

# The MULTiple trials of generating a SINGLE data set

## Taking biomarker data through the Clinical Stages

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# Presentation Summary

## Study investigating the utility and comparability of multiplex and singleplex assays

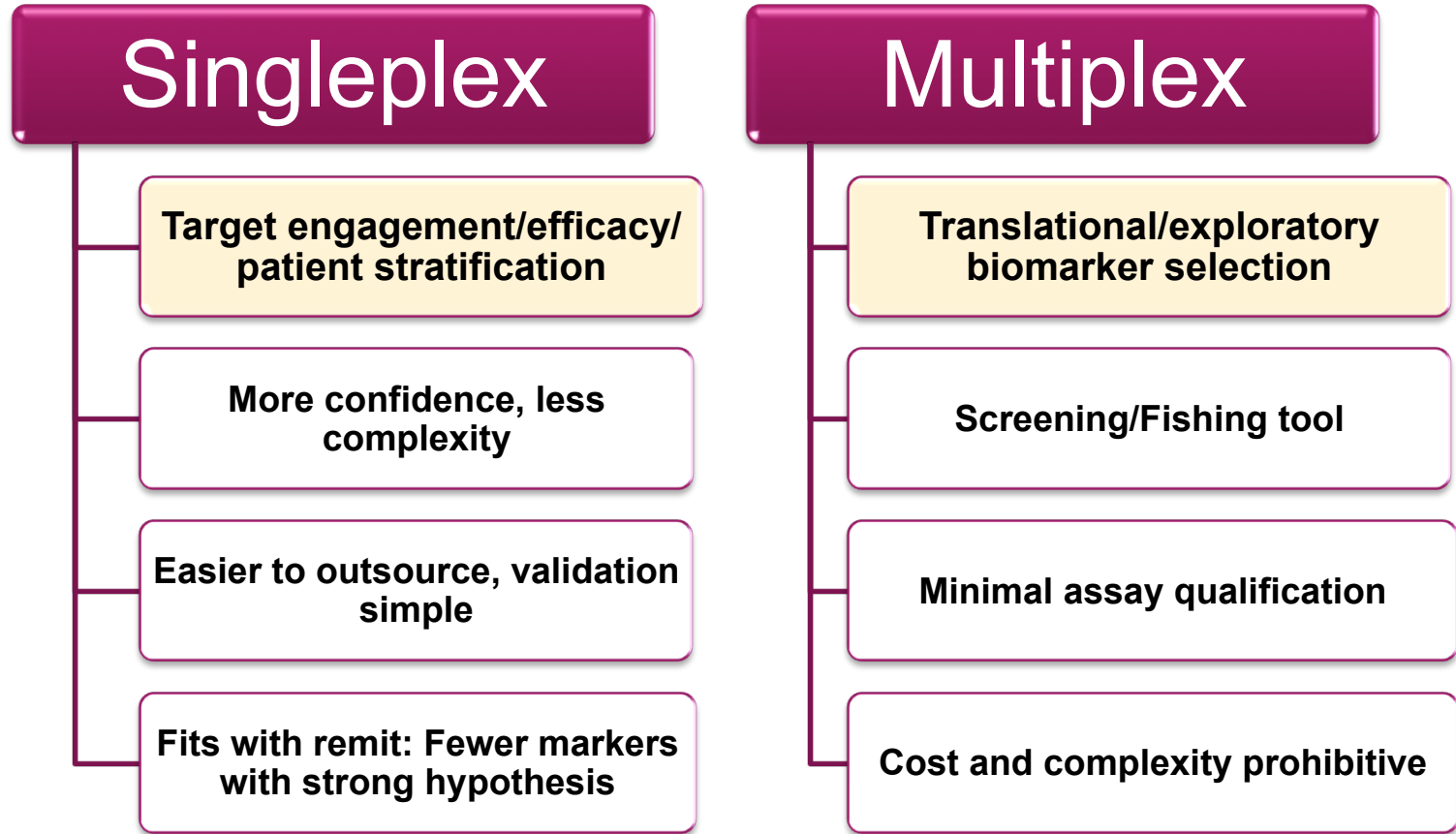
What are the major analytical differences and considerations?

Do we get the same results from a multiplex and singleplex assay?

Can we use multiplex and singleplex assays seamlessly through the drug development process to measure the same biomarker?



# Our historical perspective



## The Multiplex shift



# Recent experience with multiplex



## Request to measure 5 markers in human plasma

- 4 weeks to complete
- Chose multiplex analysis due to time and budget constraints
- MSD V -Plex Pro-inflammatory panel
- Designed and performed tailored package of work
  - Focus on assay precision and limit of quantification



## Requirement for validated IL-8 and TNF-Alpha assays

- V-Plex IL-8 assay validated in human serum
- Cross validation of V-Plex TNF-Alpha assay vs. original high sensitivity assay (plasma/sputum)
- Validation according to internal SOP
  - P&A with matrix QCs, LLOQ, Matrix recovery...



