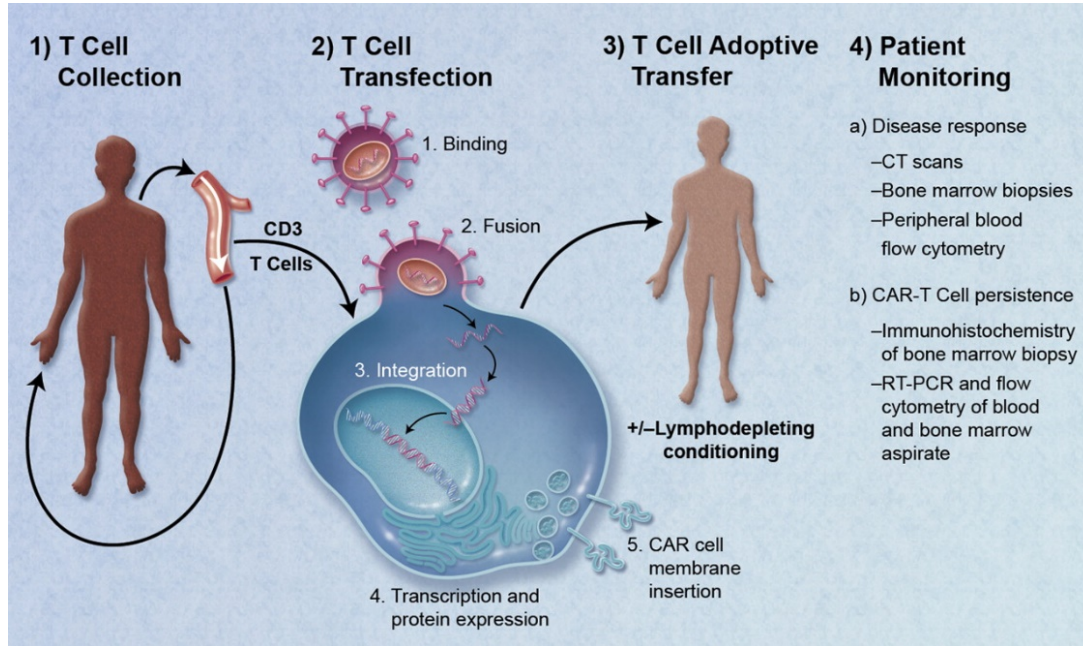
## Ex-vivo gene therapies

Beyond correcting genetic deficiencies directly in the patients' cells, gene therapy can also provide a cell with capabilities not present in its natural state (e.g. CAR-T therapeutics).

