

TAKING INSULIN ANALYSIS TO THE NEXT LEVEL

APPLICATION OF NEW SCIENCE IN ADVANCED LC- MS BASED WORKFLOWS

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OUTLOOK

1 Charles River Insulin project

2 Top-down insulin analysis

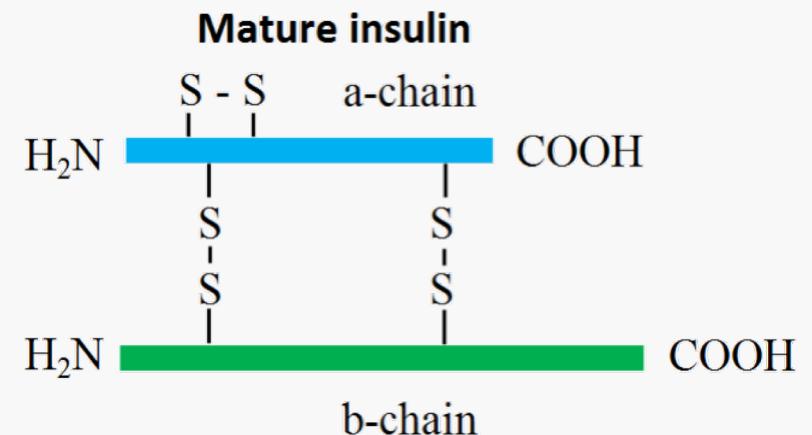
3 Bottom-up insulin analysis

4 Conclusions

Charles River Insulin project

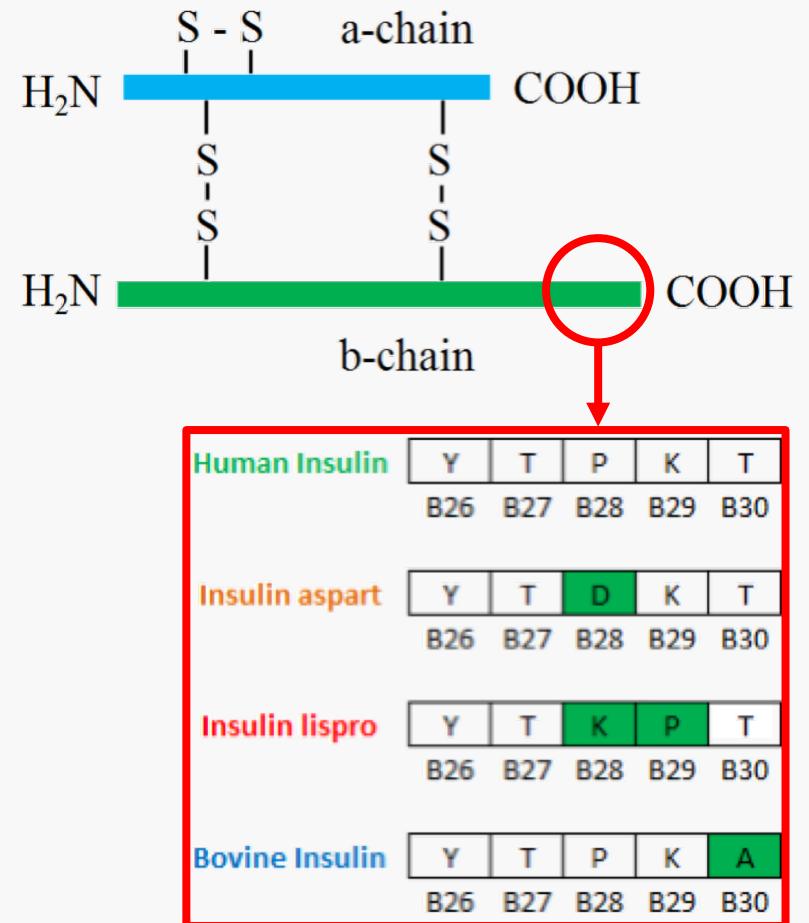
INSULIN

- Diabetes
- Insulin hormone
 - Regulates blood sugar levels
 - Small protein (biopharmaceutical)
- Quantification of Insulin
 - Pharmacokinetics
 - Therapeutic drug monitoring
 - Diagnostics
 - Doping control
 - Forensics



INSULIN ANALOGUES

- Analogues of human or animal origin
 - Human insulin
 - Bovine insulin
- Drug analogues by recombinant biosynthetic technologies
 - Insulin aspart
 - Insulin lispro
 - Insulin glargine
- Analogue specific region on b-chain
 - Insulin aspart: aspartic acid (D) on B28
 - Insulin lispro: interchange lysine (K) and proline (P)
 - Insulin glargine:
 - Two additional arginine (R) at end of b-chain
 - Glycine (G) on A21
 - Bovine insulin: alanine (A) on B30



TOP-DOWN OR BOTTOM-UP

- Top-down
 - Eluent conditions highly critical
 - Multiple charge states
 - Sensitivity lower
 - CID fragmentation less effective
 - Optimization by infusion
- Bottom-up
 - Proteolysis: enzymatic breakdown of protein
 - More complex sample preparation
 - Few charge states
 - Higher sensitivity
 - Optimization after LC separation



Figure sources:

Waters cooperation , 2011, 720003397EN rev B
Figure of QTRAP 6500+ provided by courtesy of AB Sciex Pte. Ltd

Top-down insulin analysis

PROJECT OBJECTIVE TOP-DOWN

- To set up analogue specific method for:
 - Insulin aspart
 - Insulin lispro
 - Insulin glargine
- Wide analytical range:
 - LLOQ below normal fasted endogenous insulin levels: 350 pg/mL
 - ULOQ at maximum endogenous insulin level: 10000 pg/mL

