

# Developing LC-MS/MS methods for quantifying mAbs: Transitioning from pre-clinical to clinical matrices.

LGC

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# Overview



- Existing methods for mAb extraction and analysis
- Improvements to existing methods
- Potential patent issues
- Alternative enzymes and surrogate peptides
- Moving into human

# Methods for extracting mAb's

- Immunoprecipitation:
  - Protein A/G extraction
  - Anti-human IgG extraction
  - Anti-idiotypic
  - Target protein
- Pro's – selective extraction methods
- Con's – expensive reagents
  
- Chemical extraction
  - Protein precipitation
- Pro's – cheap
- Con's – non-specific

# Protein A/G selectivity issues

Species IgG	Weak	Strong	Useful for mAb extraction?
Human		✓	Not really
Cyno		✓	Not really
Rat	✓		Yes
Mouse	✓		Yes
Rabbit		✓	Not really

- Protein A will extract IgG from monkey and human serum
- An issue with capacity...
  - IgG = ~ 15 mg/mL,
  - 25  $\mu$ L of serum = ~ 375  $\mu$ g of IgG
  - Protein A magnetic beads will capture 8  $\mu$ g protein per mg of beads
  - Requires 46 mg beads per 25  $\mu$ L....
  - Supplied at 20mg/mL = ~2.5 mL of beads per sample

# Protein precipitation based method



- The pellet digestion method

- Bioanalysis (2012) 4(1), 17–28

Pellet digestion: a simple and efficient sample preparation technique for LC–MS/MS quantification of large therapeutic proteins in plasma

Zheng Ouyang<sup>1</sup>, Michael T Furlong<sup>2</sup>, Steven Wu<sup>1</sup>, Bogdan Slecza<sup>1</sup>, James Tamura<sup>3</sup>, Haiqing Wang<sup>4</sup>, Suzanne Suchard<sup>5</sup>, Anish Suri<sup>5</sup>, Timothy Olah<sup>1</sup>, Adrienne Tymiak<sup>1</sup> & Mohammed Jemal<sup>\*1</sup>

<sup>1</sup>Bristol-Myers Squibb, Research & Development, Bioanalytical & Discovery Analytical Sciences, Route 206 & Province Line Road, Princeton, NJ 08543, USA

- Precipitate all large proteins and digest pellet

- **Discard** supernatant from protein crash

- Referenced a paper from my PhD:

- Analysis of IGF-I after GH administration to athletes
- **Keep** supernatant from protein crash

<sup>16</sup> Kay RG, Barton C, Velloso CP *et al.* High-throughput ultra-high-performance liquid chromatography–tandem mass spectrometry quantitation of insulin-like growth factor-I and leucine-rich a-2-glycoprotein in serum as biomarkers of recombinant human growth hormone administration. *Rapid Commun. Mass Spectrom.* 23(19), 3173–3182 (2009).



# Pellet digestion method

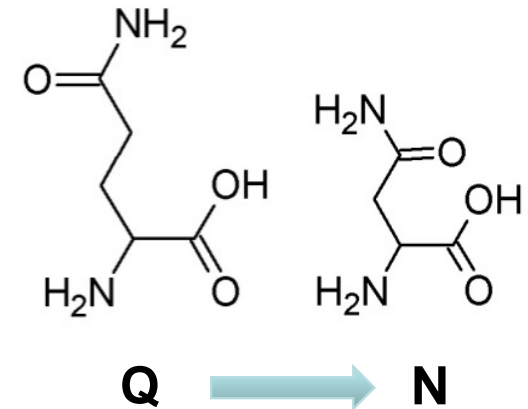
- 25  $\mu\text{L}$  of plasma (~2.5 mg of protein)
- Add 75  $\mu\text{L}$  of methanol (ACN was better, but gave tight pellet)
- Mix and centrifuge
- **Discard** supernatant
- Digest pellet (Sigma trypsin, 8mg/mL)
- Analyse by LC-MS/MS (2.6  $\mu\text{g}/\text{mL}$  LLOQ using 25  $\mu\text{L}$ )
  
- Can we improve on this?
  
