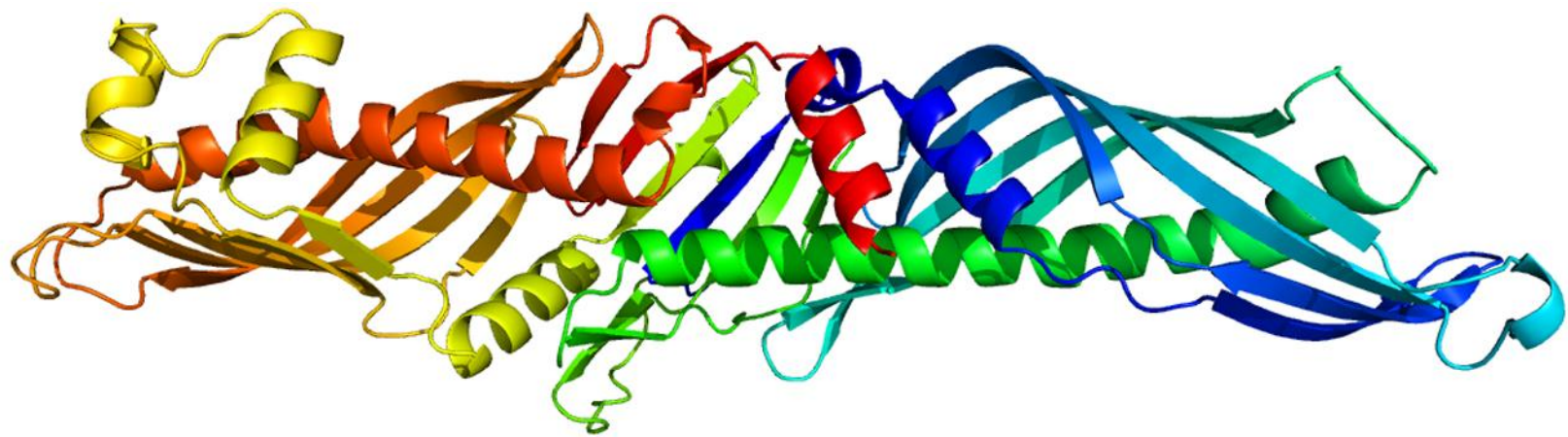


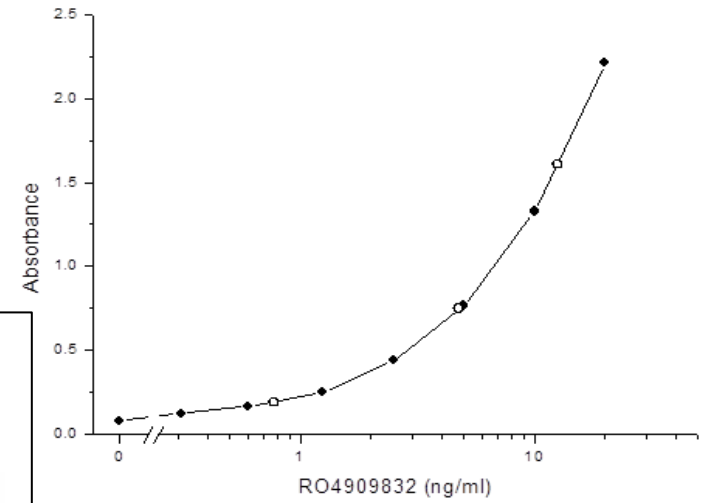
Comparison of quantitative assays for a large protein: ELISA vs. LC-MS/MS

A. Guenzi, G. Fischer, N. Justies, B. Lausecker

F. Hoffmann - La Roche Ltd., Basel, Switzerland



ELISA



ELISA (proteins and peptides)

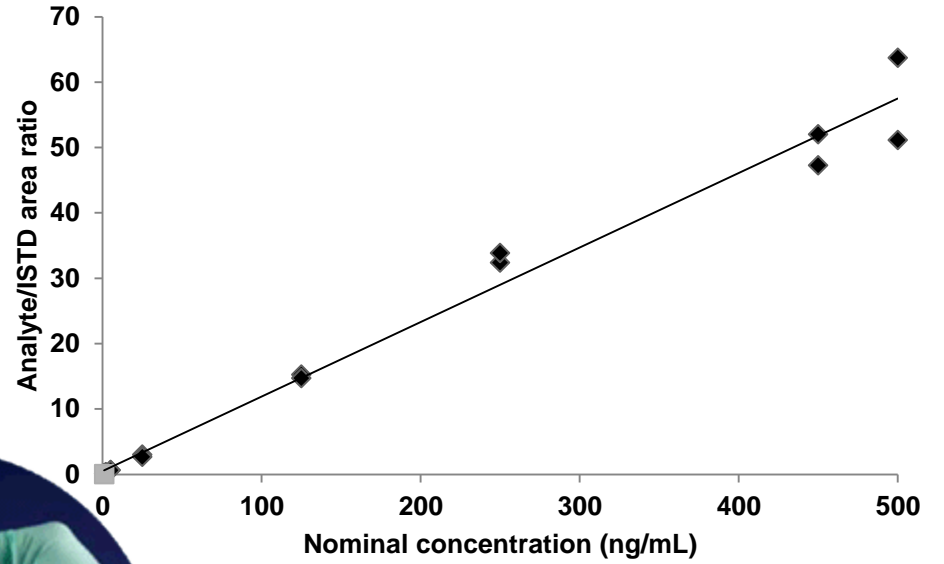
- **ADVANTAGES:**

- Highly sensitive
- Quick and easy to perform
- Inexpensive instrumentation and material
- If anti-idiotypic Abs used, only 'active' is detected

- **DISADVANTAGES:**

- Need for very specific antibodies: animal immunization, selection and purification of reagents
 - For small peptides specific Abs may be difficult to find
- Small differences in analyte sequence may require development of new reagents
- Batch-to-batch variability of reagents
- Each ELISA method needs multiple optimizations
- Each species needs optimization (cyno≠rat≠man≠...)
- Each matrix needs optimization (plasma≠serum≠...)
- Enzyme/substrate reaction is short term so read as soon as possible

LC-MS/MS



LC-MS/MS (proteins and peptides)

- **ADVANTAGES:**

- Highly sensitive
- Quick and easy to perform
- Multi-analyte assays possible
- Species-independent
- Only 'total' detectable