=> Not all cell therapies are gene therapies



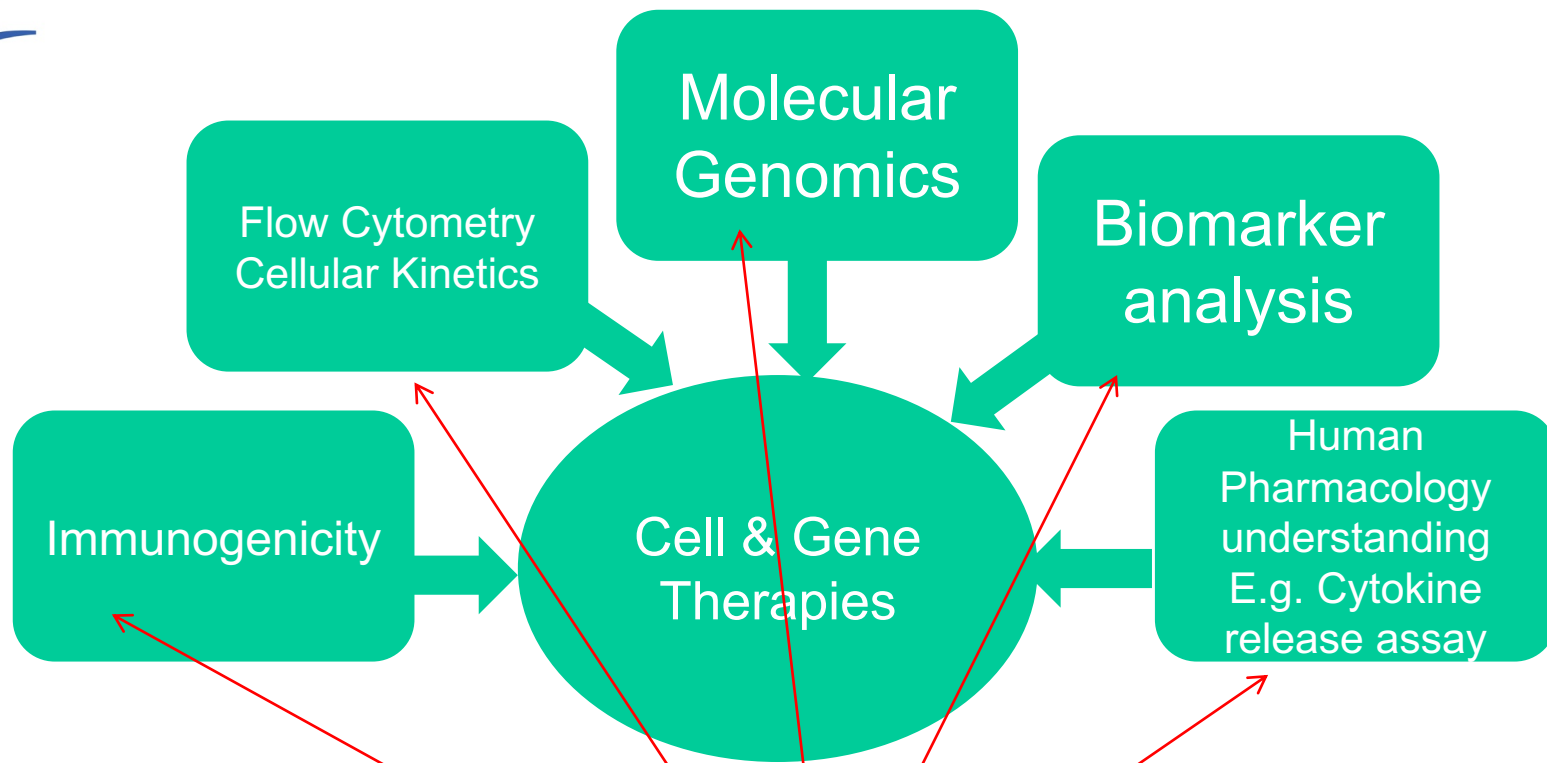
# EBF Autologous T Cell Therapies: an example of cell therapy



## ➤ CAR-T: current paradigm of cell therapies

- Strimvelis: autologous *ex-vivo* gene therapy: CD34+ enriched stem cells transduced with gamma retrovirus carrying ADA gene

## ➤ Allogenic (“off-the-shelf”) cell therapies: cells come from donors => impact on the immunogenicity assessment



BioA Activities?

## Survey results

- Companies currently working on C&GT:
  - Pharma (7) 50%
  - CRO (7) 50%
  
- Companies not currently in C&GT
  - 28 (of 67)
  