- Can we reduce the albumin level whilst improving mAb recovery?
  - Monitor an Albumin peptide along with a mAb peptide

# Albumin peptide (rodent Vs. primate)



## Rodent albumin mascot hit

#	b	b <sup>++</sup>	b <sup>*</sup>	b <sup>***</sup>	b <sup>0</sup>	b <sup>0++</sup>	Seq.	y	y <sup>++</sup>	y <sup>*</sup>	y <sup>***</sup>	y <sup>0</sup>	y <sup>0++</sup>	#
1	114.0913	57.5493					L							10
2	<b>213.1598</b>	107.0835					V	1036.5310	518.7691	1019.5044	510.2558	1018.5204	509.7638	9
3	<b>341.2183</b>	171.1128	324.1918	162.5995			Q	<b>937.4625</b>	469.2349	920.4360	<b>460.7216</b>	919.4520	<b>460.2296</b>	8
4	470.2609	235.6341	453.2344	227.1208	452.2504	226.6288	E	<b>809.4040</b>	405.2056	792.3774	396.6923	791.3934	396.2003	7
5	569.3293	285.1683	552.3028	276.6550	551.3188	276.1630	V	<b>680.3614</b>	<b>340.6843</b>	663.3348	332.1710	662.3508	331.6790	6
6	670.3770	335.6921	653.3505	327.1789	652.3665	326.6869	T	<b>581.2930</b>	291.1501	564.2664	282.6368	563.2824	282.1448	5
7	785.4040	393.2056	768.3774	384.6923	767.3934	384.2003	D	<b>480.2453</b>	<b>240.6263</b>	463.2187	232.1130	462.2347	231.6210	4
8	932.4724	466.7398	915.4458	458.2266	914.4618	457.7345	F	<b>365.2183</b>	183.1128	348.1918	174.5995			3
9	1003.5095	502.2584	986.4829	493.7451	985.4989	493.2531	A	218.1499	109.5786	201.1234	101.0653			2
10							K	147.1128	74.0600	130.0863	65.5468			1