- ✓ IL-8 & TNF- $\alpha$  both on the 5-plex plate
- ✓ Spare plates available
- ✓ Opportunity to directly compare assays

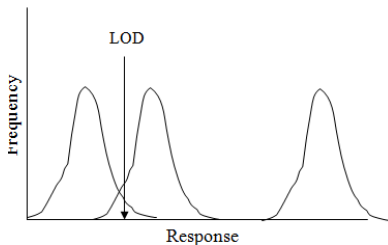


# What was missing?

What are the key features that may differ between the formats?

## Sensitivity/LOD

Differentiation of low concentration analyte from background



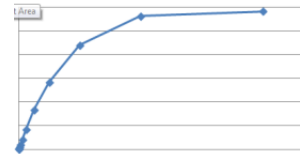
## Interference

From other matrix components but also other analyte spots in the well



## Hook Effect

Is this more prevalent in multiplex assays?



## Sample values

The ULTIMATE question:  
Do we get the same results for 'real' samples?



# Experimental design

## • 3 batch study

- Each batch run on the 3 assay types on the same day by the same analyst

IL-8 singleplex

TNF- $\alpha$  singleplex

Multiplex 5-plex

- Bulk sample preparation to minimise pre-analytical variation
- All antibodies added for multiplex plate (5 analytes in total)

### Calibration standards

- Singleplex kit standards (Multi Analyte)
- Multiplex kit standard (Multi Analyte)
- Alternative source standards (Single Analyte)

### Hook Effect

- Up to 50,000 pg/mL (Approx. 100xULOQ) in assay diluent

### Sensitivity/Limit of detection

- Diluent vs. HV Serum vs. HV Sputum
- n=26 wells of each in same plate

### Interference

- (8x) HV Serum + 50 IU/MI Rheumatoid Factor
- (8x) HV Serum + 1000 pg/mL Interleukin-6
- (8x) HV Serum + diluent (control)

### Sample Concordance

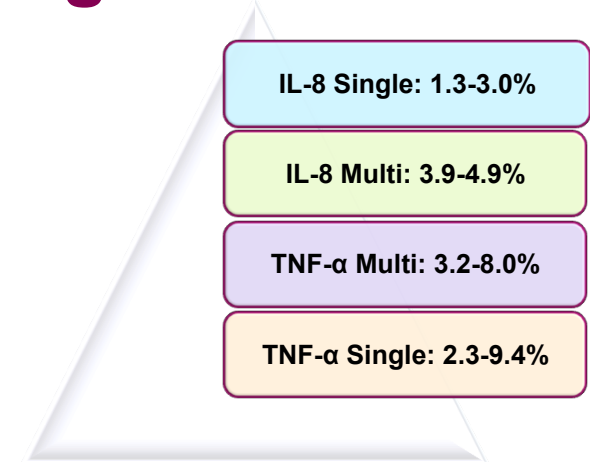
- Patient serum and LPS sputum samples
- No spike addition
- Diluted according to likely concentration & DTT effect