- Companies who said they will work on it in the future:
  - 3

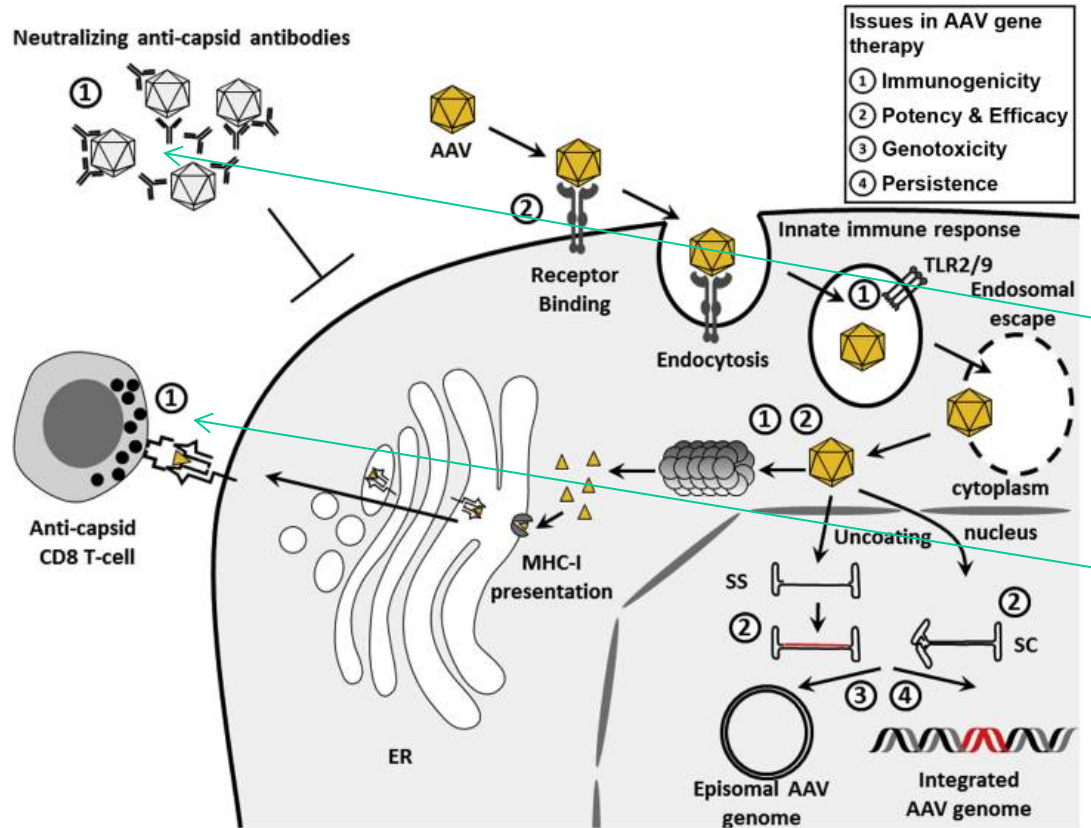


What platforms do you use?	Replies (n)	Replies (% of companies)
PCR	11	79%
Flow cytometry	11	79%
LBA	13	93%
ELISPOT	9	64%
Cell Counters	9	64%
Mass spectrometry	10	71%
Enzymatic assays	11	79%
Clinical analysers	7	50%
other:	Multiplex cytokine analysis, and biomarkers in general Cell-based functional assay (potency and nAb) Immunohistochemistry, Imaging (PET/CT), RNASeq/Nanostring, ex vivo functional assay for viral replication (cell bioassays)	

## Which endpoints do you support?

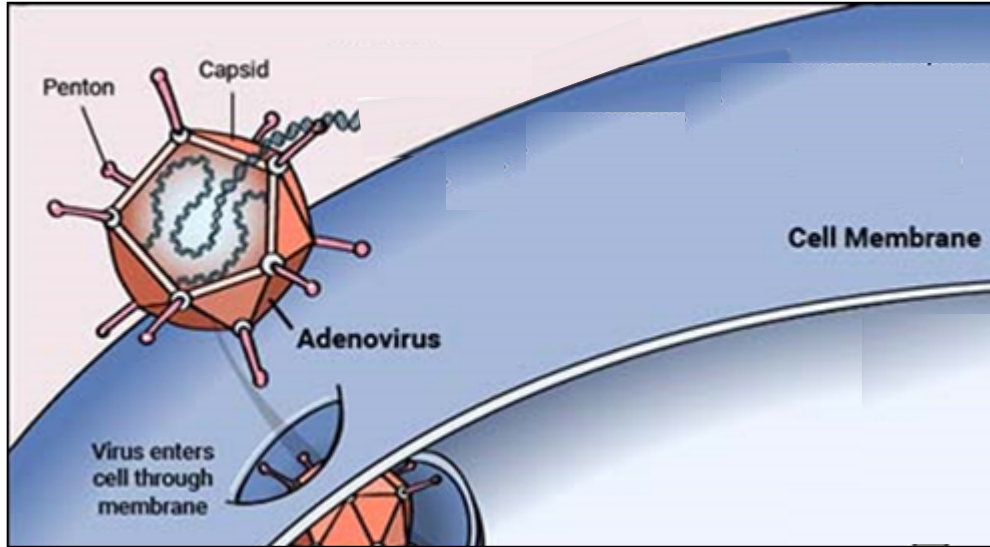
Patient selection/stratification	6	43%
Primary or secondary endpoint	13	93%
Exploratory endpoint	13	93%
PK endpoint	11	79%
PD endpoint	13	93%
Immunogenicity endpoint	13	93%
Bio-distribution assessment	8	57%
IND submission package	10	71%
BLA/MAA submission package	9	64%

