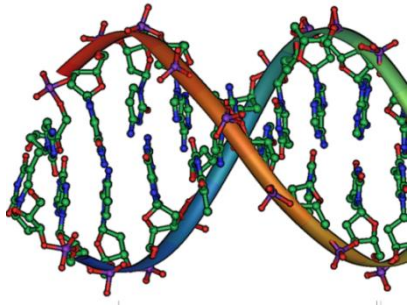


Bioanalytical LC-MS/MS of therapeutic oligonucleotides



W.D. van Dongen
PROXY Laboratories B.V.
wdvandongen@gmail.com

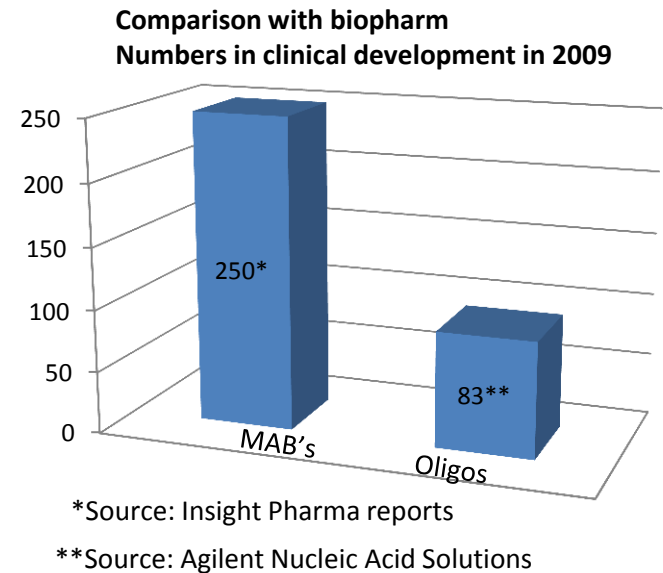
Oligonucleotide therapeutics

- RNA chains 15-25 nucleotide units
- cytosine (C) guanine (G), adenine (A) uracil (U)
- interferes with processing of genetic material
 - inhibits/decreases expression of therapeutically relevant protein
 - blocks expression of virus mRNA
- can treat diseases “undruggable” using small molecules or MABs
 - diseases with a genetic background e.g. DMD, cystic fibrosis, specific cancers and rare diseases
 - common diseases e.g. Hep-C, atherosclerosis, lupus, psoriasis
- Two types
 - single stranded: *e.g.* antisense RNA
 - double stranded: *e.g.* short interfering RNA



Oligonucleotide therapeutics

- Currently two drugs on the market
 - Vitravene[®] for cytomegalovirus infection (*herpes*)
 - Macugen[®] for wet macular degeneration (*loss of vision in the center of the visual field*)
- Many in development
 - >250 therapeutic programs
 - >100 in the clinic (2011)
 - >5 in phase III



Bioanalysis of oligonucleotides

- Method-of-choice: ELISA
 - based on hybridization of a probe or a capture and/or detection strand complementary to asRNA or siRNA
 - unsurpassed sensitivity (25 pg/ml)
- Drawbacks
 - cannot distinguish full-length oligonucleotides from truncated shortmer metabolites
 - overestimation of parent oligonucleotide
 - Cannot determine intact siRNA

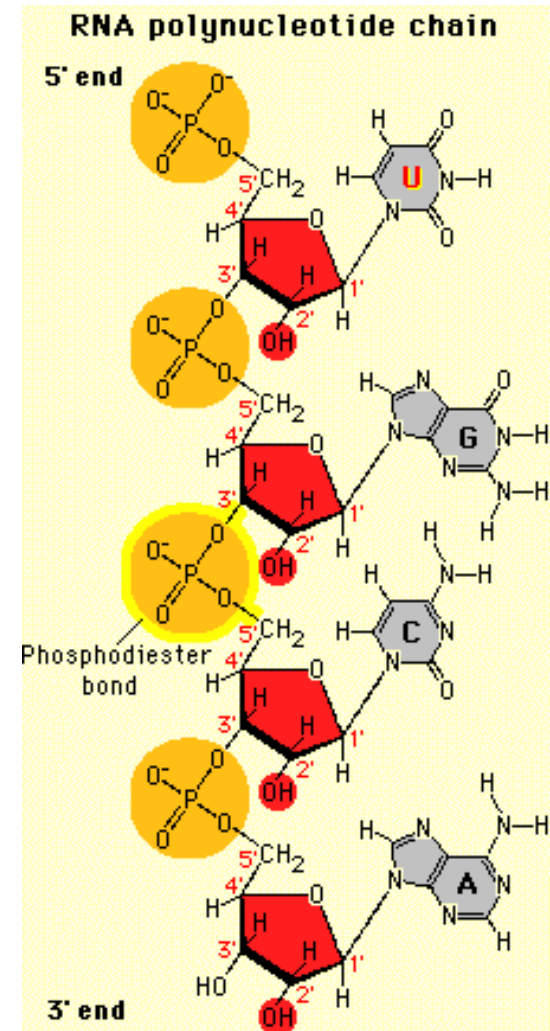
Bioanalysis of oligonucleotides

- LC-MS
 - unsurpassed selectivity : accurate levels
 - indentify and quantify metabolites
- Drawbacks
 - “best” reported sensitivity of validated method:
4 ng/ml [Deng et al, J Pharm Biomed Anal. 52(4),
571-579 (2010)]

Bioanalytical LC-MS/MS of therapeutic oligonucleotides

Highly challenging from an analytical perspective:

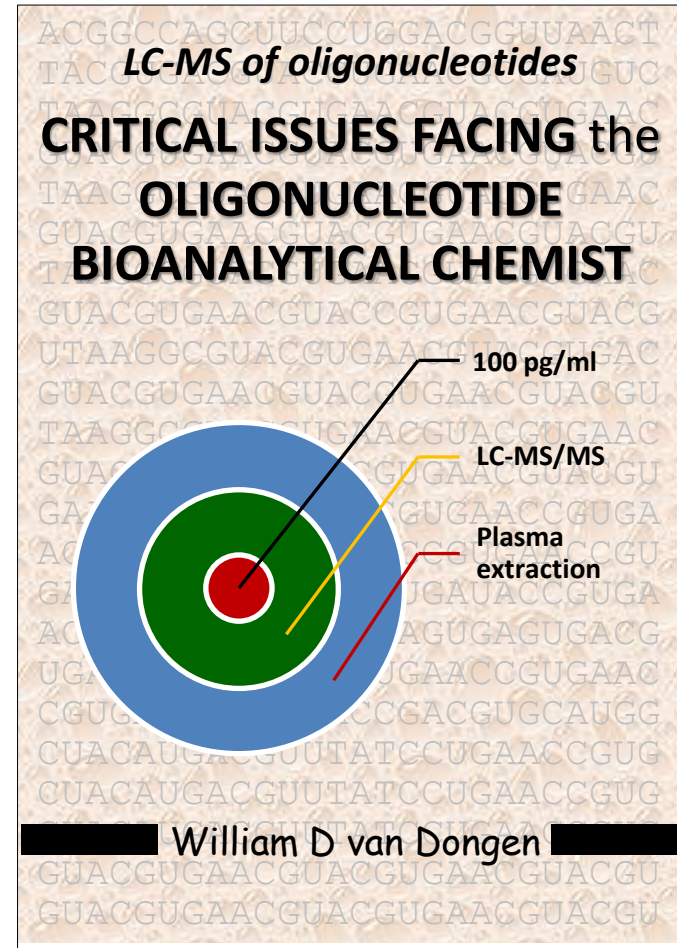
- Acidic proton at each phosphodiester bond
- Highly charged poly-anionic backbone
- Extremely polar
- Phosphorothioate linkage