- **DISADVANTAGES:**

- Multiply charged ions may reduce sensitivity
- Need for digestion (proteins) to 'simplify' to peptides
- Is a signature peptide formed during digestion?
- Lipophilicity of peptides
- Wide range of concentrations possible (from pg/mL to µg/mL)
- Mass range of MS instruments

**ELISA and LC-MS/MS data complement each other,
they are not in competition**

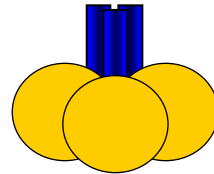
CASE STUDY:

Analysis of a protein (300 kDa including lipidation)

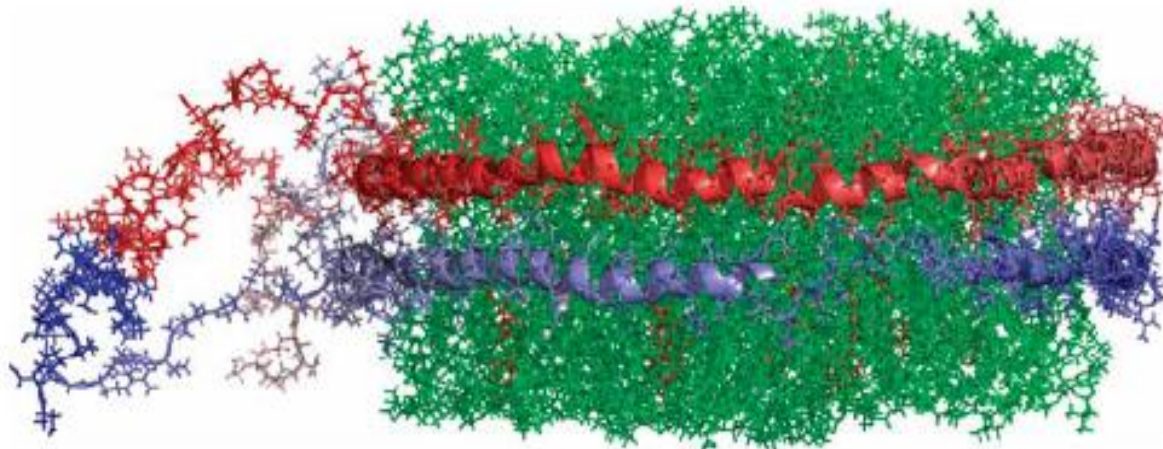
by ELISA and LC-MS/MS

The protein

A complex, trimeric fusion protein (150 kDa)