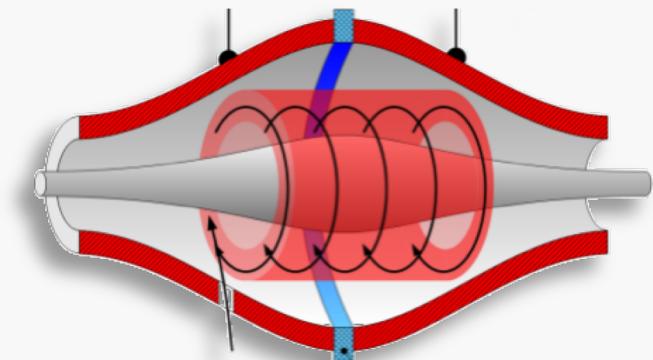
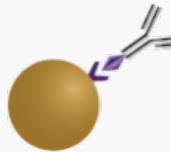


Figure sources:

Waters cooperation , 2011, 720003397EN rev B
<http://planetorbitrap.com/data/fe/image/QExactive.jpg>

LIGAND BINDING PURIFICATION

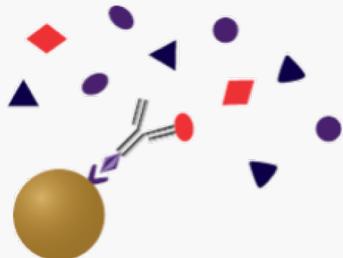
- Biotinylated antibodies



- Antibodies immobilized on streptavidin magnetic beads
 - Strongest non-covalent bond in nature



- Insulin capture



- Unspecific binding removed by bead wash



- Antigen elution

- Acidify to remove antigen from antibody

Figure sources:

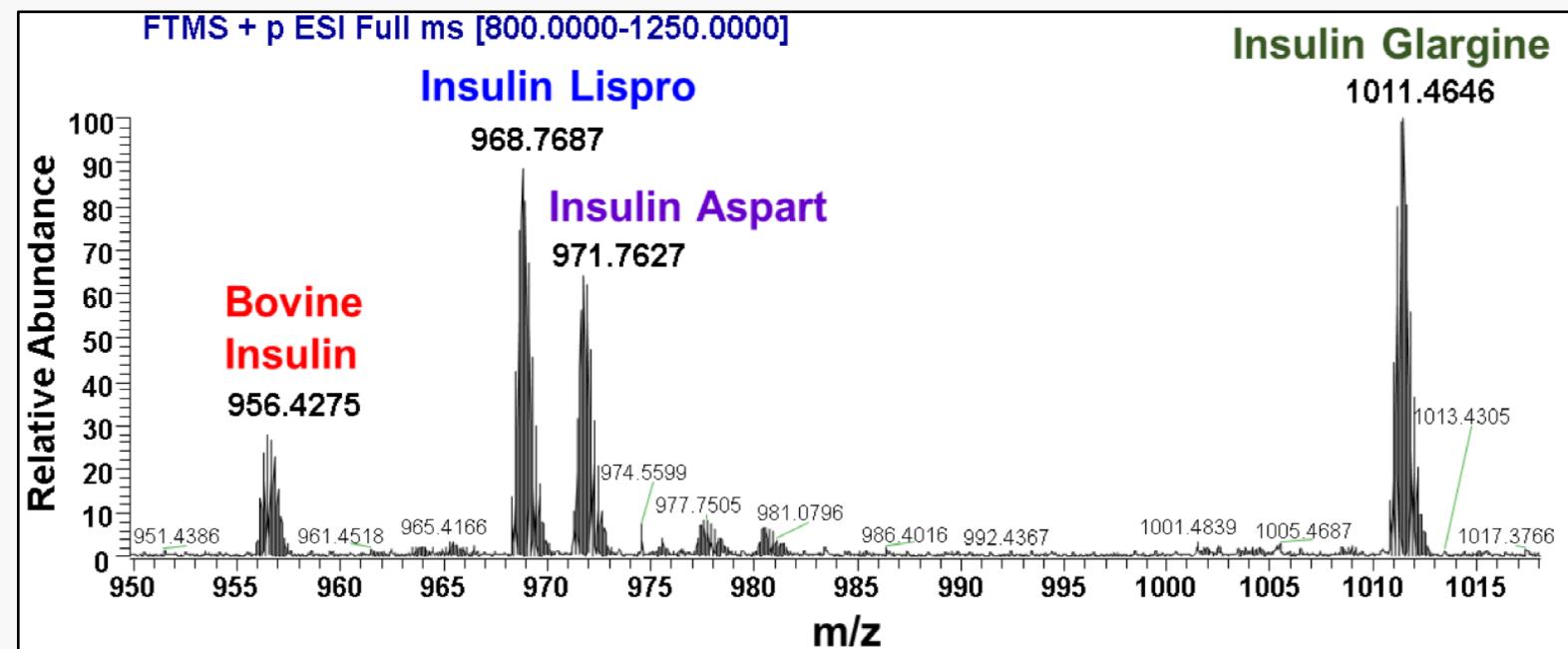
Dynabeads Streptavidin manual, ThermoFisher, 2016, CO020153 0316