Bioanalytical LC-MS/MS of oligonucleotides

Critical issues:

- LC-MS efficiency
 - retention vs. ionisation
- ESI-MS
 - multiple negative charge states
 - H⁺-alkali⁺ exchange at phosphate groups
- MS/MS
 - fragmentation of multiple charged OGNs
- Quantitation
 - internal standard selection
- Sample preparation
 - SPE
 - LLE



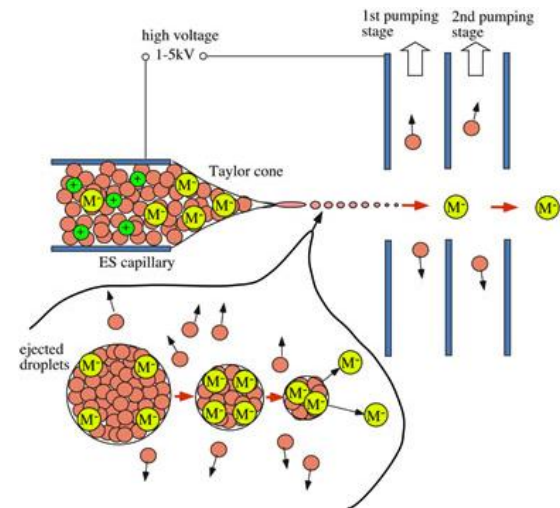
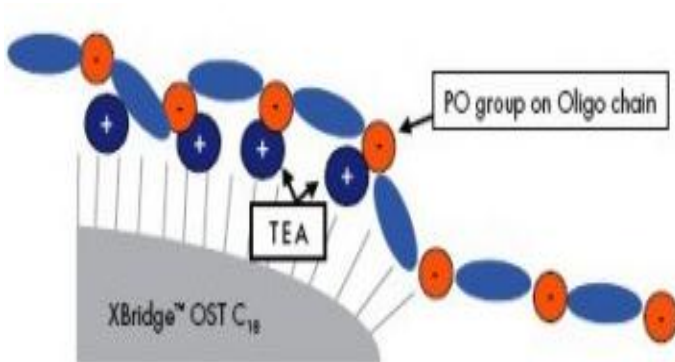
LC-MS efficiency:

chromatographic retention **vs.** ionisation efficiency dilemma

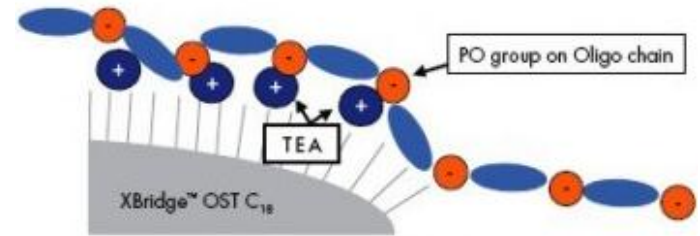
LC
low
low
ionpair

organic
pH
ions

ESI
high
high
low



IPLC-MS of oligonucleotides

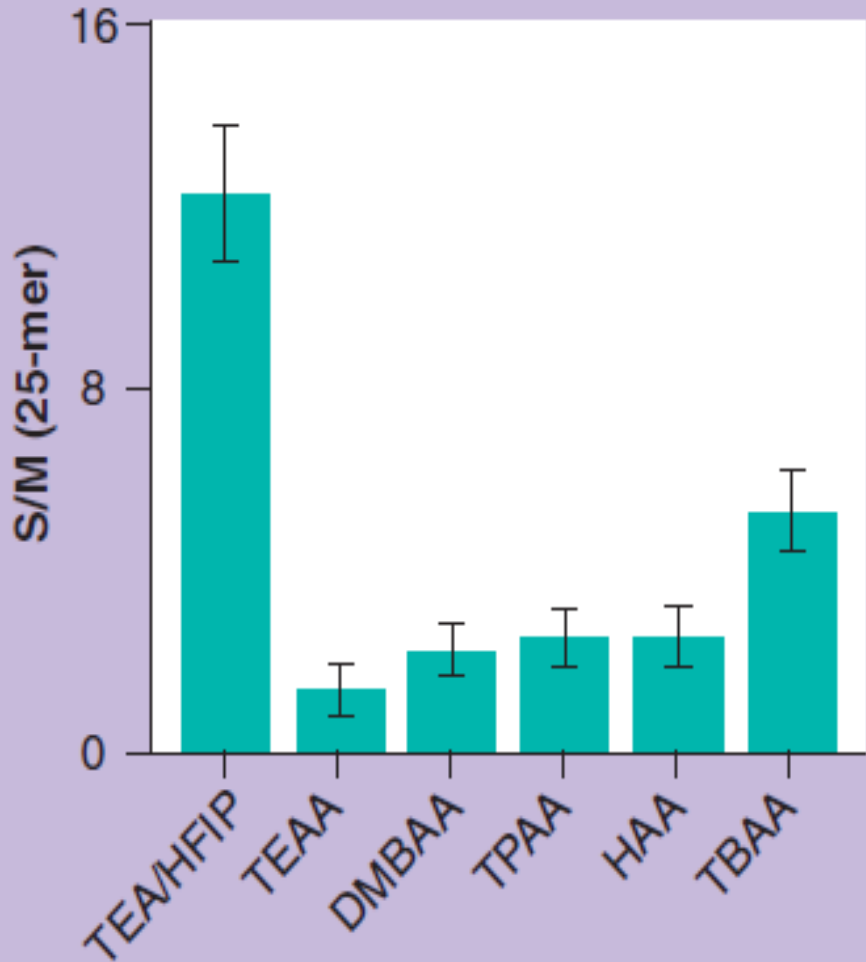


- C18 column
- 10 mM triethylamine (TEA)
- 100 mM hexafluoro-2-propanol (HFIP)
- MeOH/MeCN gradient

IPLC-MS of oligonucleotides

- LC: HFIP increases hydrophobicity ion-pair
- ESI: HFIP dynamic liquid/gas phase pH adjuster
 - pK_a 9, 99% not charged at pH 7, bp 57°C
 - evaporates during ESI
 - volatile HFIP depletes at droplet surface
 - pH at the surface rises to 10
 - OGN-TEA ion pair dissociation
 - desorption OGN into the gas phase.
- And: HFIP reduces cation exchange

Single-quadrupole S/N ratios (n=3) of 20 pg (dT)25 IPLC-MS



TEA/HFIP:

15mM triethylammonium /400mM hexafluoro-2-propanol

TEAA:

100mM triethylammonium acetate

DMBAA:

100mM butyldimethylammonium ac.