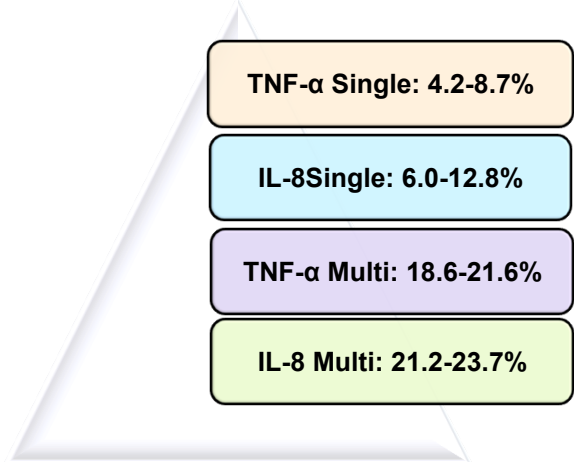
# Results



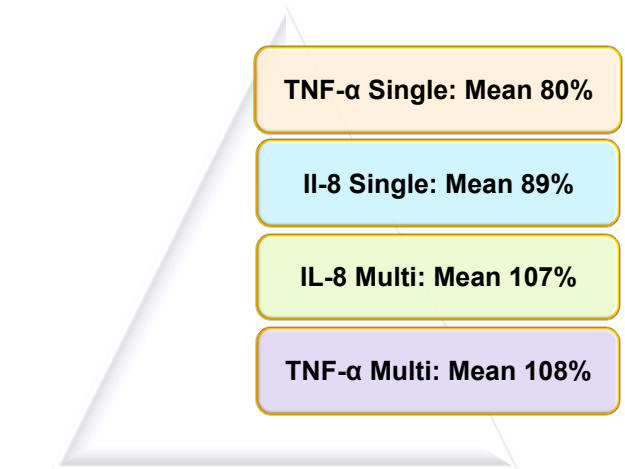
# Original assessments



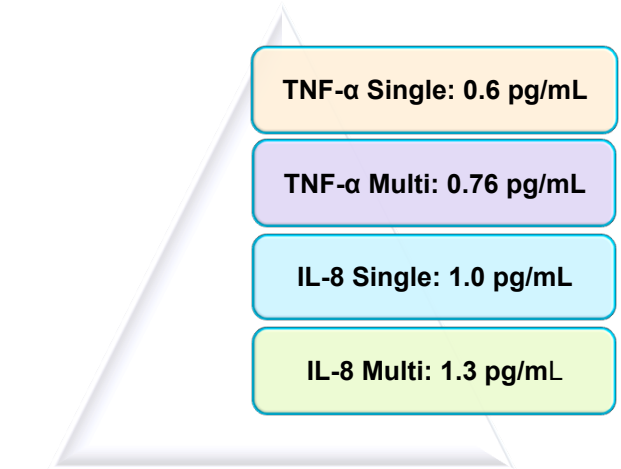
**Intra-assay precision**



**Inter-assay precision**



**Matrix specificity**

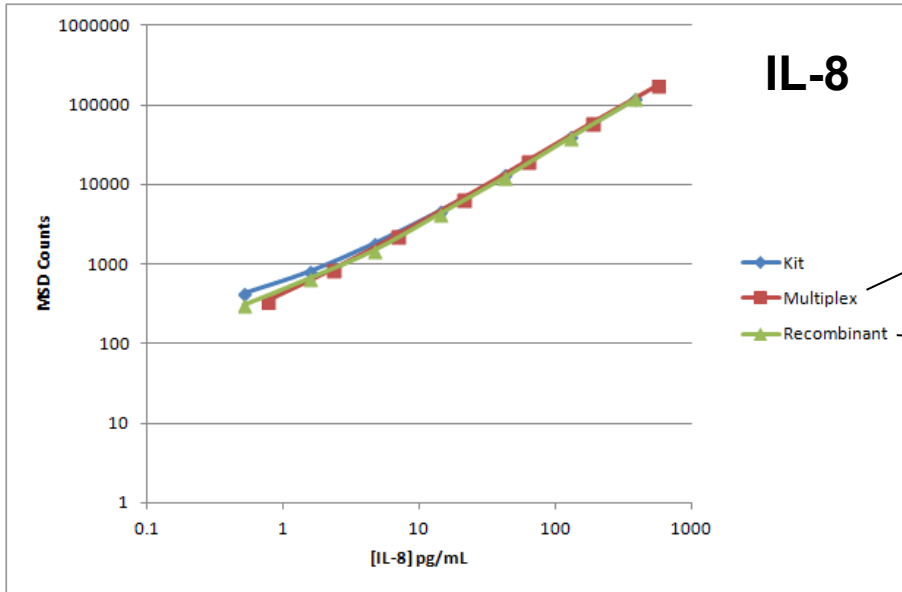


**LLOQ**





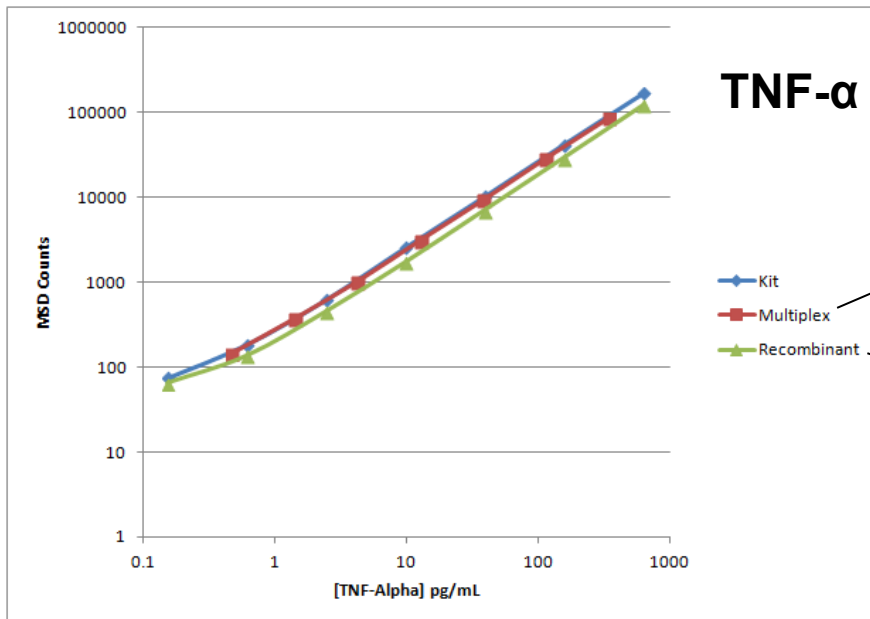
# Calibration standards (Singleplex plates)