<http://www.magnamedics.com/index.php/mm-separator-m96>

DOI: 10.1016/j.optlaseng.2017.09.016

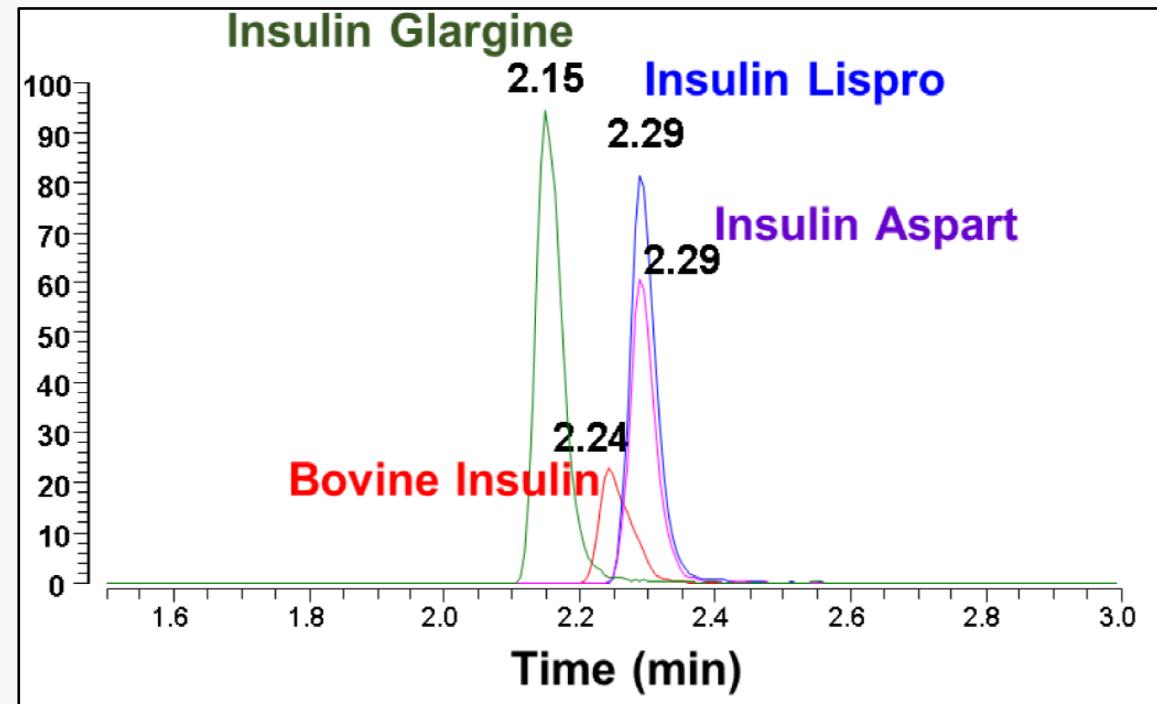
HIGH RESOLUTION MASS SPECTROMETRY, OPTIMIZATION

- HR-MS Thermo Q-Exactive
- Easy optimization of $(M+6H)^{6+}$ ions
- Analytes:
 - Insulin lispro
 - Insulin aspart
 - Insulin glargine
- Internal standard:
 - Bovine insulin



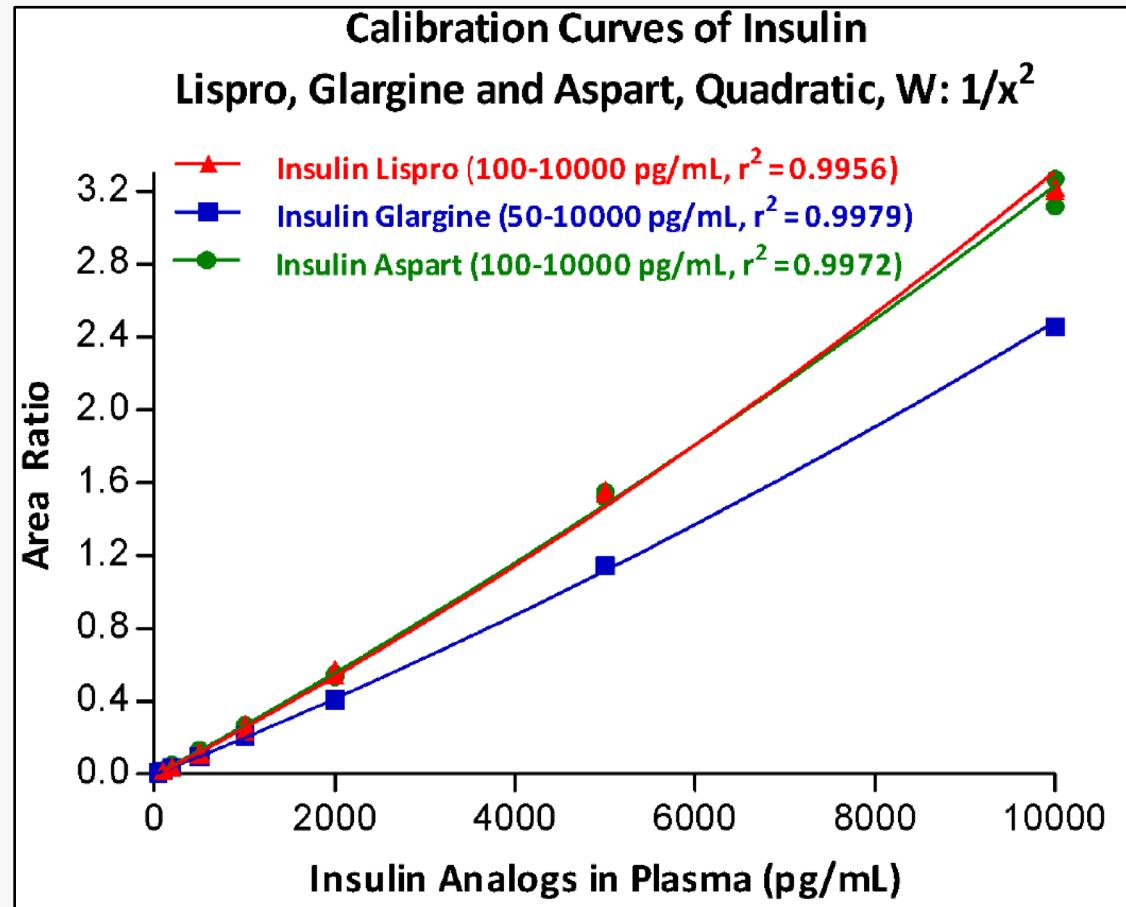
SAMPLE PREPARATION & CHROMATOGRAPHIC SEPARATION

- Matrix: Minipig plasma
- 250 µL of sample processed by Ligand binding purification
- UPLC
- Reversed phase column
- 0.4 mL/min



RESULTS LBA-LC-HRMS

- Quantification by adding up analogue isotope responses
- Extracted ion chromatogram (EIC)
- Quadratic regression
- Good correlation $r^2 > 0.99$ ($n = 2$)
- Background signal in matrix blank was less than 20% of LLOQ