TPAA:

100mM tripropylammonium acetate

TBAA:

100mM tributylammonium acetate

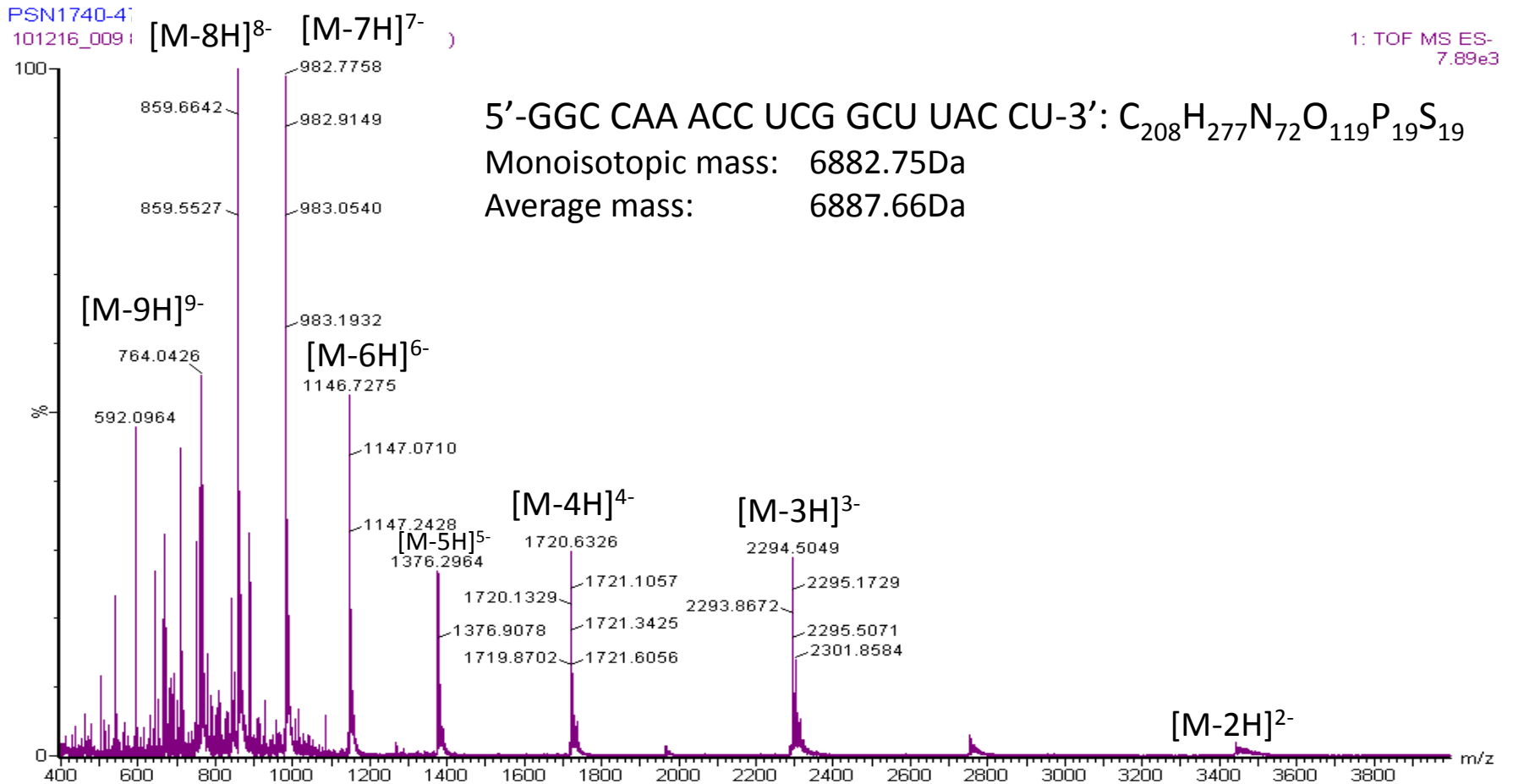
HAA:

100mM hexyl ammonium acetate

McCarthy *et al.* 5th Symposium on the Practical Applications of Mass Spectrometry in the Biotechnology and Pharmaceutical Industries. The Meritage Resort, Napa, CA, USA, 9–11 September 2008

ESI-MS:

formation of multiple negative charge states



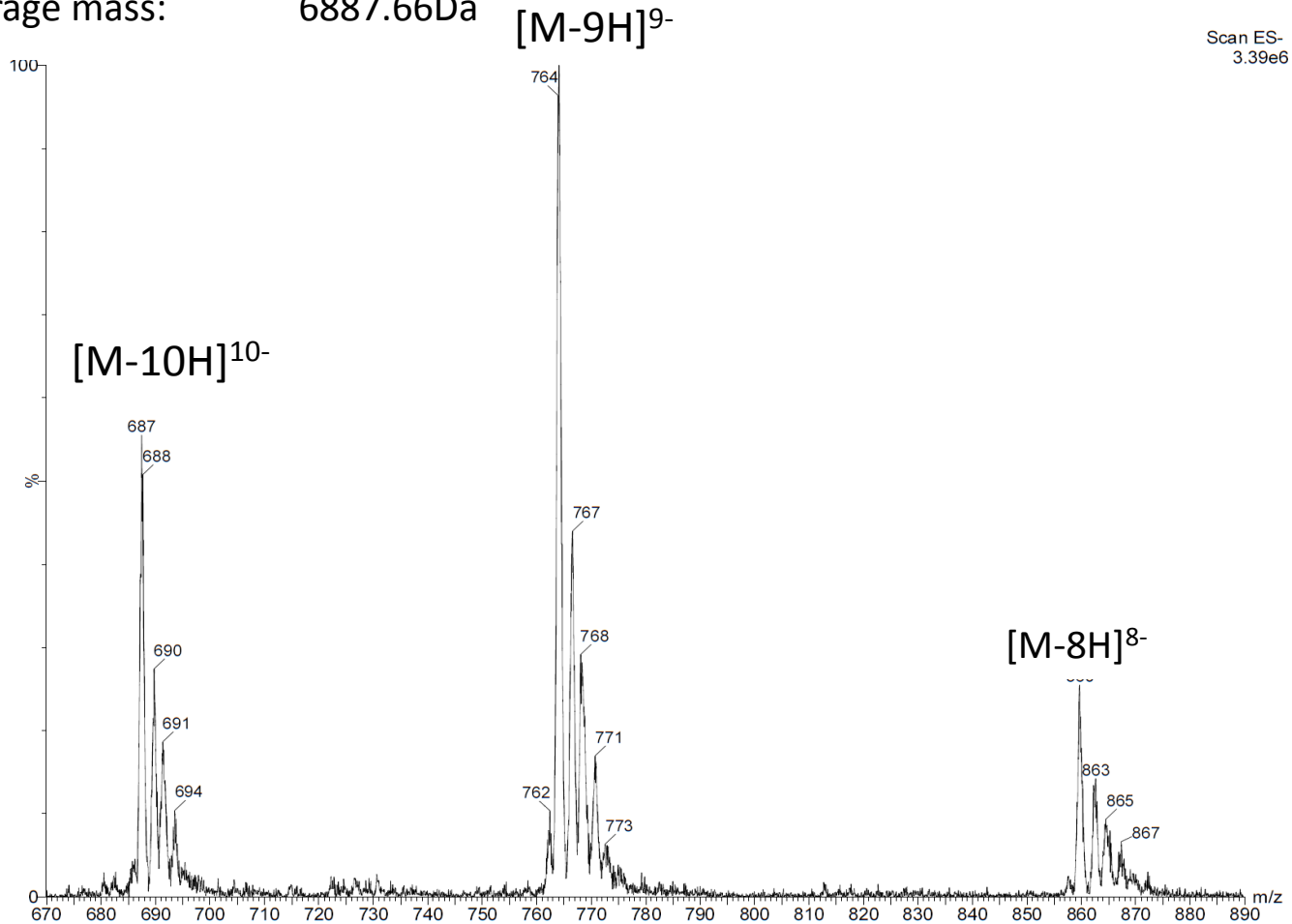
ESI-MS:

formation of multiple negative charge states

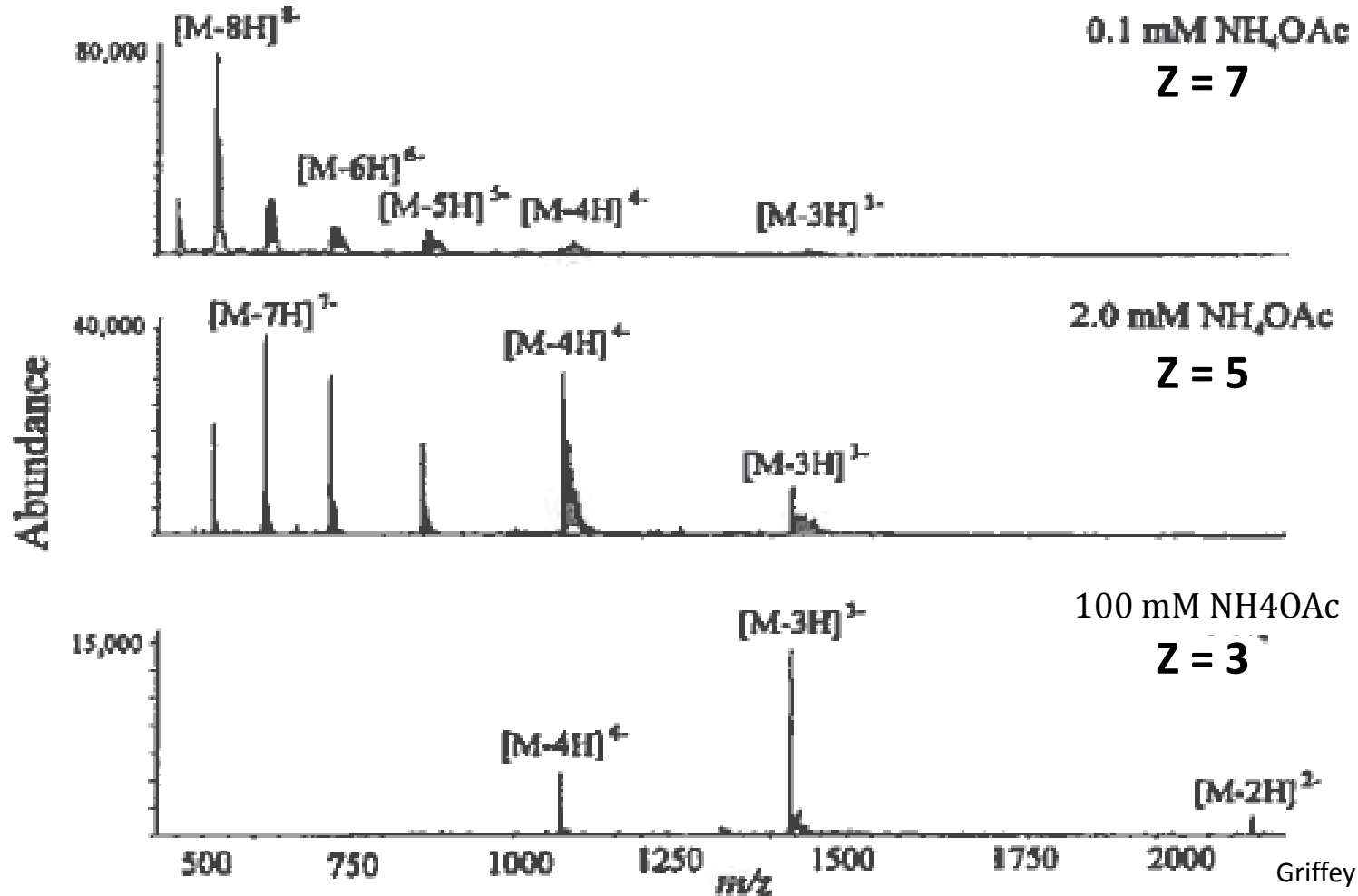
5'-GGC CAA ACC UCG GCU UAC CU-3': C₂₀₈H₂₇₇N₇₂O₁₁₉P₁₉S₁₉

Monoisotopic mass: 6882.75Da

Average mass: 6887.66Da



5'-GAGACTGCAAGCG-3'



Griffey *et al*,
JAmSoc 8,
155-160 (1997)

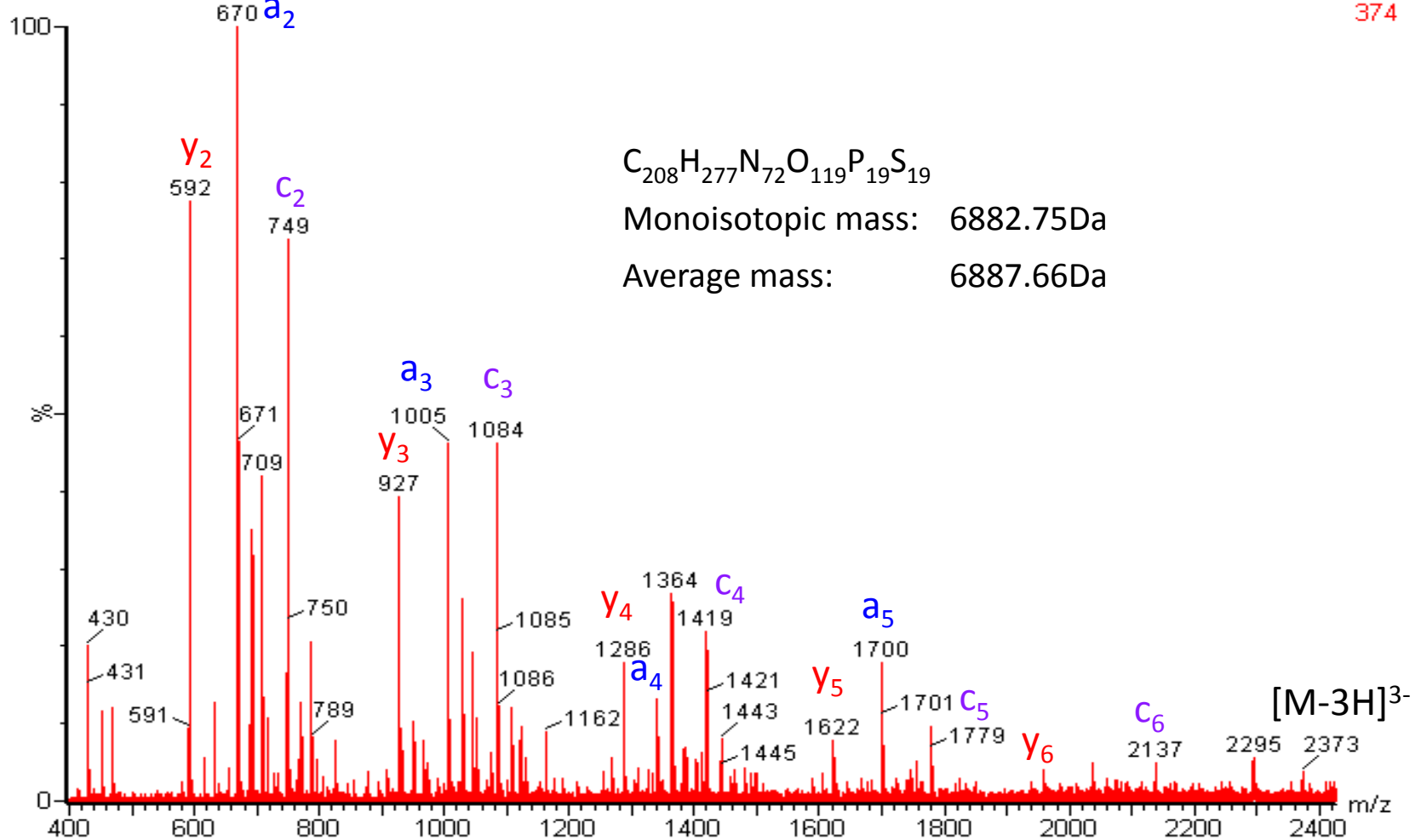
5'-GGCAAACCUCGCUUACCU-3'