# Gene Therapy – Capsid immunogenicity



**Pre-existing response:**  
ADA & Neutralising Ab assay

**Cell mediated response:**  
ELISPOT and/or Flow assay

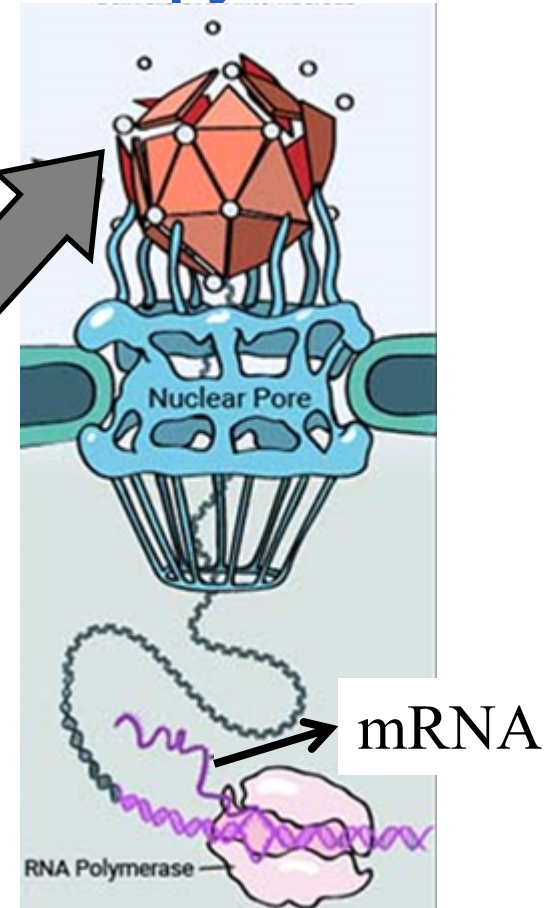
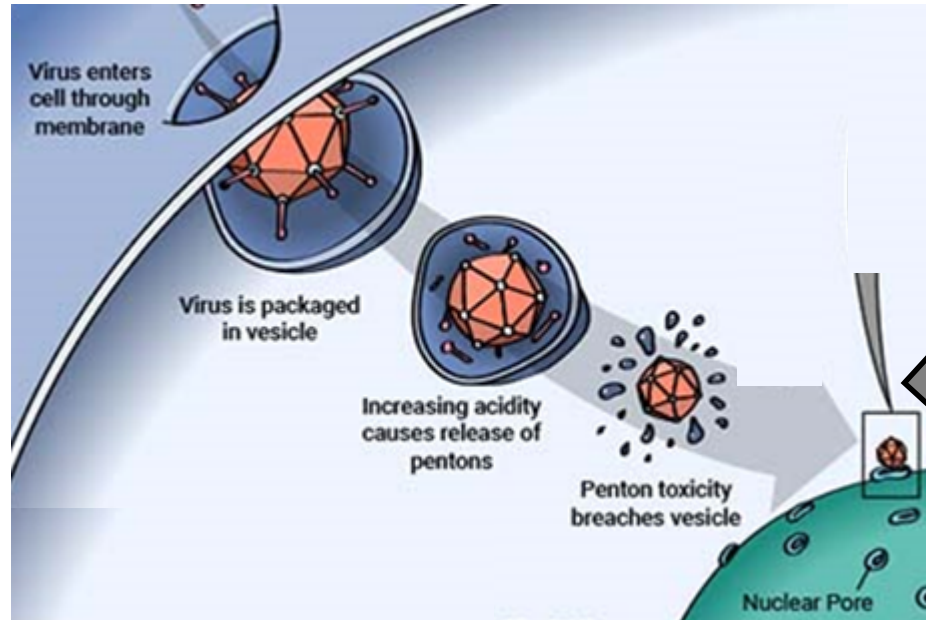


## BioA Challenges:

As a community where does Bioanalysis contribute to this Class of therapeutic?