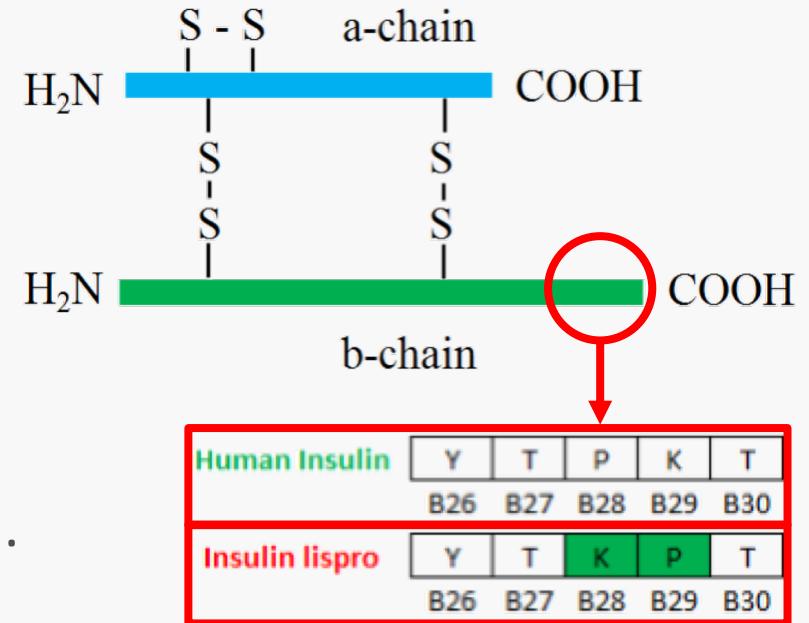
Data and figure sources:

Poster AAPS PharmSci360, Nov 2018, T0930-03-017

A Hybrid LBA/LC-MS Assay for Multiplex Quantitation of Insulin Analogs in Plasma, H. Wang, et al., CRL Ashland

HUMAN INSULIN / INSULIN LISPRO SEPARATION CHALLENGE

- Interchange lysine (K) and proline (P)
- Intact insulin LC-MS
 - Separation by LC very hard
 - MW identical
 - Comparable fragments
 - Interference according to S. Taylor et al. (2016)¹
- Intact insulin 2D-LC-MS published by E. Chambers et al. (2014)²
 - Limited retention time difference (4.28 / 4.30 minutes)
- What about only analyzing analogue specific region?
 - Bottom-up approach