95% concordance

94% concordance

**Multiplex kit: 99% concordance**



95% concordance

72% concordance

**Multiplex kit: 73% concordance**

# Interference testing (HV serum)

## IL-8

Sample	Control (Single)	Control (Multi)
1	50.7	56.8
2	101	103
3	217	223
4	3.2	3.98
5	5.42	6.54
6	68	81.5
7	17.5	20.1
8	5.71	6.24

+ IL-6 (1000 pg/mL)

Recovery (Single)	Recovery (Multi)
98	104
97	101
104	100
97	115
108	117
100	89
102	102
81	91
<b>Mean recovery</b>	<b>Mean recovery</b>
98.4	102.4

+ RF (50 IU/mL)

Recovery (Single)	Recovery (Multi)
103	107
98	100
109	109
101	102
106	110
100	87
107	100
82	89
<b>Mean recovery</b>	<b>Mean recovery</b>
100.8	100.5

## TNF- $\alpha$

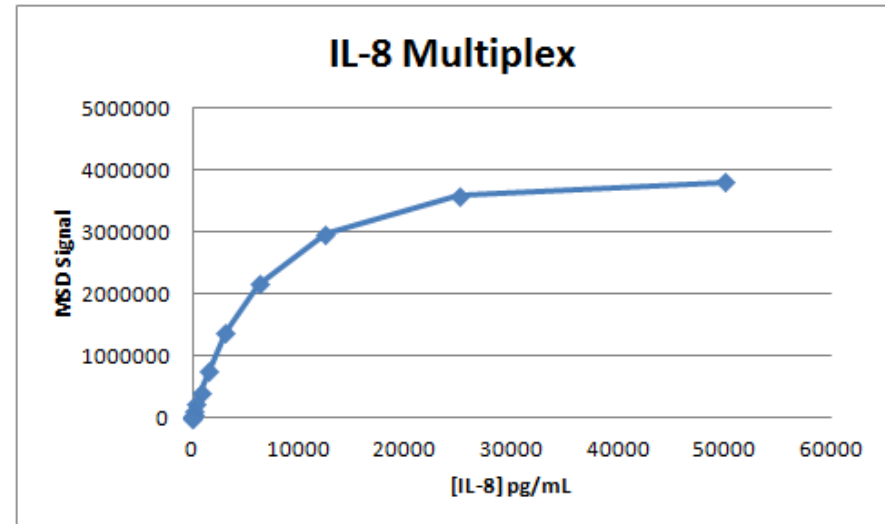
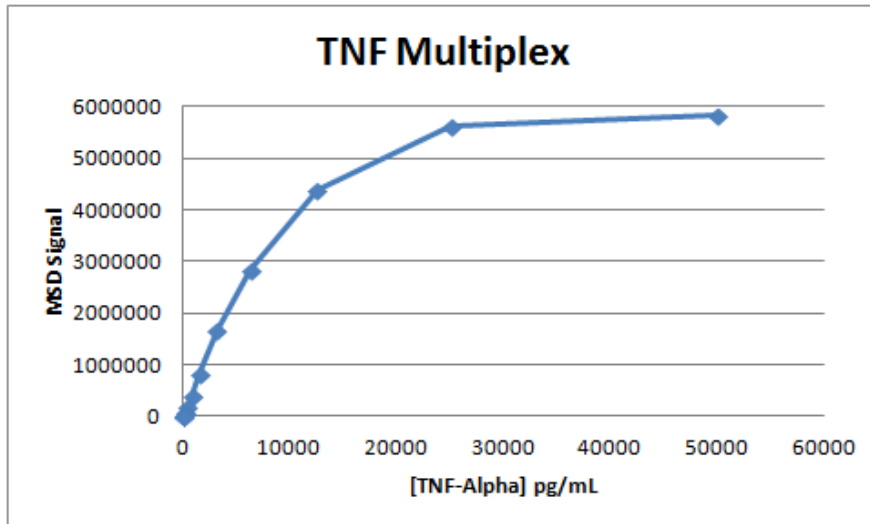
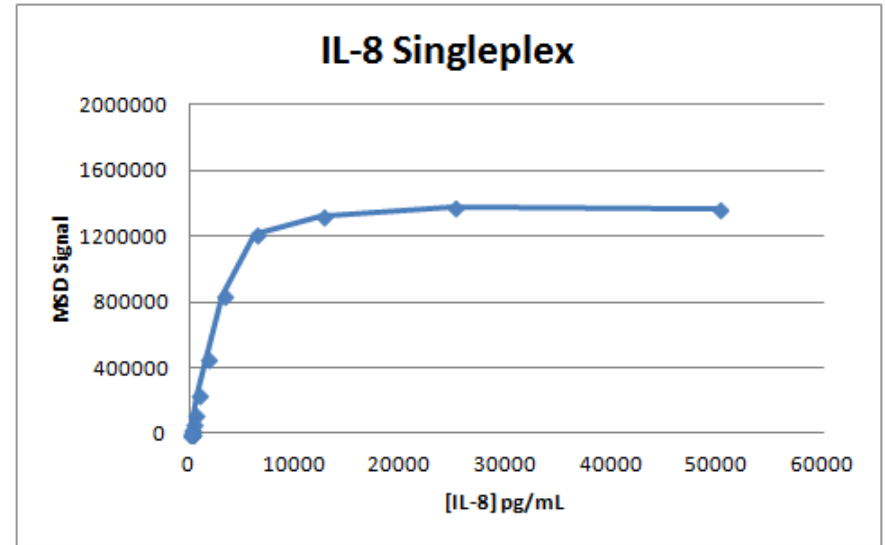
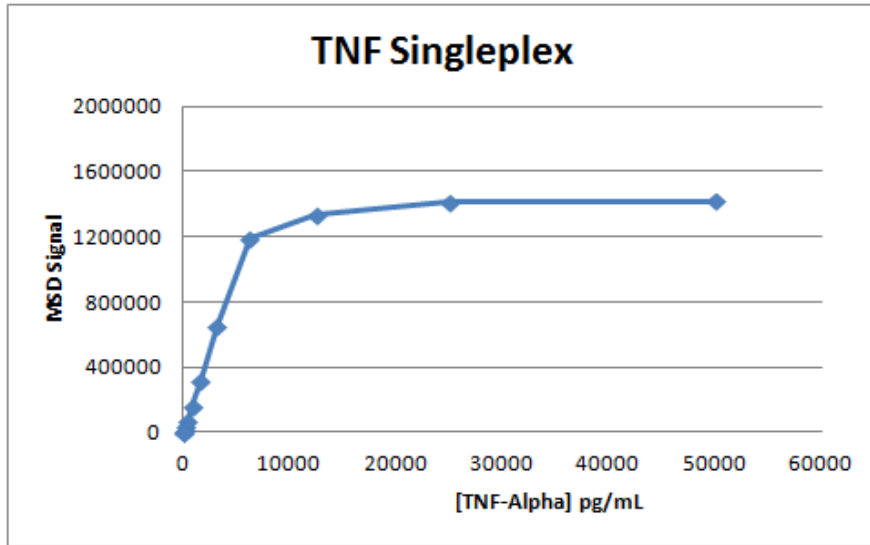
Sample	Control (Single)	Control (Multi)
1	0.326	0.431
2	0.4	0.533
3	0.405	0.383
4	0.732	0.743
5	0.976	1.14
6	1.76	2.05
7	0.74	0.714
8	1.18	1.36