Q: Which lab should do this work and what regulation should we follow (FDA, ISO, CLIA). What are the specific considerations for BioA ?

# EBF Exposure and integration Gene therapy



## Exposure and integration – Gene therapy

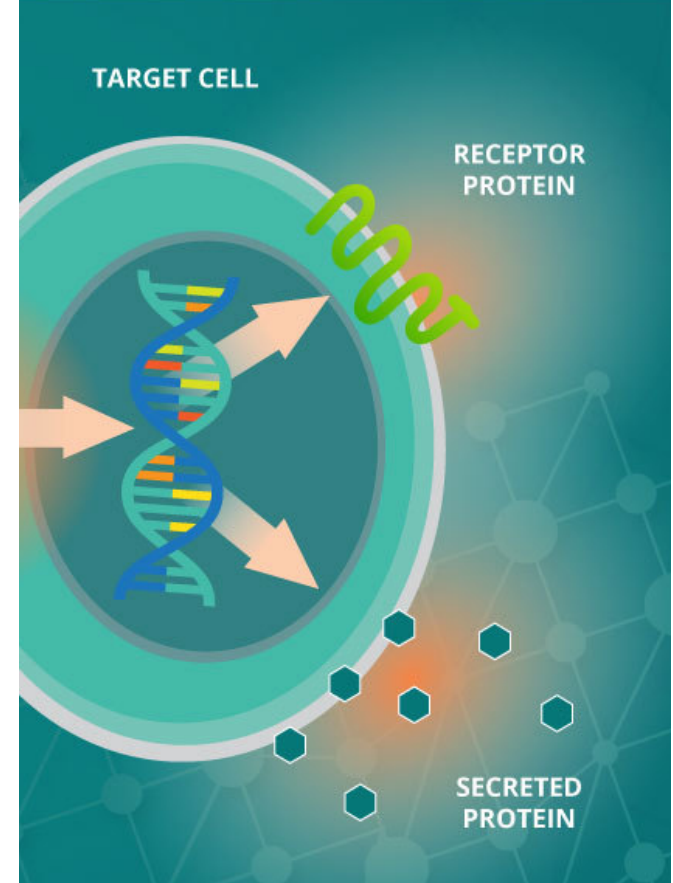
- qPCR / RT-PCR
- Hybridisation ELISA (ASO, siRNA)
- Branched ELISA
- Oligonucleotide by MS
- Viral Capsid detection by MS
- Lipid particle by MS
  
- Pre-clin biodistribution – immunohistochemistry, in-situ hybridisation
- Shedding

Q: How can we bring this under the BioA umbrella?

Q: Historic settings of the assays no appropriate for current needs  
(example hybridisation assays)

## Transgene product

- Cell surface receptor
  - Soluble protein or excreted protein
  - Functional protein
  - Functional enzyme
  
  - Transgene product Immunogenicity
    - transcription factor immunogenicity in clinical
- Q: Who has seen this?
- Native protein
  - Engineer protein



Q: Is it needed to do transgene product immunogenicity?

Q: How can we estimate the risk (safety, long lasting effects)?