MS/MS of [M-3H]³⁻

1740 0.1 mg/ml 5µl/min 15%B 0.3ml/min,CE60

110107_MSMS_002 122 (1.084) Cm (5:338)

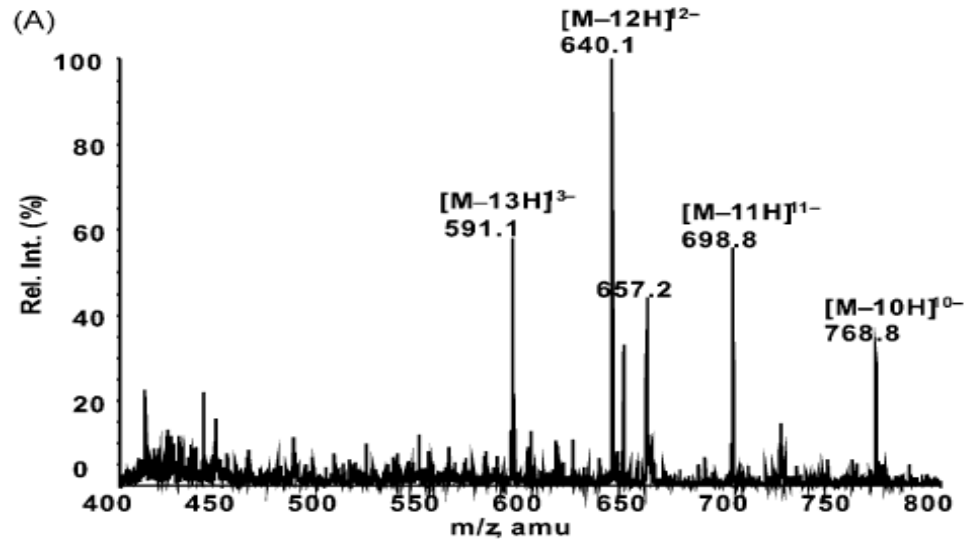
TOF MSMS 2293.21ES-
374



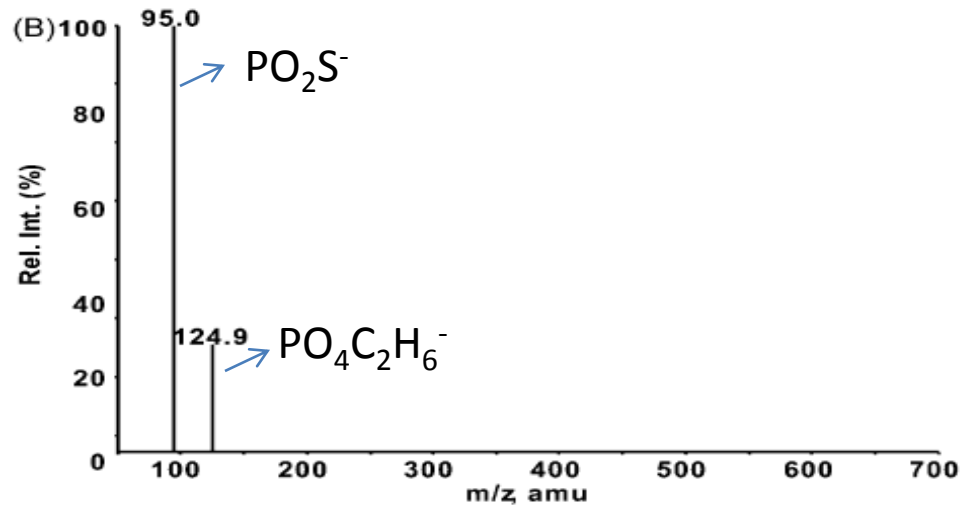
5'-TCTCCAGCGTGCGCCAT-3'