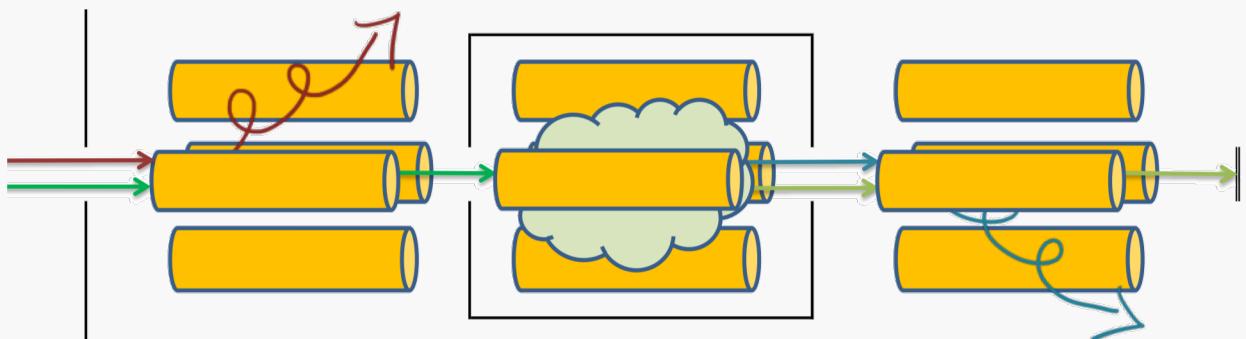
¹ DOI: 10.1016/j.cca.2016.01.019

² DOI: 10.1021/ac403055d

Bottom-up insulin analysis

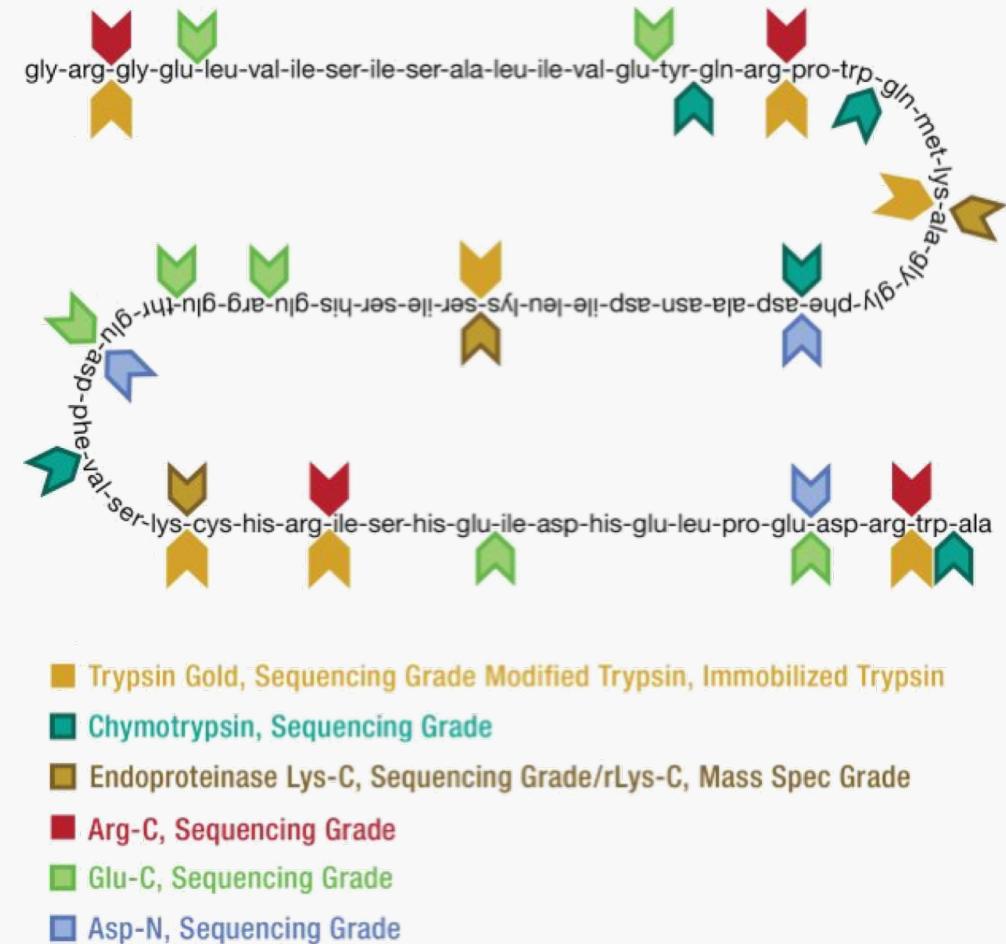
PROJECT OBJECTIVE BOTTOM-UP

- To set up an LC-MS (QqQ) method for Insulin lispro
 - Analogue specific for insulin lispro
 - Larger Dynamic range compared to ELISA (2 decades)
 - Range: 0.07 to 20 ng/mL



BOTTOM-UP: PROTEOLYSIS

- Enzymatic breakdown of protein to peptides
- Trypsin protease
 - Arginine residue (R)
 - Lysine residue (K)
 - No cleavage when proline at C-terminus
- Glu-C protease
 - Glutamic acid residue (E)
 - Aspartic acid residue (D) in presence of PO_4
- Next: *In-silico* proteolysis



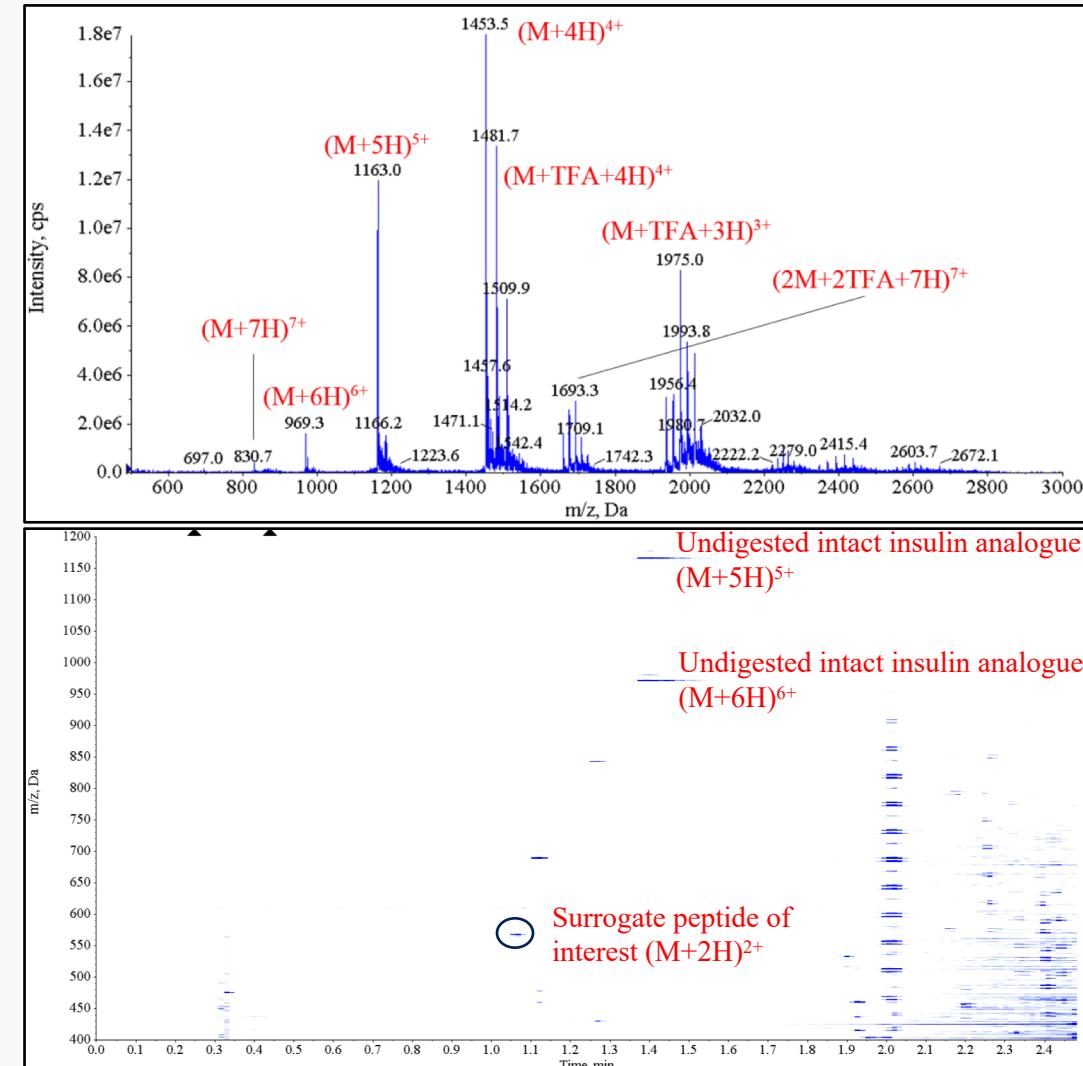
PROTEOLYSIS

- Proteolysis
 - Enzymatic breakdown of protein to surrogate peptides
 - Trypsin protease
 - Glu-C protease
 - Glutamic acid residue (E)
 - First: *In-silico* proteolysis
- BLAST
 - Unique surrogate peptides
 - Generic surrogate peptides