Recovery (Single)	Recovery (Multi)
108	111
125	123
79	108
107	107
113	110
93	95
98	126
110	107
<b>Mean recovery</b>	<b>Mean recovery</b>
104.1	110.9

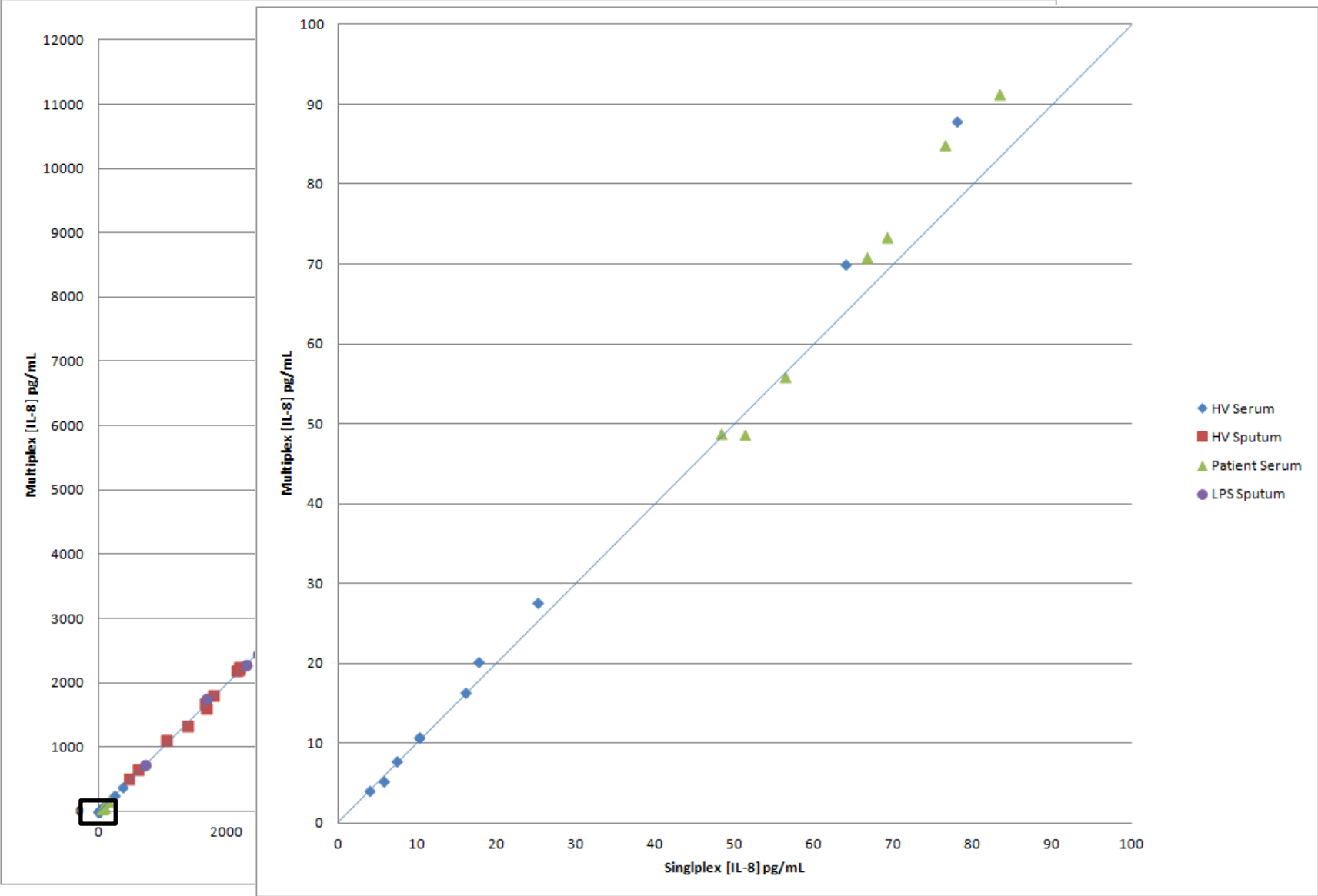
Recovery (Single)	Recovery (Multi)
1215	935
980	583
1173	1099
533	431
504	427
307	268
512	507
381	319
<b>Mean recovery</b>	<b>Mean recovery</b>
700.6	571.1