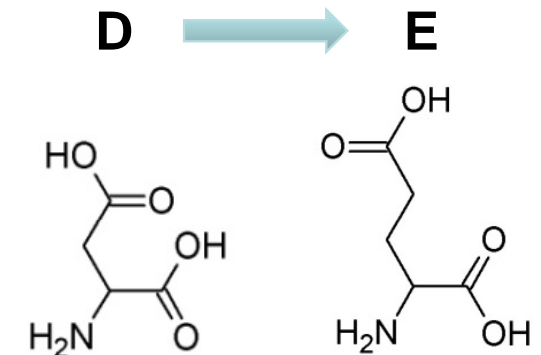
**LVQEVTDFAK Q-N=+14 Da 575.3 / 809.4**

**Generic cross species albumin peptide transition = 575.3 / 937.5**

**LVNEVTDFAK D-E=-14 Da 575.3 / 823.4**

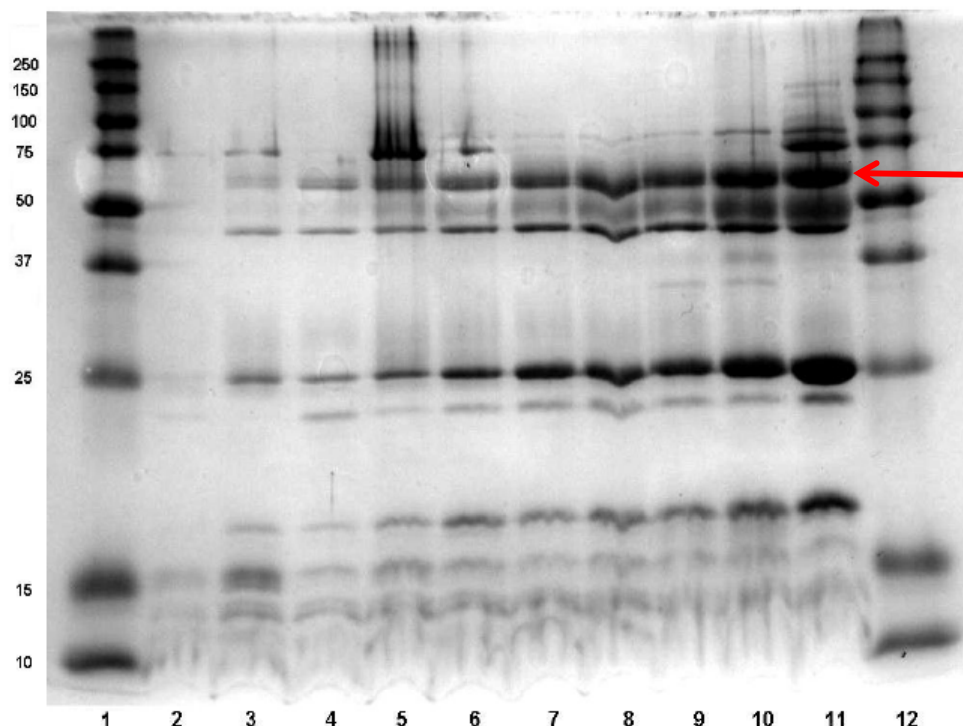
## Primate albumin mascot hit

#	b	b <sup>++</sup>	b <sup>*</sup>	b <sup>***</sup>	b <sup>0</sup>	b <sup>0++</sup>	Seq.	y	y <sup>++</sup>	y <sup>*</sup>	y <sup>***</sup>	y <sup>0</sup>	y <sup>0++</sup>	#
1	114.0913	57.5493					L							10
2	<b>213.1598</b>	107.0835					V	1036.5310	518.7691	1019.5044	510.2558	1018.5204	509.7638	9
3	327.2027	164.1050	310.1761	155.5917			N	<b>937.4625</b>	<b>469.2349</b>	920.4360	460.7216	919.4520	460.2296	8
4	<b>456.2453</b>	228.6263	439.2187	220.1130	438.2347	219.6210	E	<b>823.4196</b>	412.2134	<b>806.3931</b>	403.7002	805.4090	403.2082	7
5	<b>555.3137</b>	278.1605	538.2871	269.6472	537.3031	269.1552	V	<b>694.3770</b>	347.6921	677.3505	339.1789	676.3665	338.6869	6
6	656.3614	328.6843	639.3348	320.1710	638.3508	319.6790	T	<b>595.3086</b>	<b>298.1579</b>	578.2821	289.6447	577.2980	289.1527	5
7	785.4040	393.2056	768.3774	384.6923	767.3934	384.2003	E	<b>494.2609</b>	247.6341	477.2344	239.1208	476.2504	238.6288	4
8	932.4724	466.7398	915.4458	458.2266	914.4618	457.7345	F	<b>365.2183</b>	183.1128	348.1918	174.5995			3
9	1003.5095	502.2584	986.4829	<b>493.7451</b>	985.4989	493.2531	A	<b>218.1499</b>	109.5786	201.1234	101.0653			2
10							K	147.1128	74.0600	130.0863	65.5468			1



# Protein crash solvent selection

- Adding water to serum prior to crash gave better recovery of proteins
- Take supernatant after protein crash and run on SDS-PAGE gel