Insulin analogue	Trypsin proteolysis	Glu-C proteolysis
Human insulin		RGFFYTPKT
Bovine insulin	GFFYTPK	RGFFYTPKA
Rat insulin-1		RGFFYTPKS
Rat insulin-2	GFFYTPMS	RGFFYTPMS
Insulin lispro	GFFYTKPT	RGFFYTKPT
Insulin aspart	GFFYTDK	RGFFYTDKT

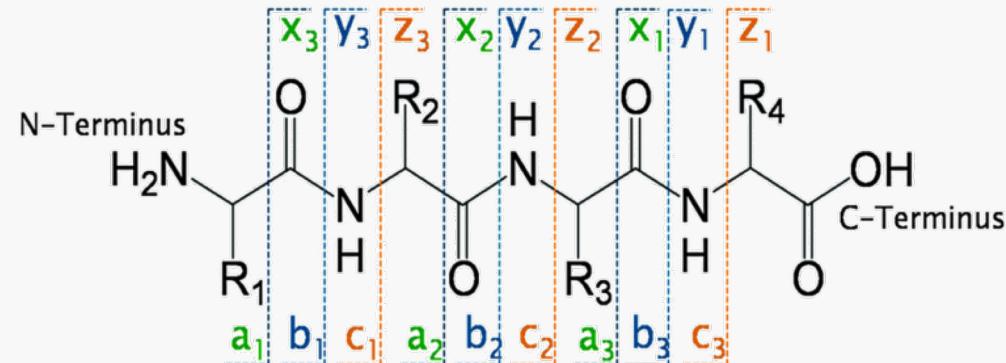
MASS SPECTROMETRY, PRECURSOR ION OPTIMIZATION

- Intact (top-down)
 - Like a small molecule: direct infusion
- Analogue specific region (bottom-up)
 - Infusion optimization not feasible
 - Sample too complex
 - LC separation before optimization needed.



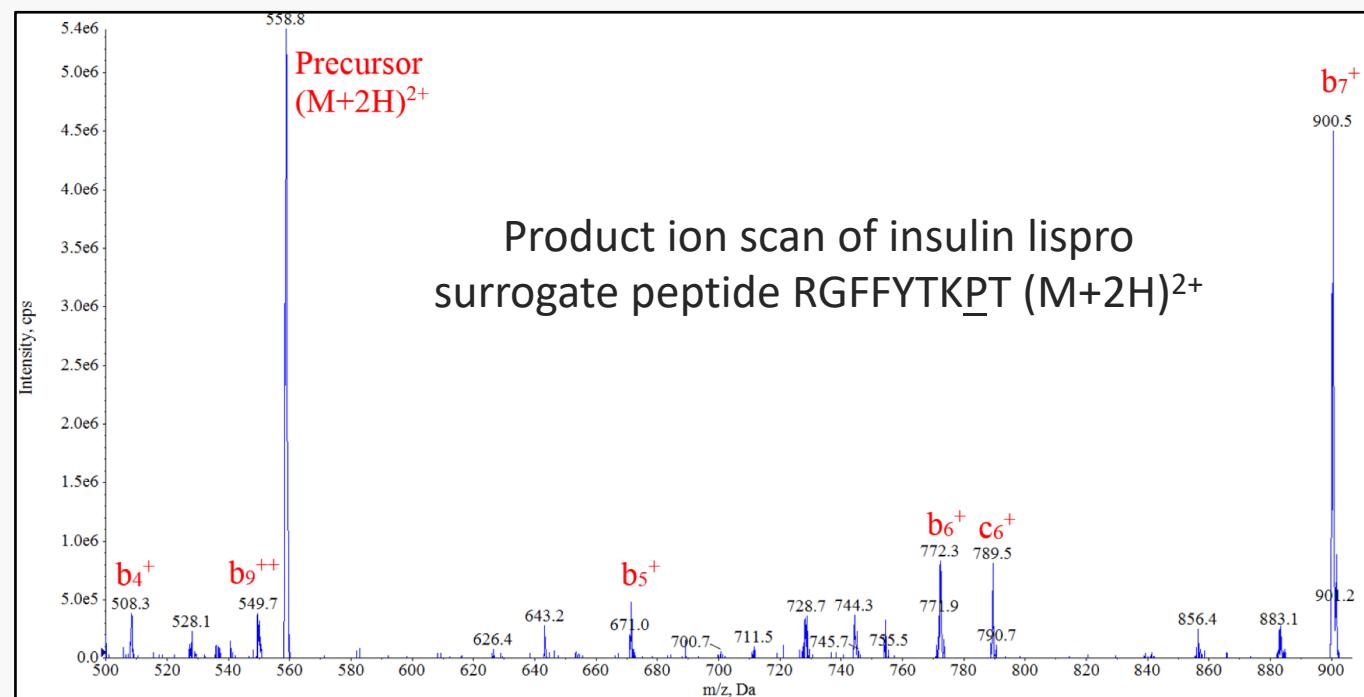
MASS SPECTROMETRY, PRODUCT ION OPTIMIZATION