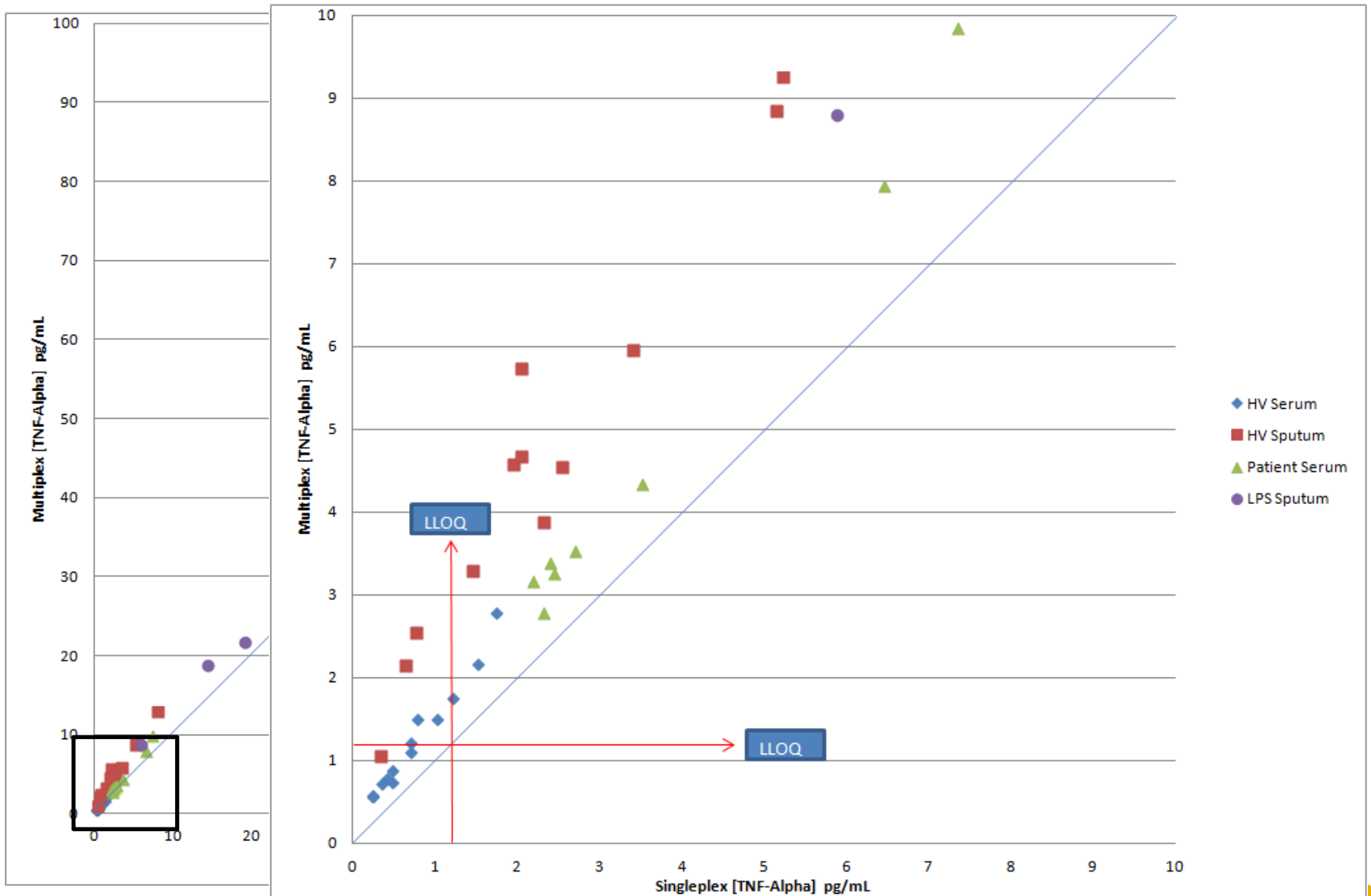
# Hook Effect



# Sample concordance: IL-8

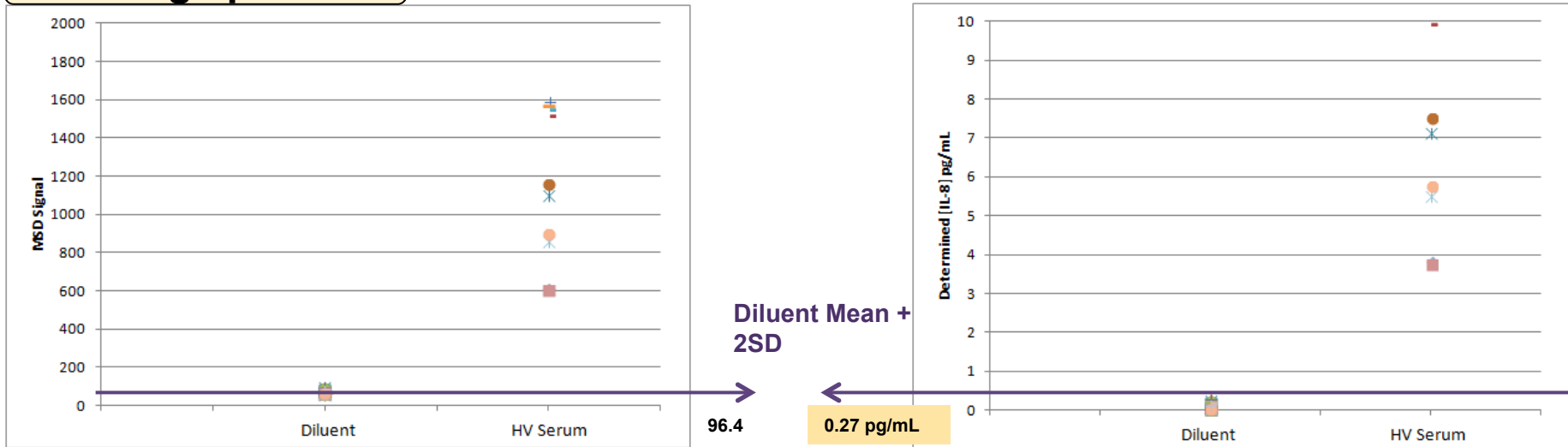