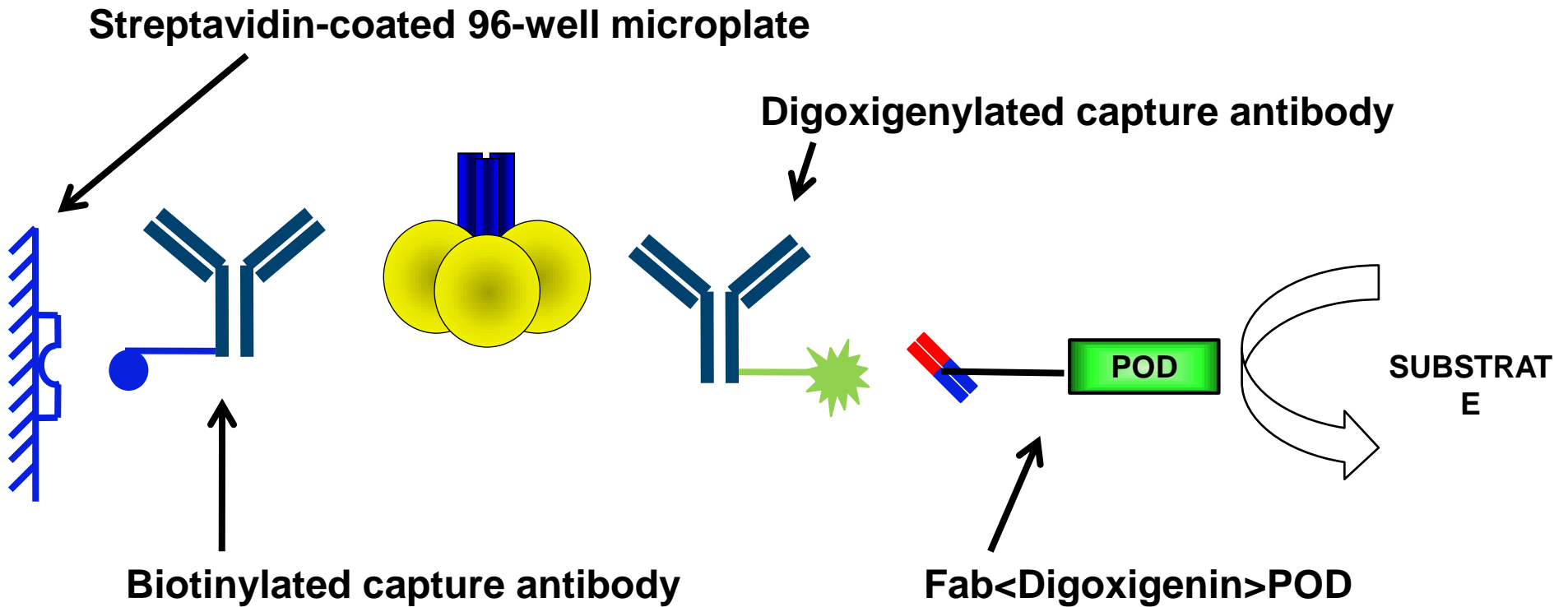


containing phospholipids



Molecular weight: 300 kDa

ELISA (PK) assay



Validation results of the ELISA cynomolgus plasma

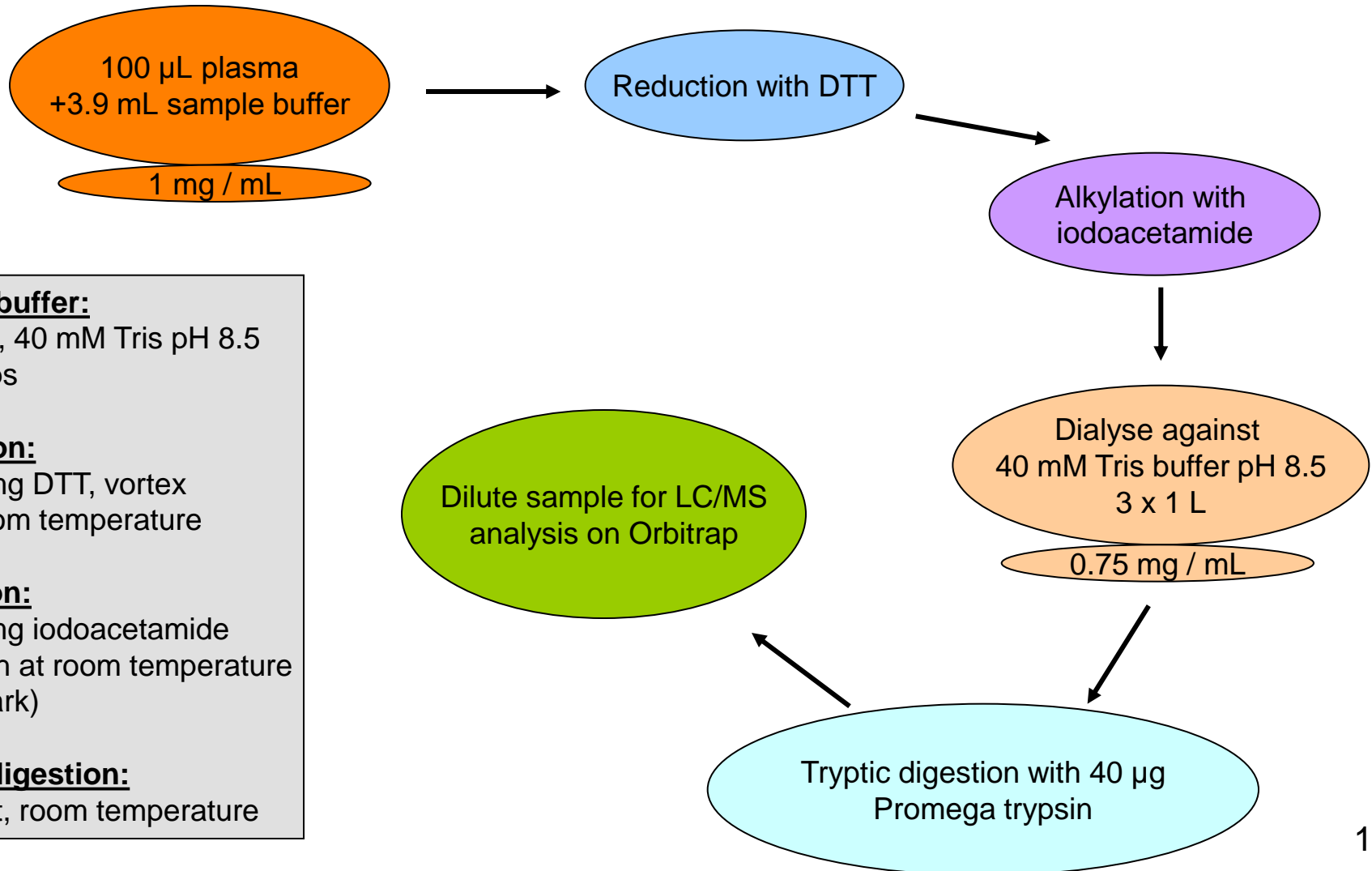
- **Minimum required dilution (MRD):** 1%
- **Validated range (in 100% plasma):** 1.02 to 65.0 µg/mL
- **Validated range (in assay buffer):** 10.2 to 650 ng/mL
- **Precision and accuracy (intra-assay):** <9.2% and 89.9 – 101.3%
- **Precision and accuracy (inter-assay):** <4.7% and 99.9 – 111.2%
- **Selectivity, specificity:** within Guidance criteria
- **Dilution linearity and parallelism:** no hook effect up to 6.50 mg/mL
- **Assay signal stability:** readout immediately after addition of SDS

- **During application (routine analysis of studies) the ELISA-measured concentrations did not fit with other pharmacodynamic observations**
- **The possibility of exploiting for quantitative purposes the results obtained by proteomics studies were considered**



An orthogonal analytical method (LC-MS/MS) able to confirm the ELISA data was developed

Starting point for LC-MS assay: Proteomics qualitative procedure



Sample buffer:

8 M urea, 40 mM Tris pH 8.5
2% Chaps

Reduction:

Add 15 mg DTT, vortex
1 h at room temperature

Alkylation:

Add 60 mg iodoacetamide
vortex 1 h at room temperature
(in the dark)

Tryptic digestion:

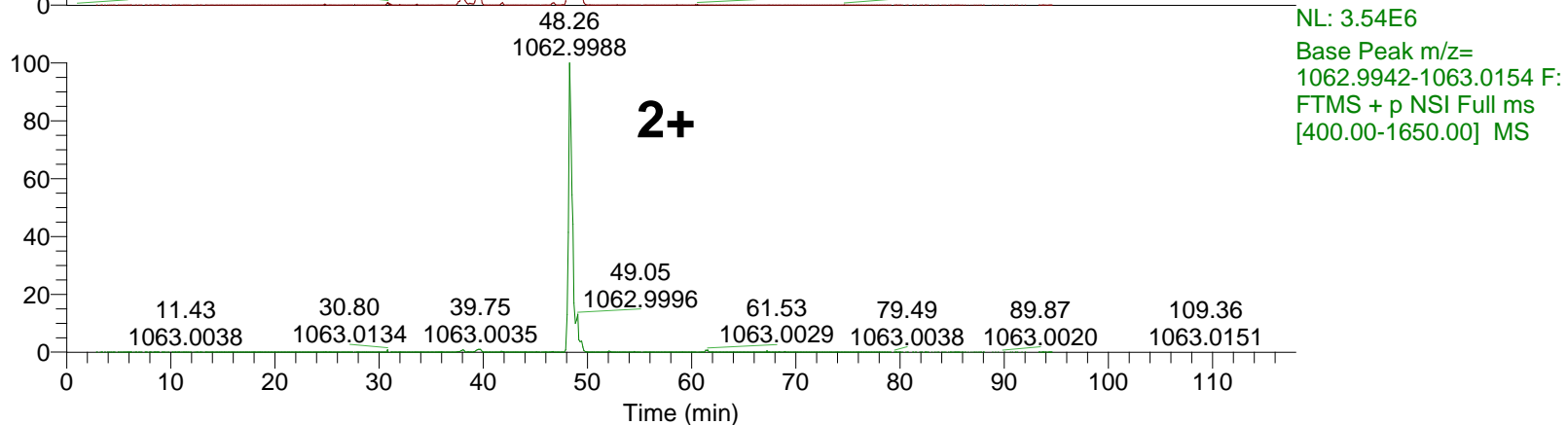
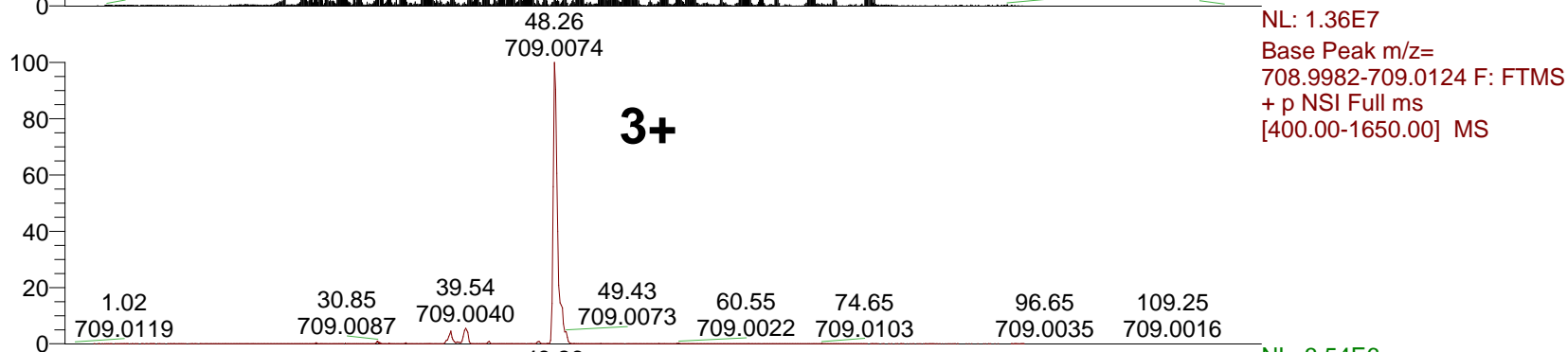
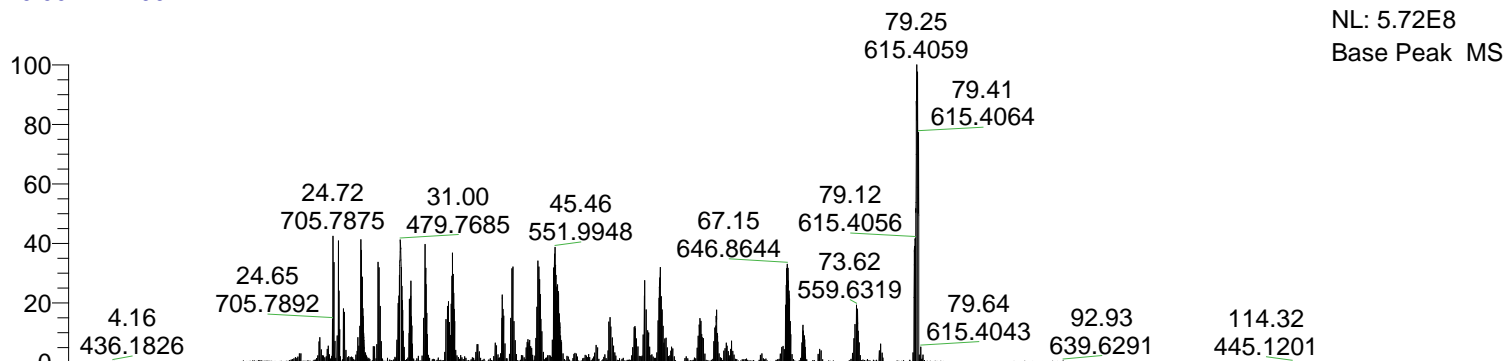
overnight, room temperature

Starting point for LC-MS: Identification of signature peptide

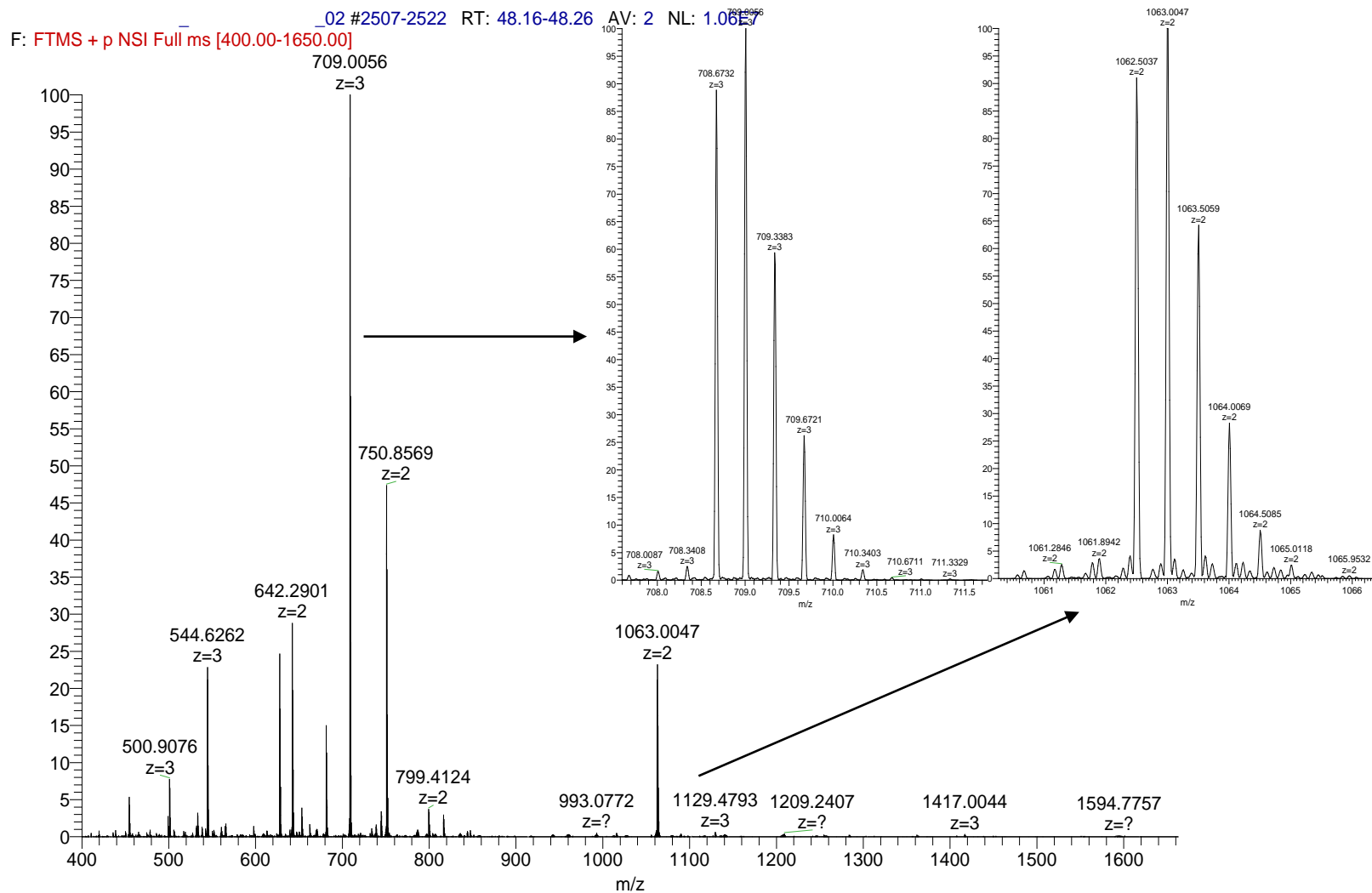
- **Samples containing proteins (e.g., plasma) are digested to peptides with trypsin. Peptides are selected as surrogates of the protein by the following criteria:**
 - **The fragmentations (transitions) are unique for one single protein**
 - **The peptide contains no Cys, Met or other commonly modified residues**
 - **The peptides lie in the mass range 800 – 2000 MW**