## Cell Therapies

CAR-T is used as case modality



## Pre-clinical development and animal models

- Considerations for Immunogenicity assessment
- Building better models
- Cytokine release syndrome

Questions and discussion:

- Value of pre-clin assessments?
- Translational ability of results? (cell and gene therapy alike)

## EMA/FDA – Immunogenicity Guidance

- Predictive nature is low for clinical risk
- Used to aid data interpretation
- Different classes of therapeutic compound will have different considerations for Pre-clinical immunogenicity assessment based on associated risks

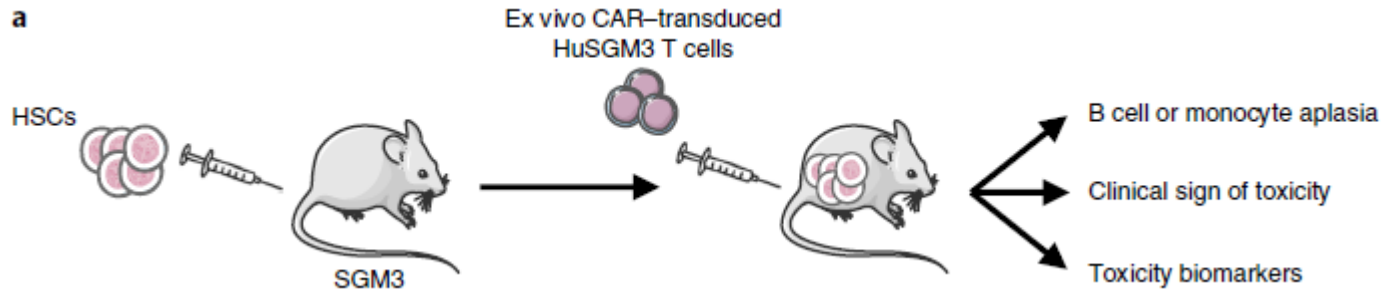
# The Challenge for Industry

- Nonclinical assessment of immunomodulatory therapies lags behind traditional toxicology, because of the complexity of the immune system and its interaction with disease states

Current models do not fully predict outcomes



# Building better models



**Figure 1.3:** (adapted from Norelli et al. Nat Med. 2018): Generation of CRS model. SGM3 mice are i.v. infused with human hematopoietic stem cells (HSCs). Four weeks after that, they receive CAR-modified T cells and are monitored for B cell or monocyte aplasia, clinical signs of toxicity and toxicity biomarkers.

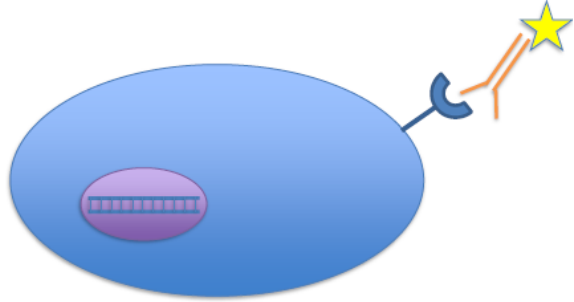


Q: How do we address the gaps in translating non-clinical work

# Cell therapy – Exposure, persistence and immunogenicity

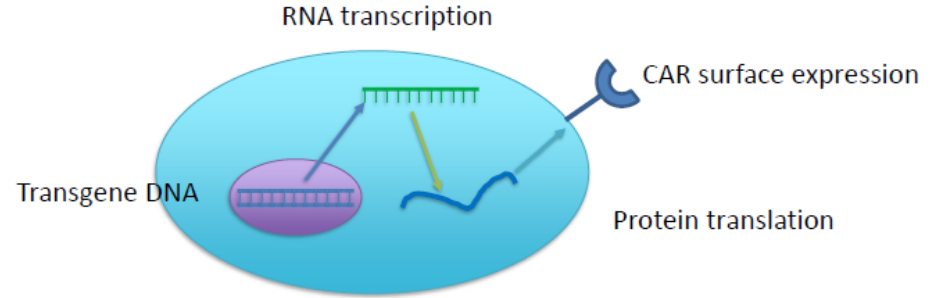
- Flow cytometry
- qPCR (real-time or endpoint, cf. WS5, Rob Nelson and Chris Cox)
- LBA, CBA

**Flow cytometry:**  
direct measure of the CAR-T  
 therapeutic agent



Technical and logistical limitations  
 Sensitivity – Stability  
 => Alternative