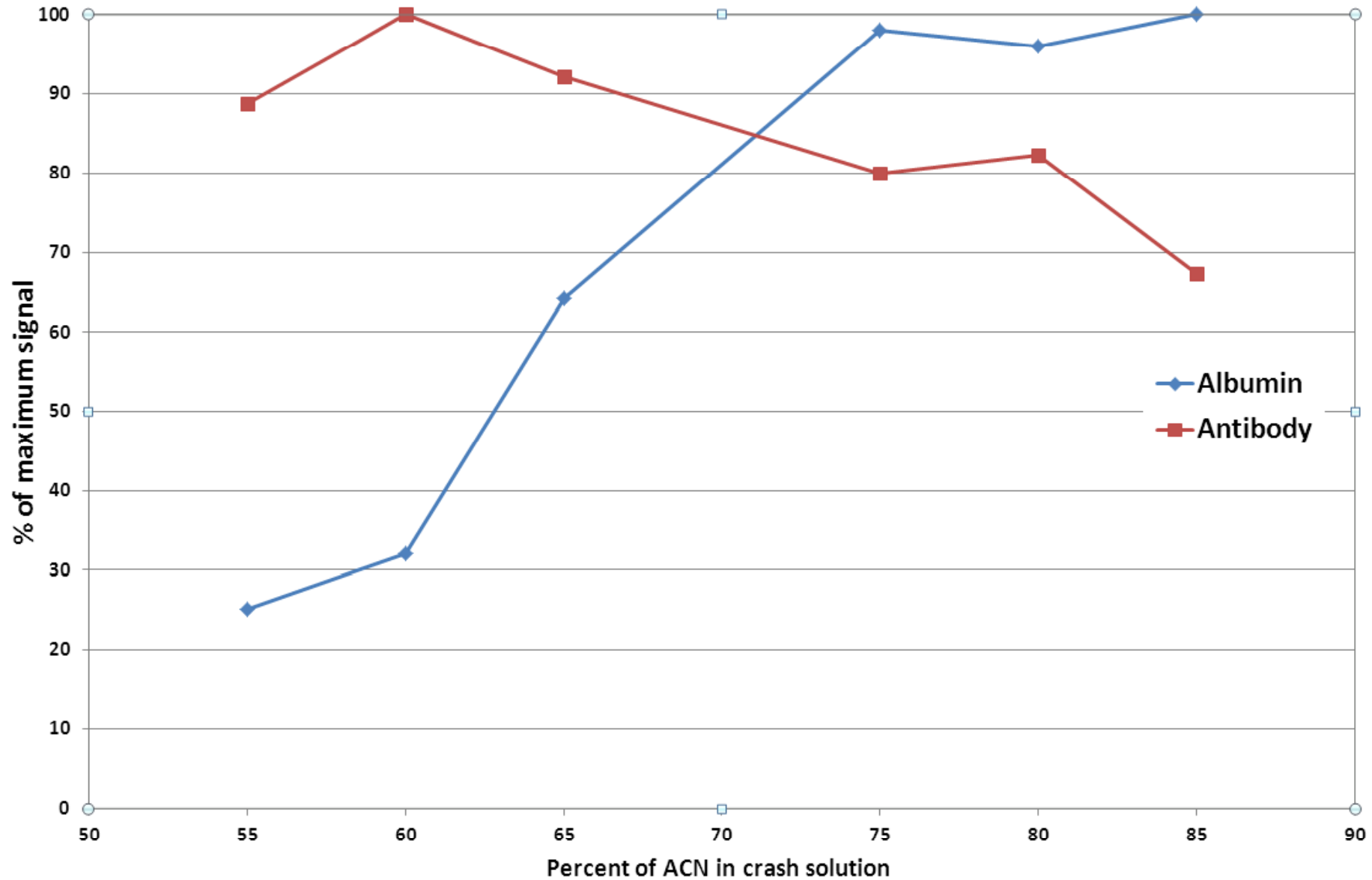
Albumin

- 1= markers
- 2= no dilution
- 3= 1:0.5      serum:water
- 4= 1:1        serum:water
- 5= 1:1.5      serum:water
- 6= 1:2        serum:water
- 7= 1:2.5      serum:water
- 8= 1:3        serum:water
- 9= 1:3.5      serum:water
- 10=1:4        serum:water
- 11=1:9        serum:water
- 12= markers

Increased dilution of serum, same ACN ratio:diluted serum (1.5 volumes ACN)