- Nomenclature



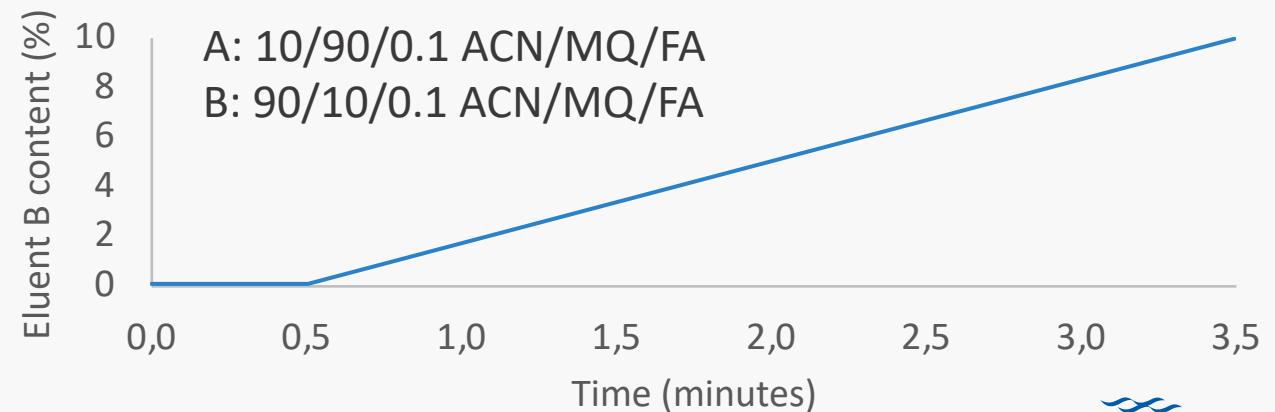
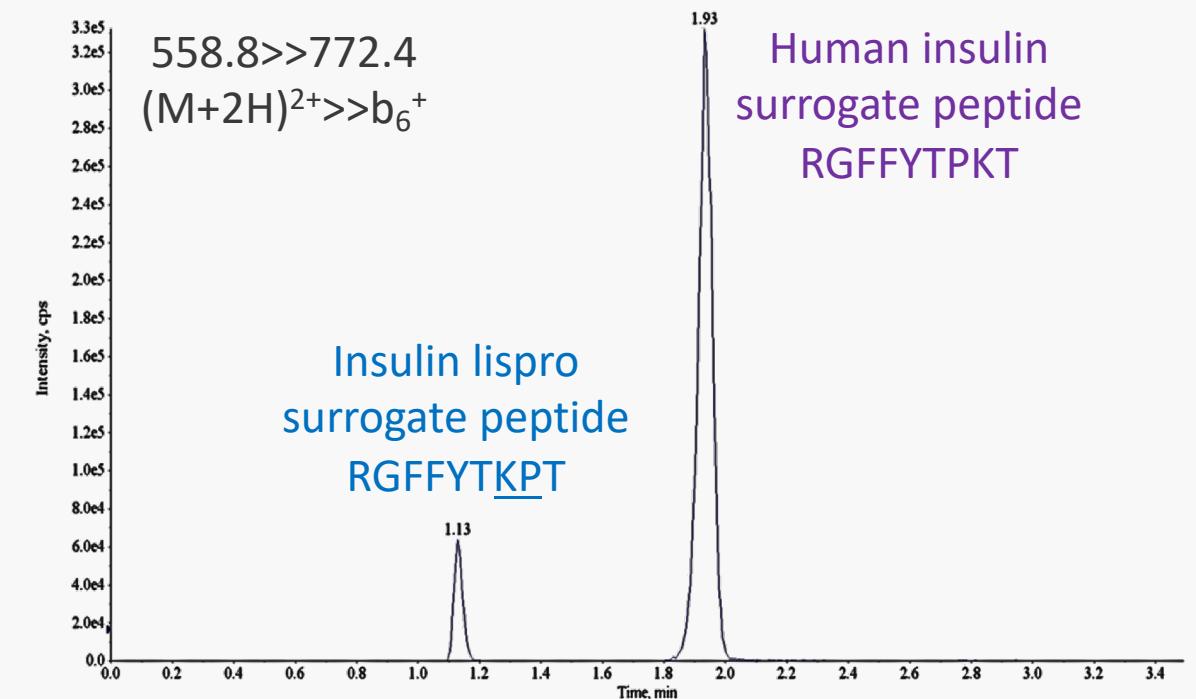
- Proline (P) effect

- Strong peptide bond (because of secondary amine)
- Predominance b_i and y_i ions formed at N-terminus proline