**Quantitative PCR:**  
 CAR-T concentration is inferred from the  
 level of transgene DNA



Does



CAR surface expression

=

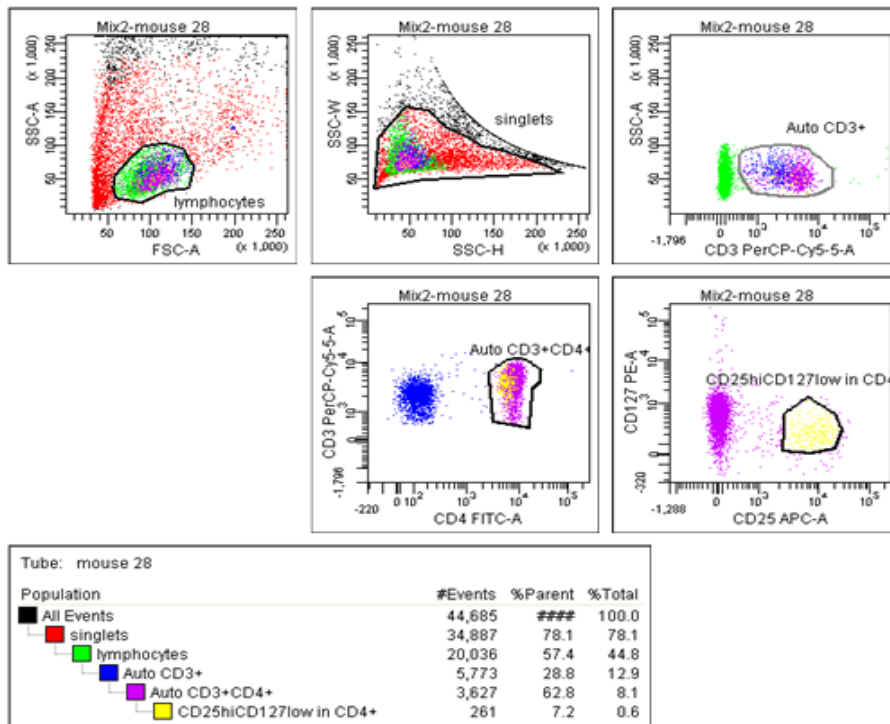
Transgene DNA



?

Correlation between flow cytometry (cell measurement) and qPCR results (transgene measurement) for exposure to CAR-T - [Mueller et al \(Blood, 2017\)](#): Cellular kinetics of CTL019 in relapsed/refractory B-cell acute lymphoblastic leukemia and chronic lymphocytic leukemia -> Correlation performed on the PK parameters

# Flow cytometry



- Generally used to look for effects of a biotherapeutic on immune cell populations in the blood or spleen homogenate
- Antibodies are used to identify cells by detecting specific antigens expressed by these cells, which are known as phenotypic markers
- Same principles as ligand binding assay but generally performed on fresh samples

# EBF

## Considerations for Cell Population & Receptor Expression

- Cell population:
  - ▶ Low numbers can make it difficult to measure and may require the implementation of more cell markers

### RECEPTOR EXPRESSION:

- ▶ Levels of expression can vary between healthy and disease states
- ▶ Receptor can be shed or internalised
- ▶ Expression levels can be variable across species
- ▶ Target may be on non-circulating tissues



## Cell therapy – Exposure and persistence

- Flow cytometry
- qPCR

Q: What should be reported and why?

Q: How important is this data?

Q: Who should do the analysis?

BioA lab with no experience in the technology or lab with technological experience but no BioA experience?

# Acknowledgment

## EBF Members

- Chris Cox – Psioxus
- James Munday – Covance
- Johannes Stanta – Covance
- Lydia Michaut – Bioagilytix
- Manuela Braun – Bayer
- Robert Nelson – Covance

## Non-EBF members

- Paula Miranda – uniCure

Thank You!



# Contact Information

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**European Bioanalysis Forum vzw**

[www.e-b-f.eu](http://www.e-b-f.eu)

## Additional Questions to the community

- Where do oligonucleotides fall into?
  - > DNA oligos and siRNA (ONPATRO) approved by FDA-CDER
  - ≠ other GTs are approved by FDA-CBER
  
- Current guidelines
  - Ambiguities in nomenclature
  - Gaps