# Sample concordance: TNF-Alpha

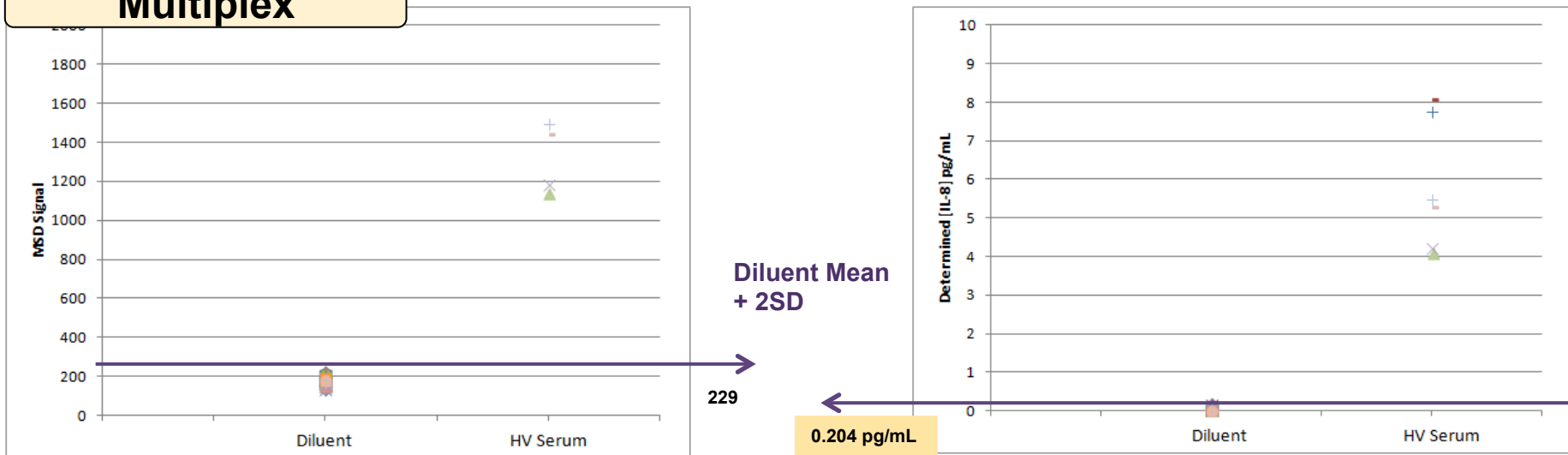


# Sensitivity/Limits of detection: IL-8

## Singleplex