# Optimisation of crash solvent

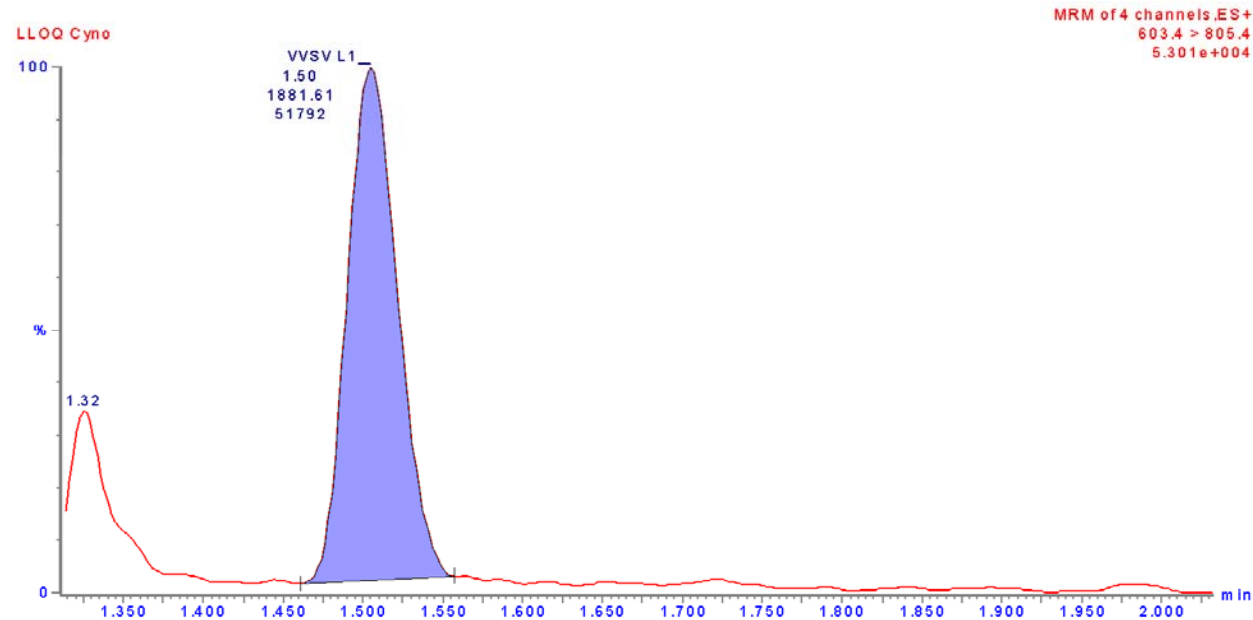
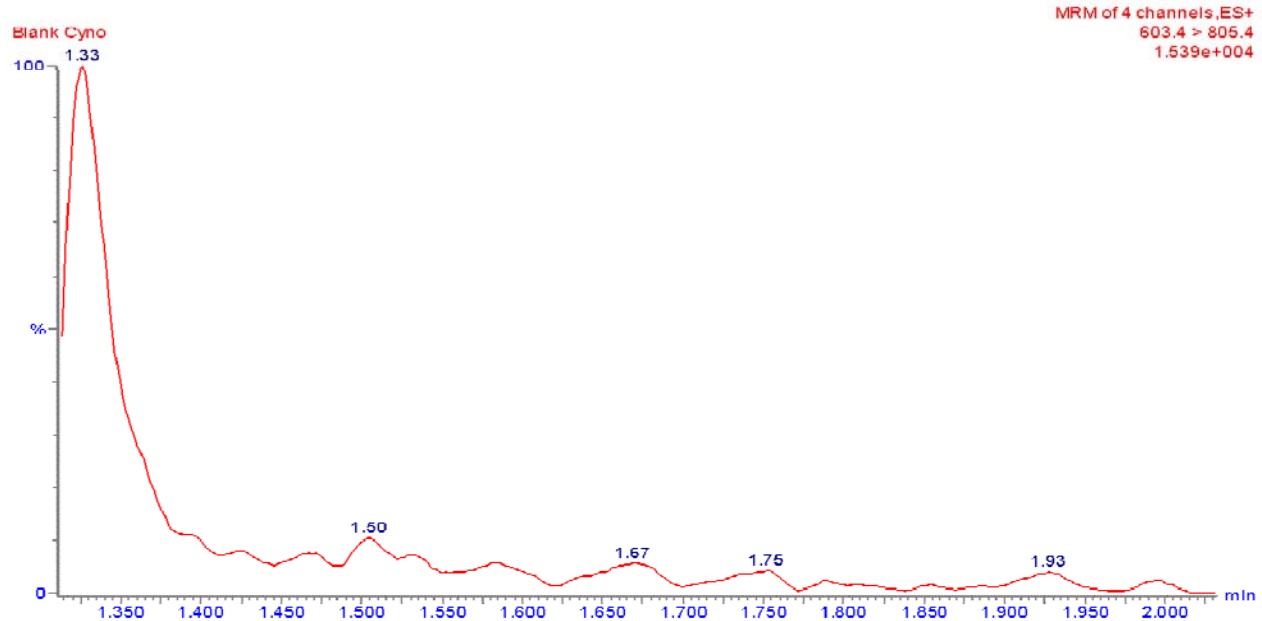




# LC-MS/MS analysis of mAb in preclinical matrices

# mAb in cyno serum at 1 µg/mL

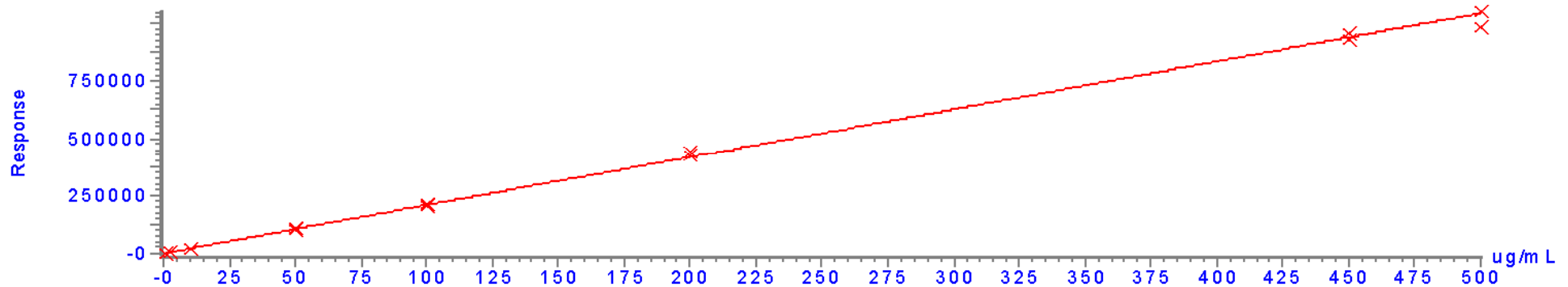
## 10 µL of serum extracted, 200 nL injected



# Cyno extraction results



Correlation coefficient:  $r = 0.998610$ ,  $r^2 = 0.997222$   
Calibration curve:  $2091 \cdot x + -151.192$   
Response type: External Std, Area  
Curve type: Linear, Origin: Exclude, Weighting:  $1/x^2$ , Axis trans: None