MS analysis of protein in plasma (proteomics)

RT: 0.00 - 117.99



Full ion scan of the signature peptide

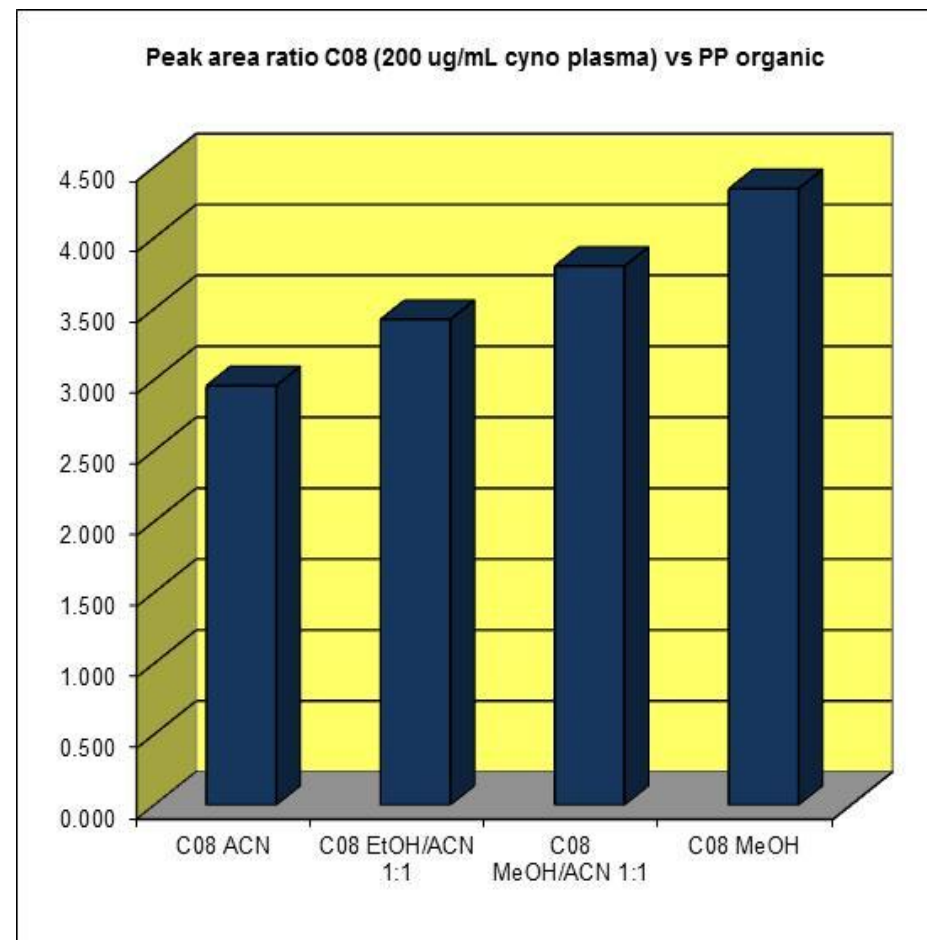
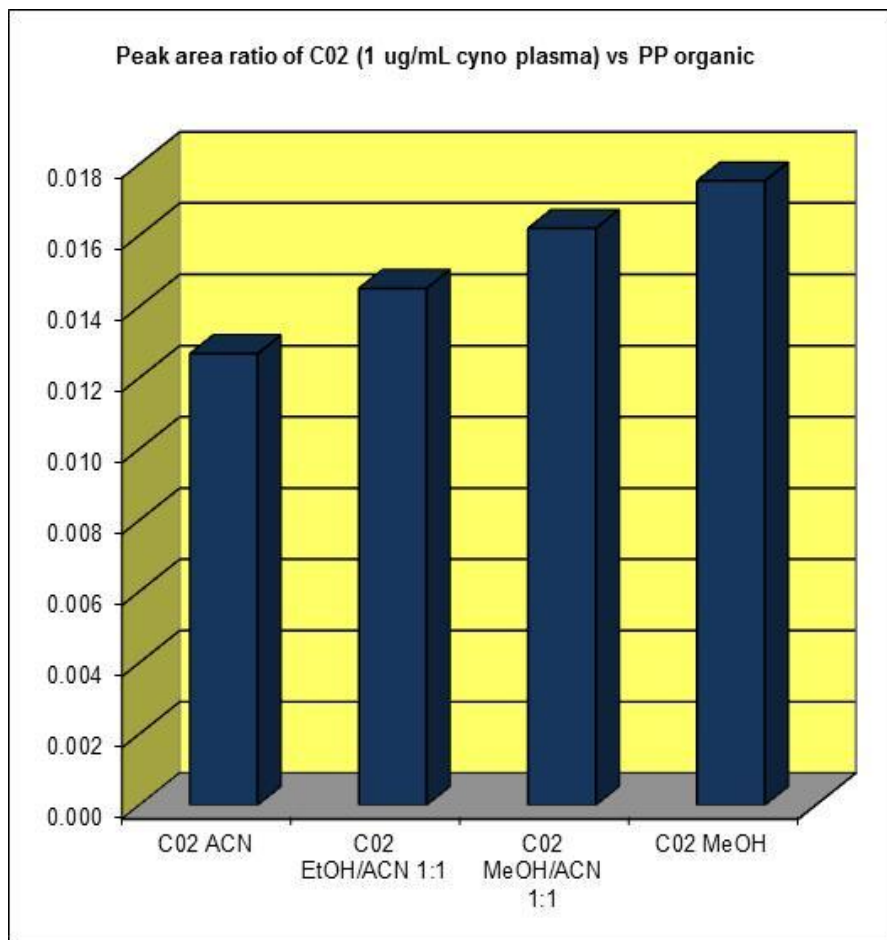