MS/MS of [M-13H]¹³⁻

Full scan m/z 400-800



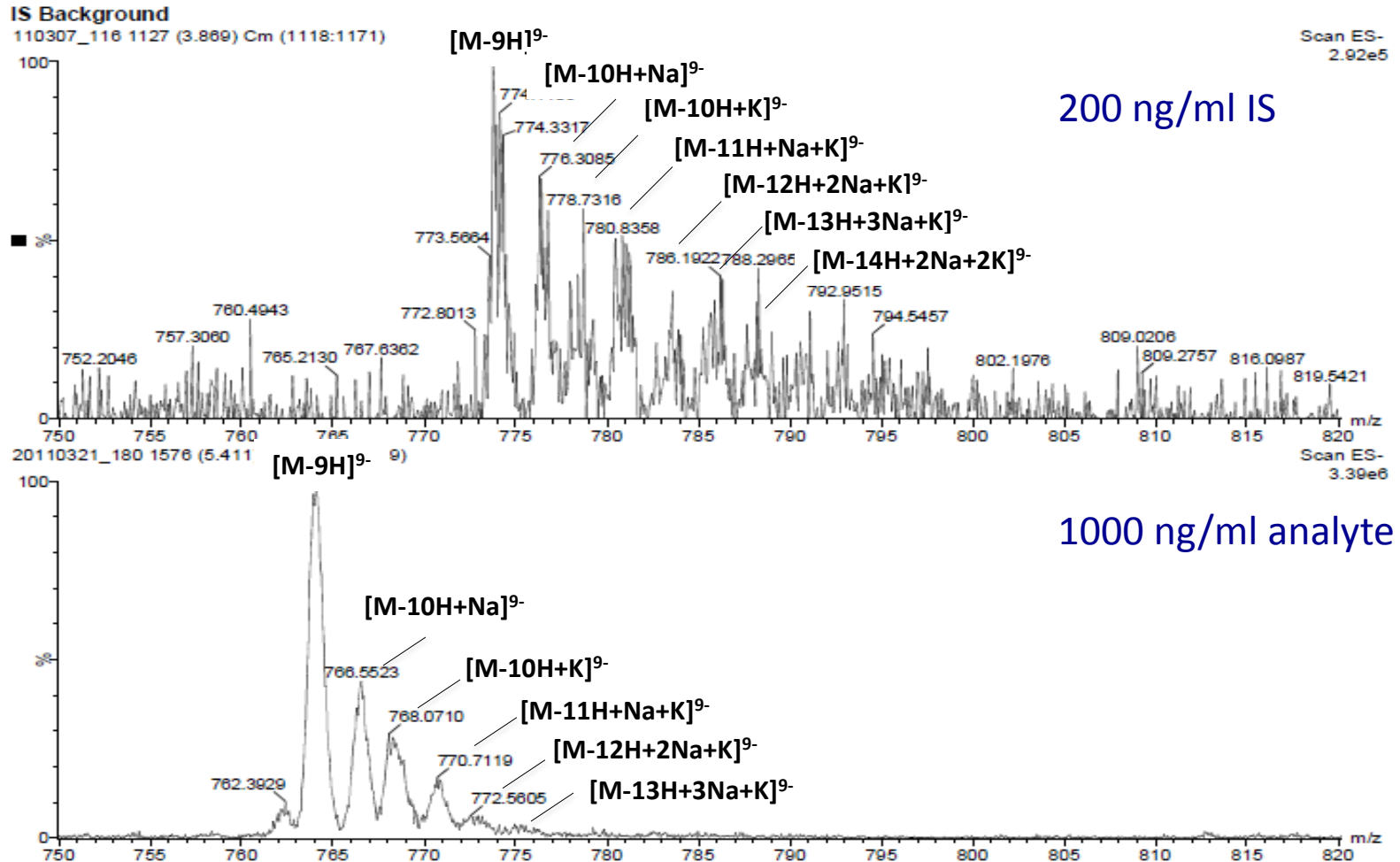
product ion scan of
m/z 591 [M-13H]¹³⁻



Deng et al, J Pharm Biomed Anal. 52(4),
571-579 (2010)

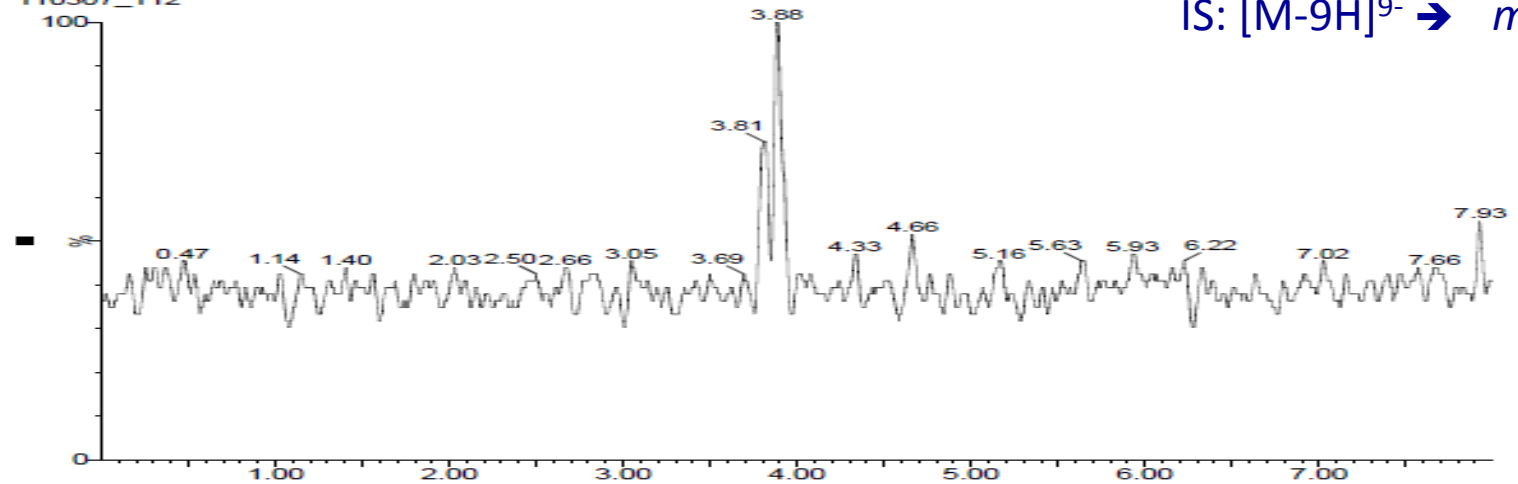
ESI-MS

H⁺-alkali⁺ exchange at phosphate groups



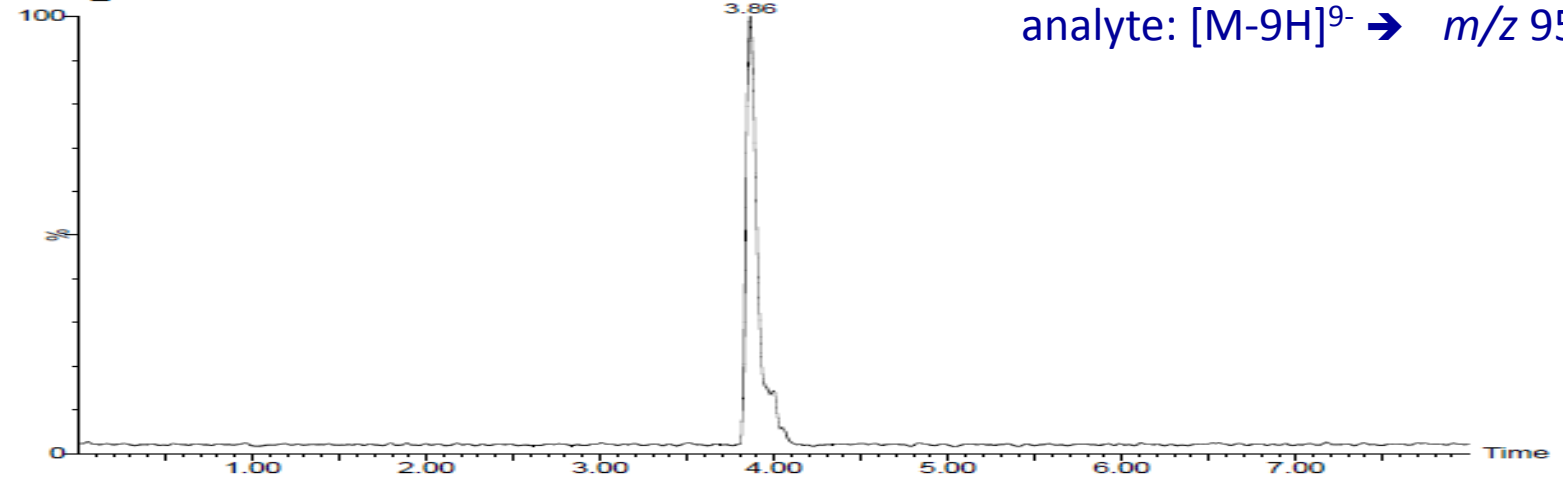
Quantitation: internal standard selection

IS Background
110307_112



IS: $[M-9H]^{9-} \rightarrow m/z 95$

110307_112



analyte: $[M-9H]^{9-} \rightarrow m/z 95$

Internal standard affairs: case 1

Drug substance:

rafAON $G_S TGCTCCATTGATG_S C$ mol mass: 4590

IS $G_S UGCUCCAUUGAUG_S C$ mol mass: 4521

Chemical modifications:

lower case S = PS linkage

LC:

no separation of rafAON and IS

SRM:

rafAON, $[M-3H]^{3-}$: 1529 \rightarrow 322 + 1529 \rightarrow 746

IS, $[M-3H]^{3-}$: 1506 \rightarrow 289

LOQ: 50 ng/ml

Internal standard affairs: case 2

Drug substance:

PF-ODN TCGTCGTTTTGTCGTTTTGTCGTT

IS: TTTTTTTTTTTTTTTTTTTTTT

mol mass:

7697

6327

LC:

no separation of PF-ODN and IS

SRM:

PF-ODN, $[M-xH]^{x-}$ 698.8, 640.1, 591.1 (n=11-13) → 95

IS, $[M-10H]^{10-}$: 631.7 (n=10) → 125

LOQ: 4 ng/ml

Deng et al, J Pharm Biomed Anal. 52(4),
571-579 (2010)

Sample preparation of plasma samples

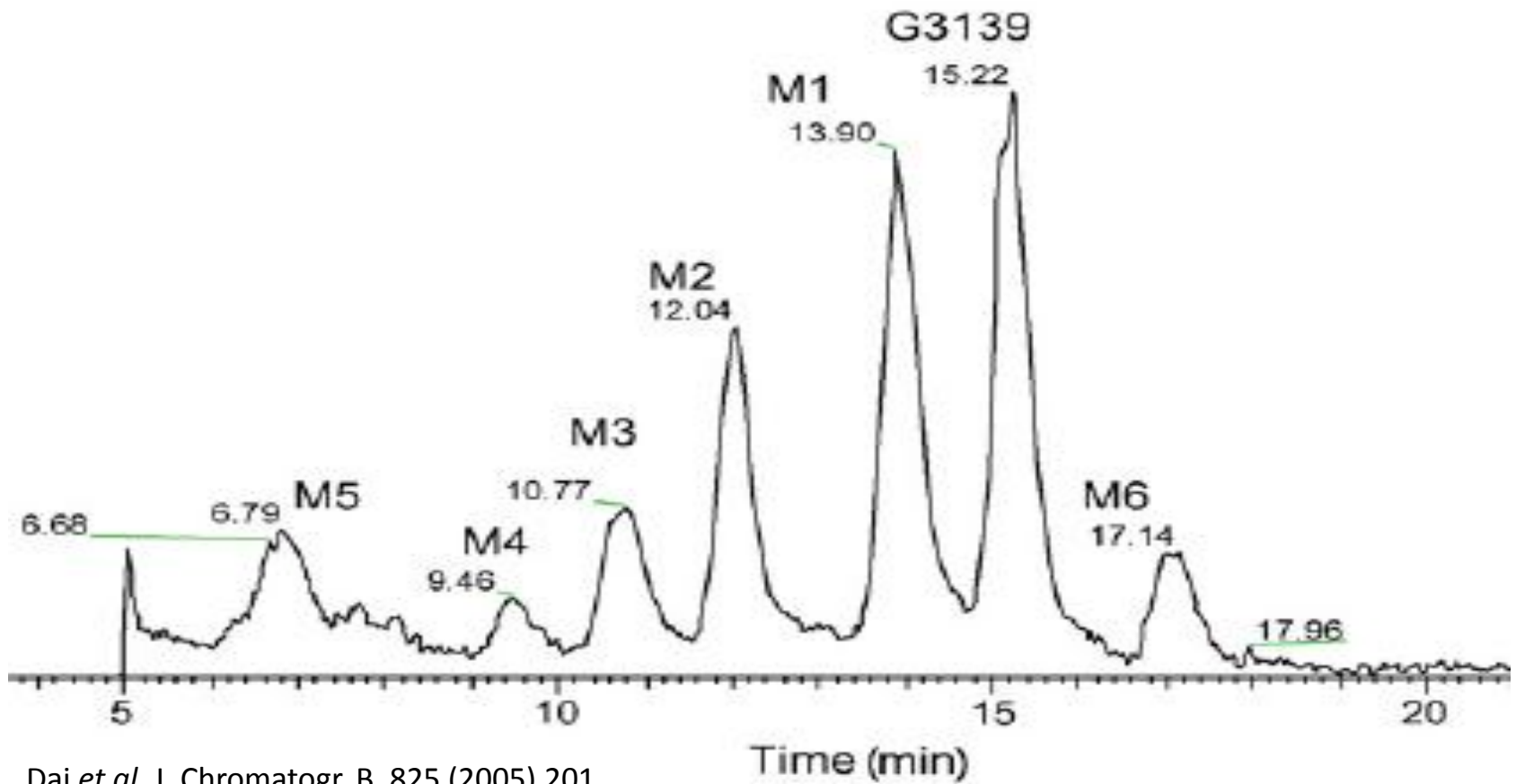
drug	sample prep	recovery	suppression
	<i>SPE</i>		
G3139	OASIS HLB	40-50%	na
rafAON	OASIS HLB	22.8 ± 6.5%	48.2 ± 3.5%
rafAON	Varian C18	80%	50%
Compound X	OASIS WAX	75%	na
Oligo1	Clarity OTX	>80%	na
	<i>LLE</i>		
Compounds A/B/C	phenol/ chloroform/ isoamyl alcohol	80%	na
	<i>LLE & SPE</i>		
PF-ODN	chlorof/phenol & Oasis HLB	70-80%	0-6%

Bioanalytical LC-MS methods of asRNA

drug	sample prep	range	validation statistics
ISIS 1083 21-mer	SPE Phenyl	5-500 ng/ml	89-107% RSD 2-15%
rafAON 15-mer	monkey (OASIS HLB) mouse (OASIS C18)	50-10,000 ng/ml 25-5,000 ng/ml	94-102% RSD 6-14% 95-101% RSD 3-11%
PF-ODN 24-mer 5'-TCGTCGTTTTGTCGTTTTGTCGTT-3'	LLE, chloroform/phenol & Oasis HLB	4-2.000 ng/ml	PF-ODN: 97-101% RSD 2-12% (n-1)5'/(n-1)3': 102-106% RSD 1-12% (n-2)5': 99 - 94% RSD 1-12 % (n-3)5': 90- 100 % RSD 6-7%