QC	Conc	Mean	%CV	%RE
LLOQ	1.00	0.954	3.0	-4.6
LOW	4.00	3.75	4.0	-6.3
MED	40.0	37.7	4.2	-5.8
HIGH	400	375	4.7	-6.3



# Patent issues...

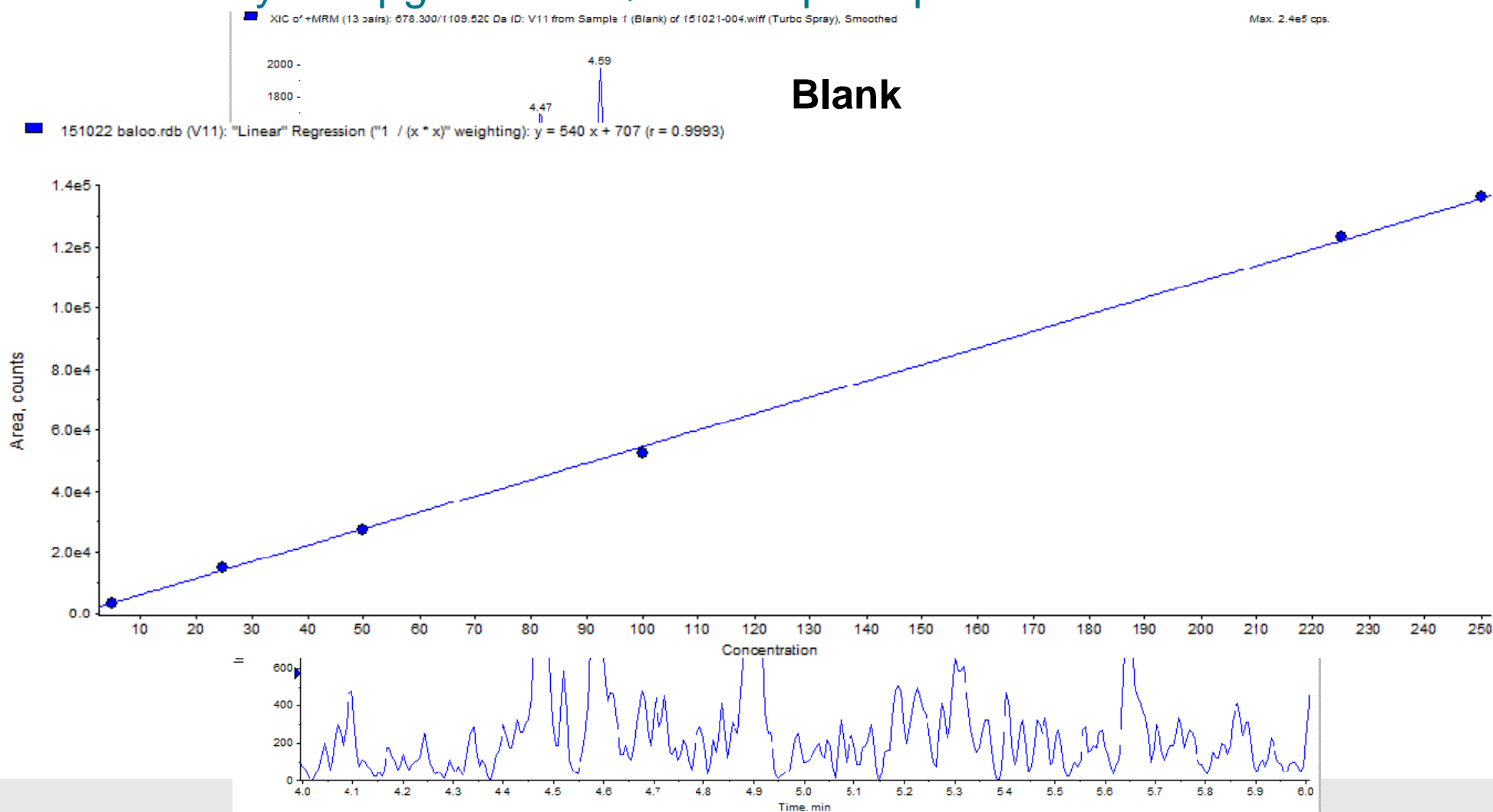
- Patent applied for using human mAb peptides in pre-clinical matrix
  - Filed in May 2012
  - Granted in the US, currently being considered in EU
- Application is specific for 8 framework specific peptides “FSP”