Multidimensional optimization of the LC-MS/MS assay

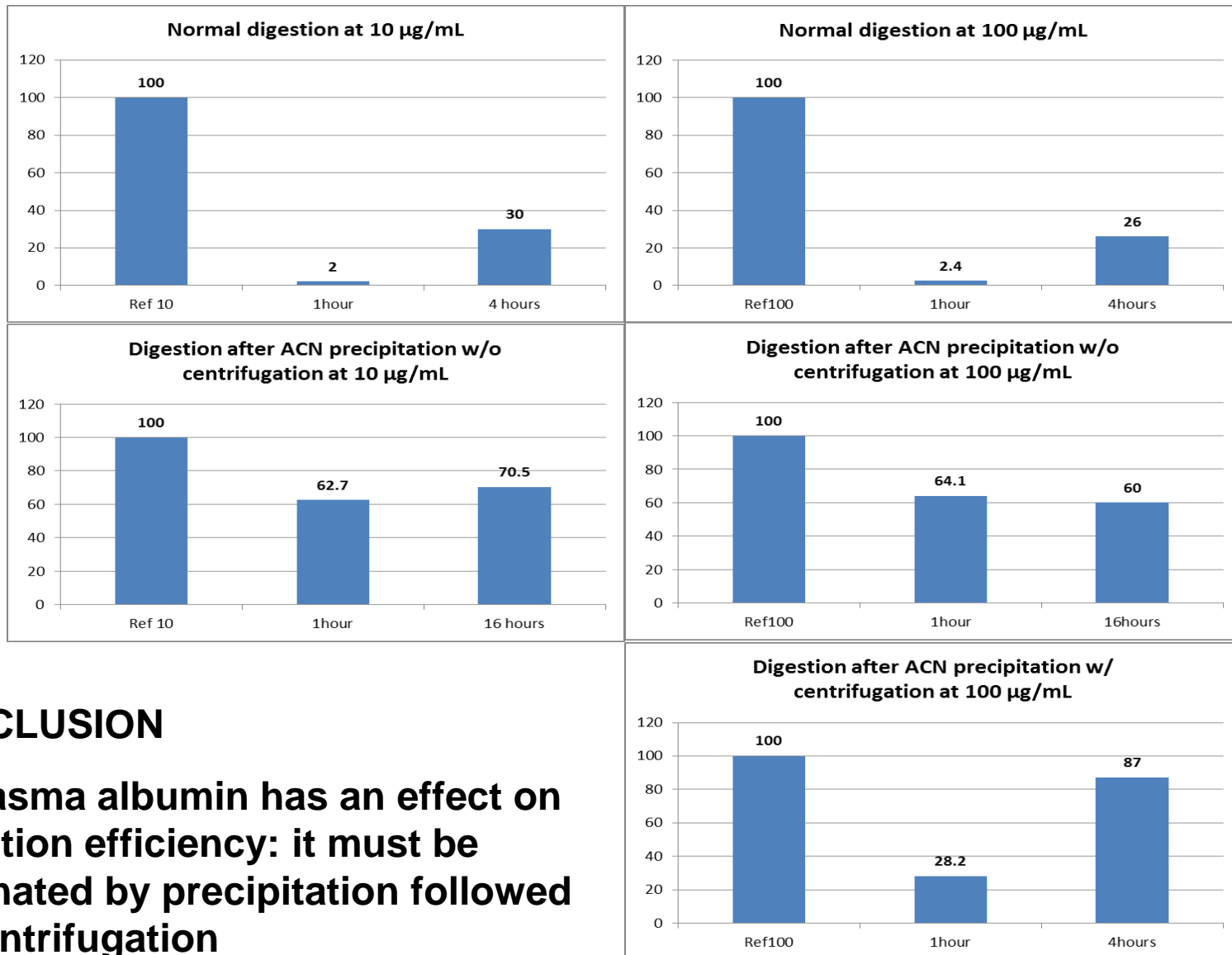
Optimization of the following parameters

- Volume of plasma
- Volume of buffer
- Trypsin concentration
- Protease-protein (mainly albumin) ratio
- Precipitating solvent
- Duration of incubation
- Chromatographic conditions
- Addition of internal standard