WD van Dongen and WMA Niessen, *Bioanalysis* (2011) 3(5), 541-564

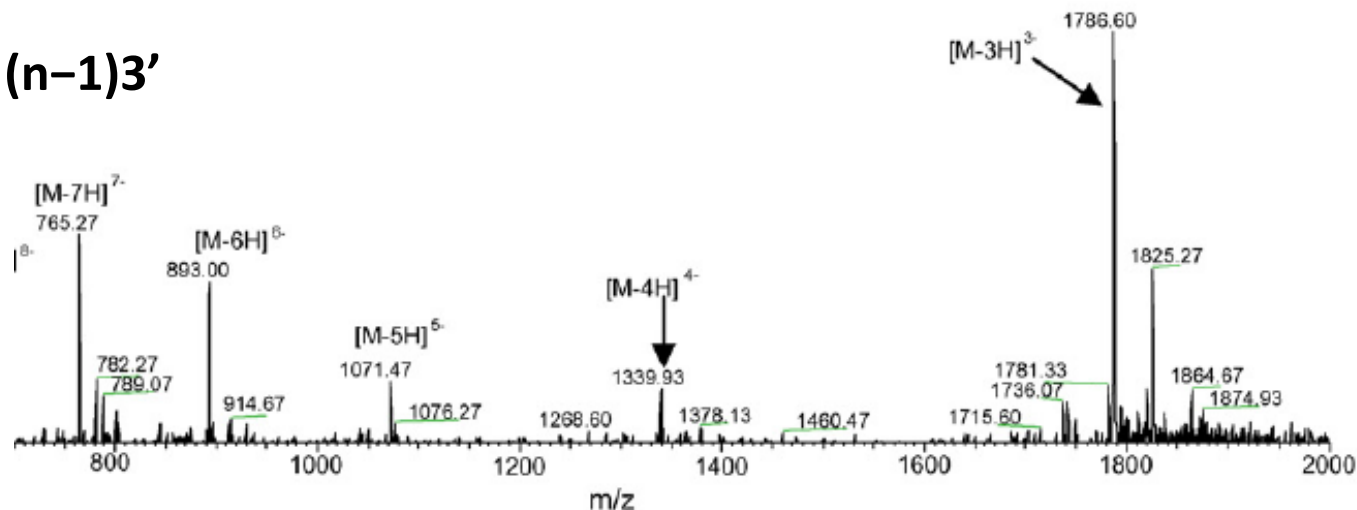
Metabolites of antisense G3139 in human plasma obtained from *in vivo* study



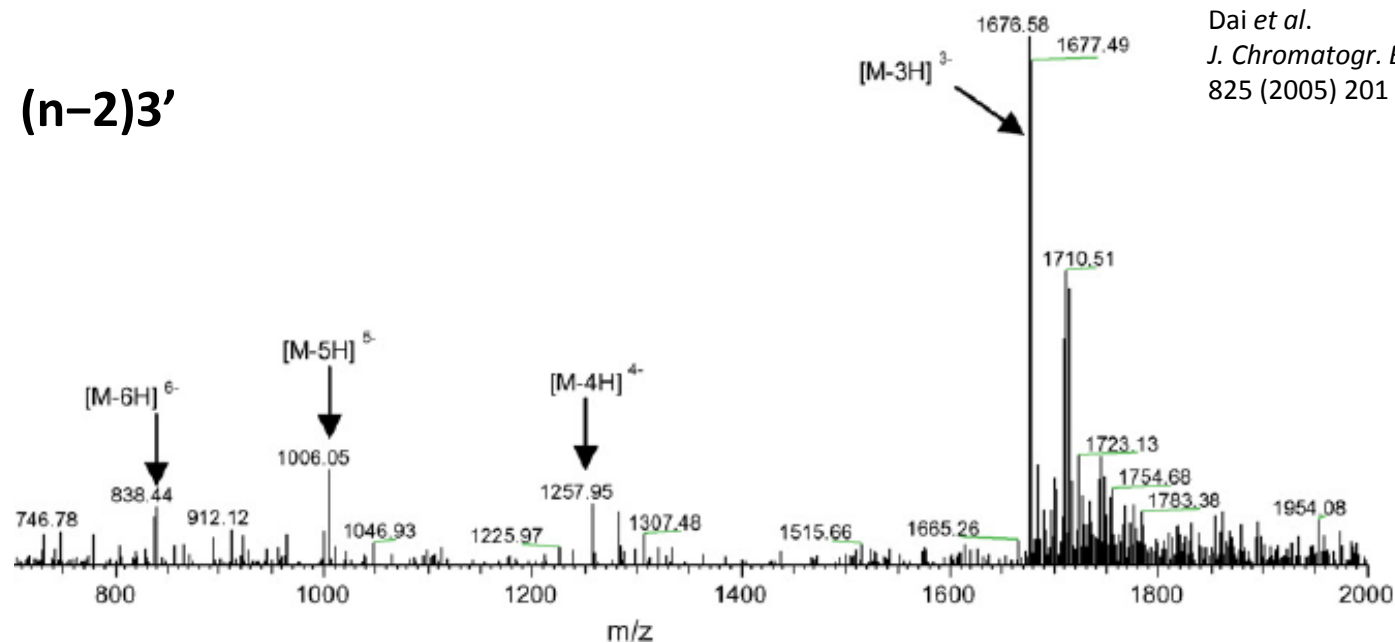
Human metabolites of G3139

G3139 5'TCTCCCAGCGTGCGCCAT'3

(n-1)3'



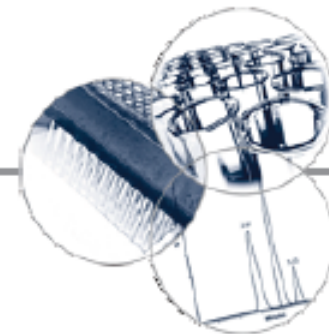
(n-2)3'



Dai et al.
J. Chromatogr. B
825 (2005) 201

Future perspective: asRNA

- Current 4 ng/ml, must and will be improved
 - UPLC
 - sensitive triple quadrupole MS (Xevo TQS, API 5500, Agilent 6490)
 - (nano-)UPLC and chip technology
 - make LLQ's in the range of low pg/ml potentially possible



Bioanalytical LC–MS of therapeutic oligonucleotides

Therapeutic oligonucleotides (OGNTs) are important biopharmaceutical drugs for the future, due to their ability to selectively reduce or knockout the expression of target genes. For the development of OGNTs, reliable and relatively high-throughput bioanalytical methods are required to perform the quantitative bioanalysis of OGNTs and their metabolites in biological fluids (e.g., plasma, urine and tissue). Although immunoaffinity methods, especially ELISA, are currently widely applied for this purpose, the potential of LC–MS in OGNT analysis is under investigation. Owing to its inherent ability to monitor the individual target OGNTs as well as their metabolites, LC–MS is now evolving into the method-of-choice for the bioanalysis of OGNTs. In this paper, the state-of-the-art of bioanalytical LC–MS of OGNTs and their metabolites in biological fluids is critically reviewed and its advantages and limitations highlighted. Finally, the future perspective of bioanalytical LC–MS, that is, lower detection levels and potential generic LC–MS methodology, is discussed.

At present, short-chain oligomers are applied and under investigation as **oligonucleotide therapeutics** (OGNTs), typically between 15 and 50 nucleotide units long. These OGNTs are built to interfere with the processing of genetic information by acting on DNA or RNA [1]. Unlike many other drugs, which target the functioning of proteins by interfering with their receptor site, OGNTs target the gene directly or at the mRNA-expression stage and thereby interfere with the production of

Another type of OGNTs is based on a naturally occurring gene silencing mechanism called RNA interference (RNAi), where short dsRNAs knock down gene expression in cells [2–5]. The synthetic 19- to 25-base pair (bp) dsRNAs used as OGNTs of this type are called **short interfering RNA** (siRNAs). The RNAi process is a multistep process: the double-stranded siRNAs are incorporated into the RNA-induced silencing complex (RISC). The central defining component of the RISC is formed by an Argonaute

William D van Dongen^{†1} & Wilfried MA Niessen^{2,3}

¹Proxy Laboratories, Archimedesweg 25, 2333 CM Leiden, The Netherlands
²Hyphen MassSpec, Liden, The Netherlands

³VU University Amsterdam, Faculty of Sciences, Section of BioMolecular Analysis, Amsterdam, The Netherlands

[†]Author for correspondence:

Tel.: +31 610 010 515

E-mail: wdvandongen@gmail.com

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therapeutics