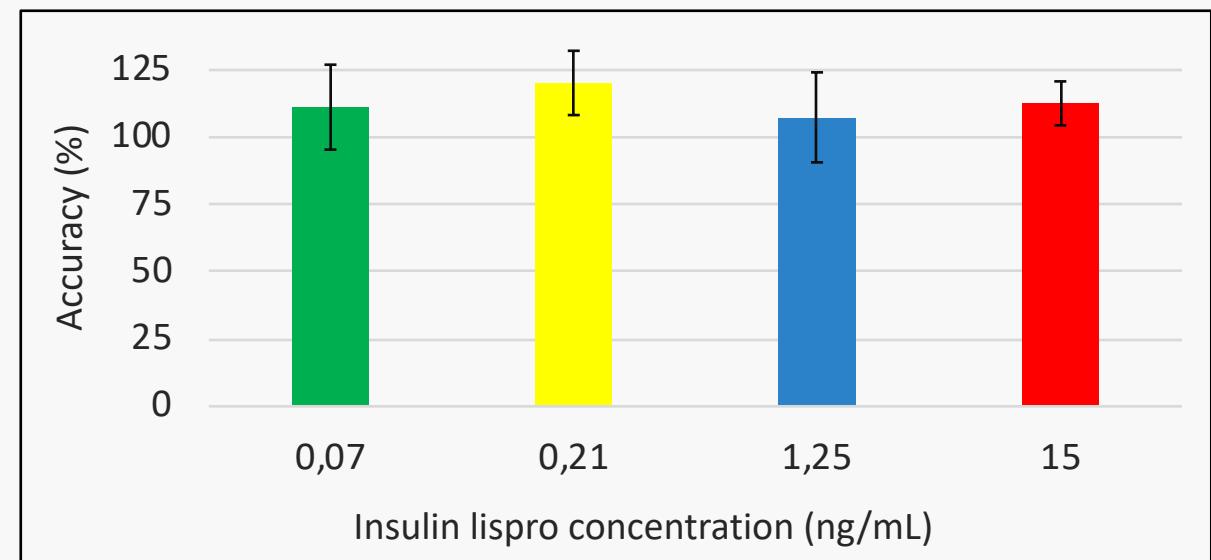
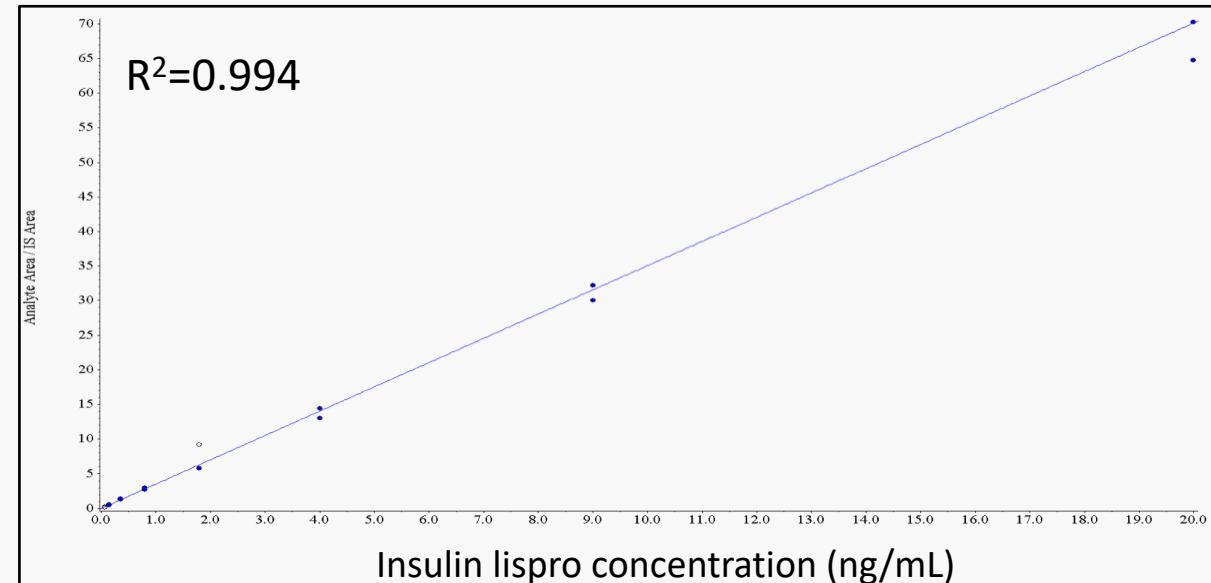
CHROMATOGRAPHIC OPTIMIZATION

- Reminder: Intact insulin quantification cannot separate insulin lispro from human insulin
- We can! Now we have the first method to perform insulin lispro and human insulin selective LC-MS quantification in one assay



INSULIN LISPRO

- 500 µL human K₂EDTA plasma processed
- Double Blank
 - No internal standard response
- Blank
 - No insulin lispro response
- Calibration samples
 - Duplicate sample preparation
 - Calibration curve according to criteria
 - 87.5 % accepted
- QC samples
 - 5 replicates of
 - QC-LLOQ 0.07 ng/mL
 - QC-L 0.21 ng/mL
 - QC-M 1.25 ng/mL
 - QC-H 15 ng/mL



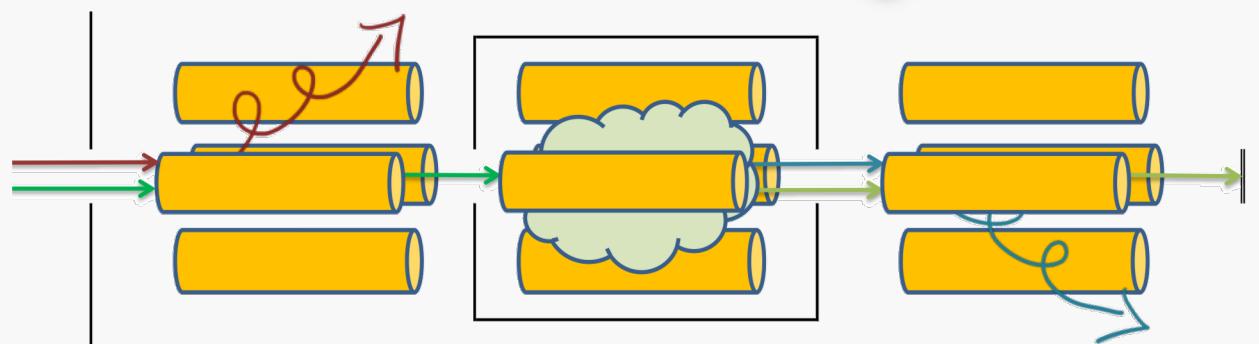
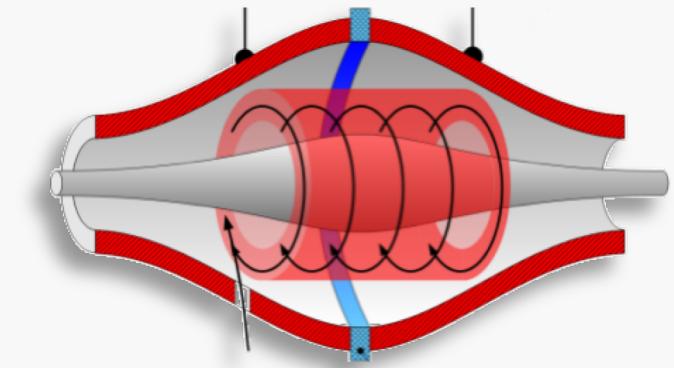
Conclusions

CONCLUSIONS

- LC-HRMS multiplex quantification
 - Easy method development
 - Method developed in analytical range 0.05 to 10 ng/mL for insulin glargine
 - 0.1 to 10 ng/mL for insulin lispro and insulin aspart
- Insulin lispro bottom-up LC-MS
 - Separation for insulin lispro and human insulin surrogate peptides
 - Method developed in analytical range 0.07 to 20 ng/mL insulin lispro

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Thank you

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