Example: choice of precipitating solvent in the preliminary step



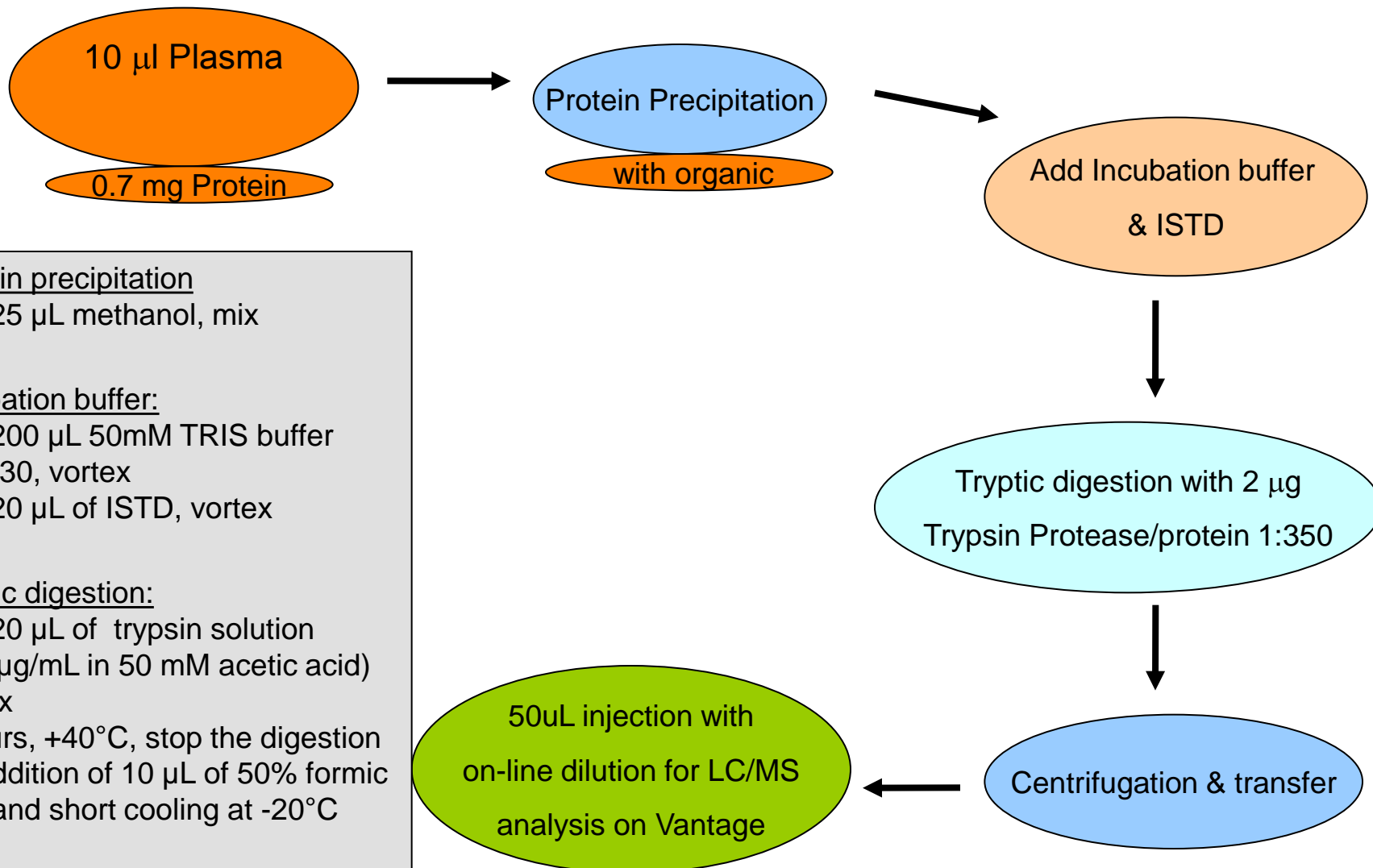
Example: protein precipitation parameters



CONCLUSION

⇒ Plasma albumin has an effect on digestion efficiency: it must be eliminated by precipitation followed by centrifugation

Cyno plasma: final sample preparation



Protein precipitation

Add 25 µL methanol, mix

Incubation buffer:

Add 200 µL 50mM TRIS buffer

pH 8.30, vortex

Add 20 µL of ISTD, vortex

Tryptic digestion:

Add 20 µL of trypsin solution

(100 µg/mL in 50 mM acetic acid)

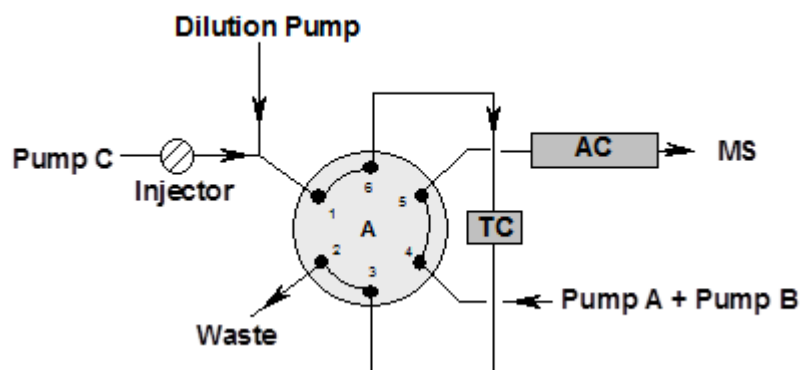
Vortex

2 hours, +40°C, stop the digestion

via addition of 10 µL of 50% formic acid and short cooling at -20°C

LC-MS/MS of the signature peptide

- The traditional approach in our group (column-switching with on-line solid-phase extraction) was followed



- 50 μL injection volume (1.8 μL plasma-equivalents)
- Pump C to dilution pump flow-rate ratio 1:6.6
- Gradient elution with acetonitrile-water-formic acid on the analytical column (Supelco Ascentis Express C18, 2.0 (i.d.) x 50 mm)
- Detection on a Thermo TSQ Vantage EM triple quad with H-ESI source

Validation results of the LC-MS/MS assay cynomolgus plasma

- Validated range (plasma): 1.00 to 200 µg/mL
- Precision and accuracy (intra-assay): <7.9% and 100.0 – 106.7%
- Precision and accuracy (inter-assay): <7.9% and 92.5 – 105.6%
- Selectivity, specificity: within Guidance criteria
- Dilution linearity: validated up to 1:100
- Matrix effect: compensated by SLIS, ca. 0.92
- Extraction recovery: between 93.5 and 97.8 (SLIS)

It does not cover digestion!



Chromatograms



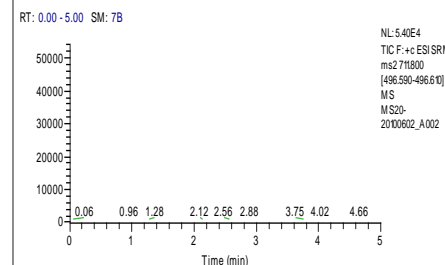
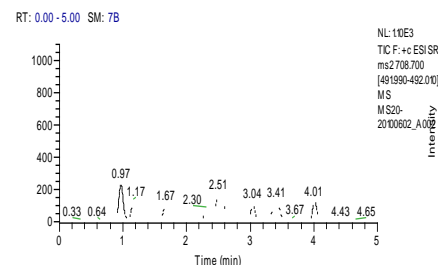
D:\Xcalibur...MS20-2\