GPSVFPLAPSSK	SEQ ID NO:1
STSGGTAALGCLVK	SEQ ID NO:2
TPEVTCVVVDVSHEDPEVK	SEQ ID NO:3
FNWYVDGVEVHNAK	SEQ ID NO:4
VVSVLTVLHQDWLNGK	SEQ ID NO:5
ALPAPIEK	SEQ ID NO:6
GFYPSDIAVEWESNGQPENNYK	SEQ ID NO:7
TTPPVLDSDGSFFLYSK	SEQ ID NO:8

- Application specifies trypsin as the enzyme
- Glu C may be potentially used instead
  - VKFNWYVDGVE - heavy chain constant region peptide
  - We have achieved 0.25 µg/mL using Protein A extraction on 25 µL plasma
  - Performed in 2013

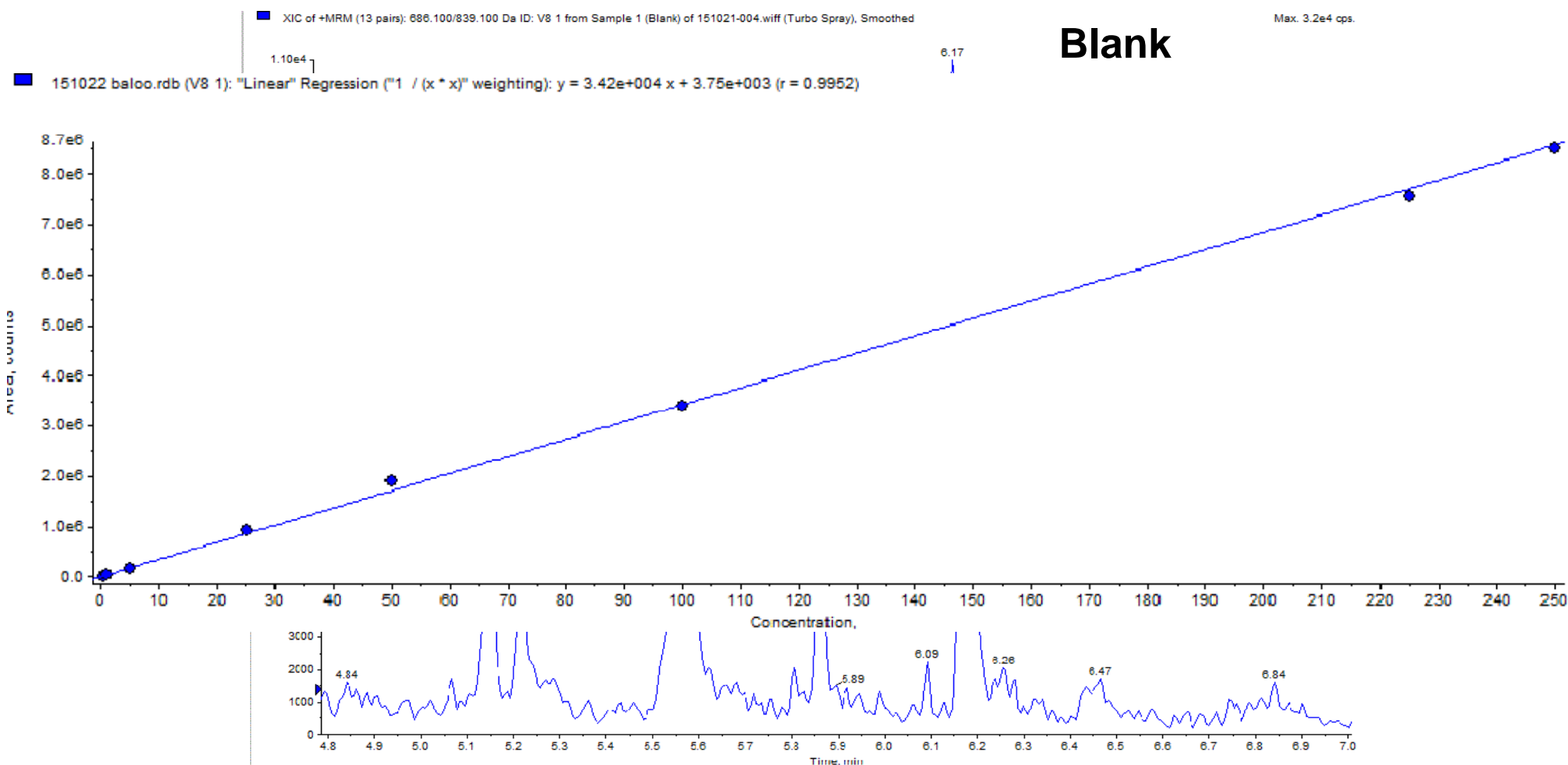
# Protein crash and Glu C peptides

- Generic peptide sequence (VKFNWYVDGVE)
  - Not present in cyno or rodent protein
- Currently at 5 µg/mL LLOQ with 10 µL of plasma