008

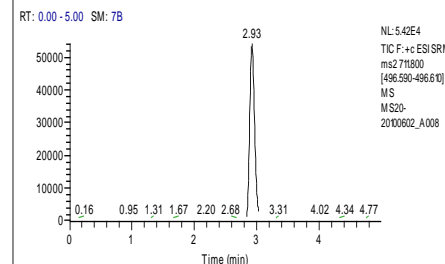
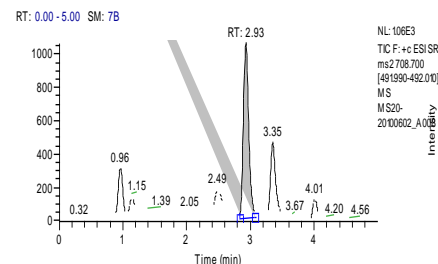
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C01

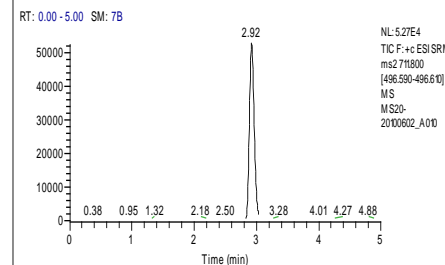
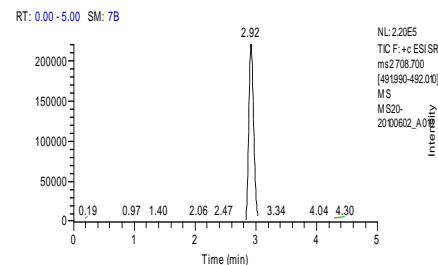
Blank Cyno plasma



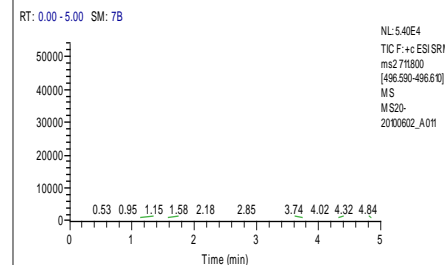
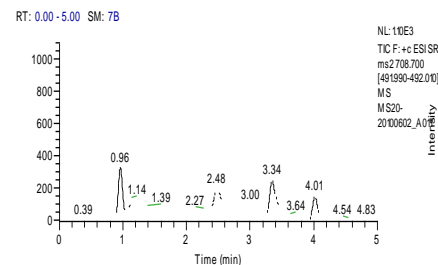
LLOQ = 1 µg/mL



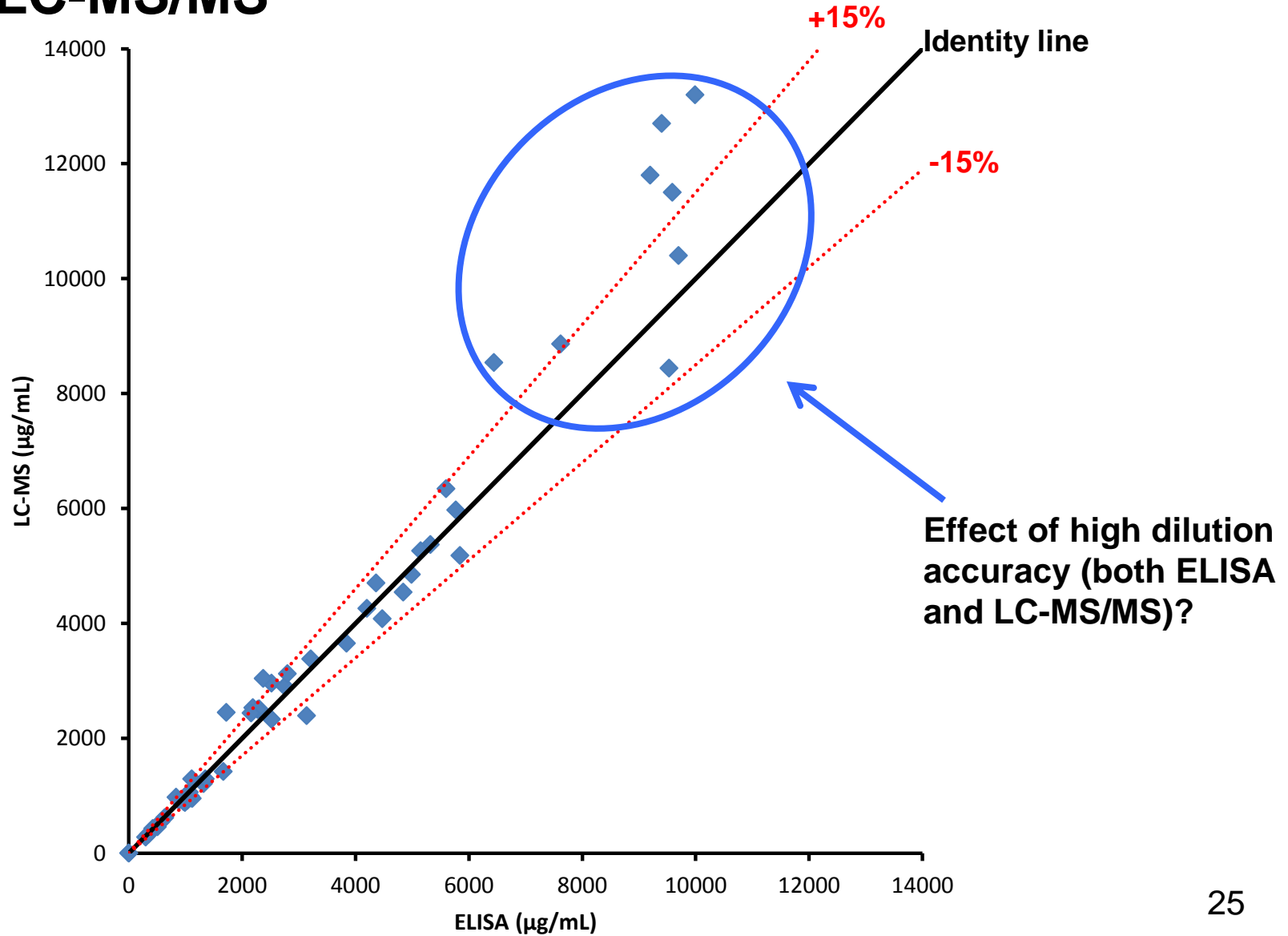
ULOQ = 200 µg/mL



**Blank after ULOQ
(carry-over test)**



Cross-validation results ELISA vs. LC-MS/MS



Acknowledgements

Non-Clinical Safety, Roche Basel

- **Katrin Schliemann**

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Pharma Research, Roche Penzberg

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