# Protein crash and Glu C peptides

- LLGGPSVFLFPPKPKDTLMISRTPE
- Common with monkey but OK in rodents
- Currently at 0.5 µg/mL LLOQ with 10 µL of plasma





# Human mAb in human serum

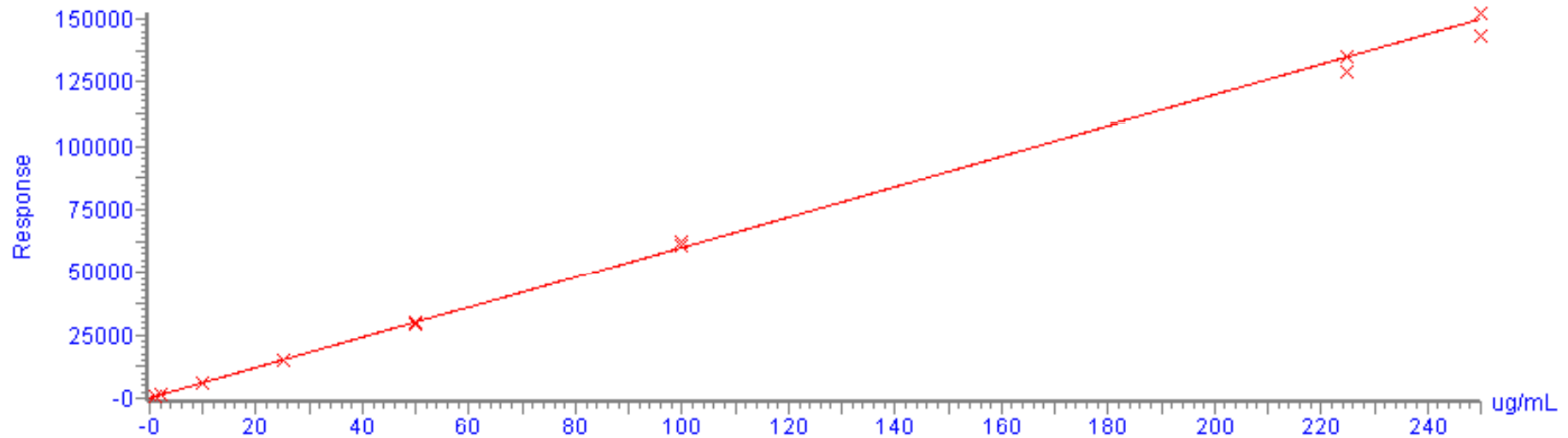




# Method development

- LC-MS/MS approach had to target a peptide from the variable region
- Found 2 that were selective and sensitive
  - 1 that was sufficient for LLOQ in question (1  $\mu\text{g/mL}$ )
- Same solvent assessment as previously performed in pre-clinical
  - Gave same results – 60% ACN in water
- Very quick extraction method development
  - Method was established within 5 working days

# Linearity and precision and accuracy



QC	Conc	Mean	%CV	%RE
LLOQ	1.00	1.10	8.3	9.6
LOW	3.00	3.13	10.1	4.3
MED	20.0	19.6	2.6	-1.9
HIGH	200	182	2.5	-8.9

# Assessment of selectivity



- 20 individual serum samples taken for analysis

Selectivity sample	% of LLOQ	Selectivity sample	% of LLOQ
Sample 1	0	Sample 11	10
Sample 2	0	Sample 12	10
Sample 3	0	Sample 13	30
Sample 4	10	Sample 14	0
Sample 5	10	Sample 15	10
Sample 6	20	Sample 16	0
Sample 7	0	Sample 17	0
Sample 8	40	Sample 18	0
Sample 9	0	Sample 19	0
Sample 10	40	Sample 20	0



# Conclusions

- A modified pellet digestion method was developed
  - Gave good sensitivity for trypsin approach
- Pellet digestion approach is species and matrix agnostic
  - Good for a fast generic mAb extraction method
- Potential patent issues
  - May potentially be circumvented in EU by using Glu C enzyme instead
- Sensitivity is not far off tryptic peptides
  - Still in development..



# Acknowledgements

## Colleagues at LGC:

- Szabolcs Szarka (Sabi)
- Kjetil Hansen (KJ)
- Mark Hows (Mark)

## NovImmune

- Rob Nelson
- Florence Guilhot



# Thank you for listening

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