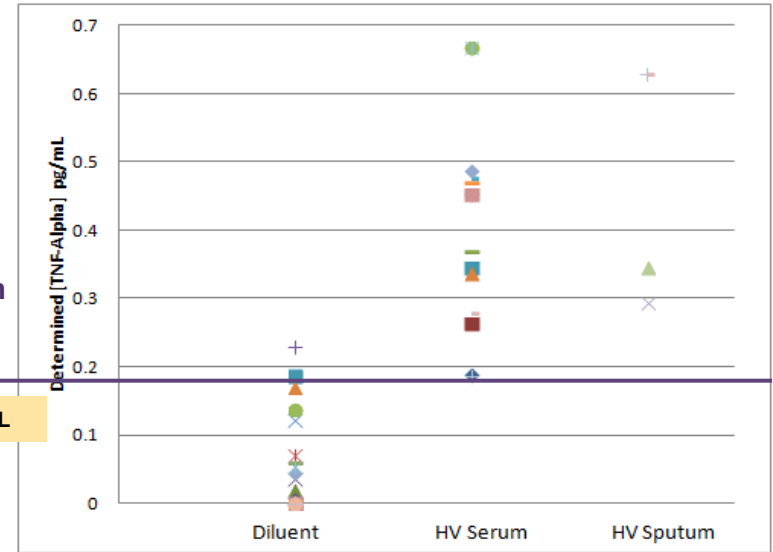
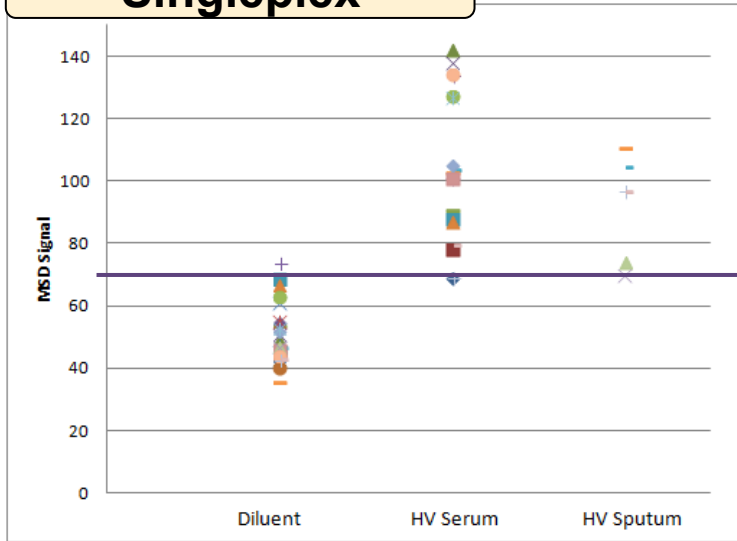


## Multiplex

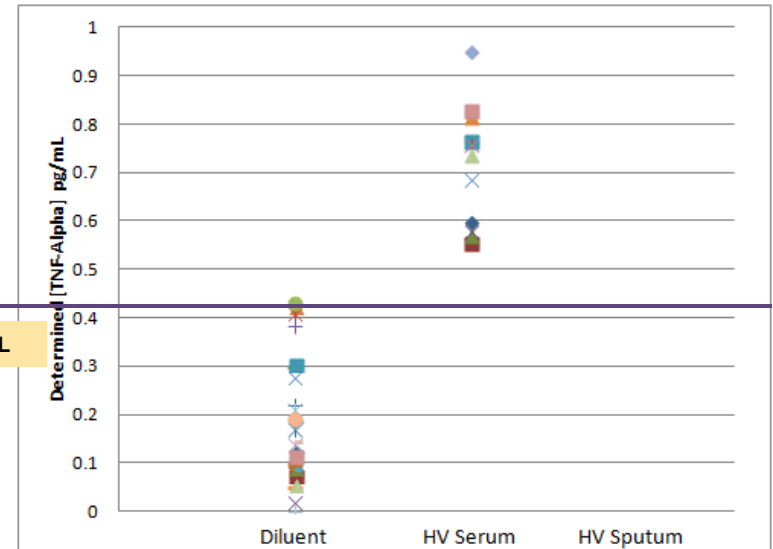
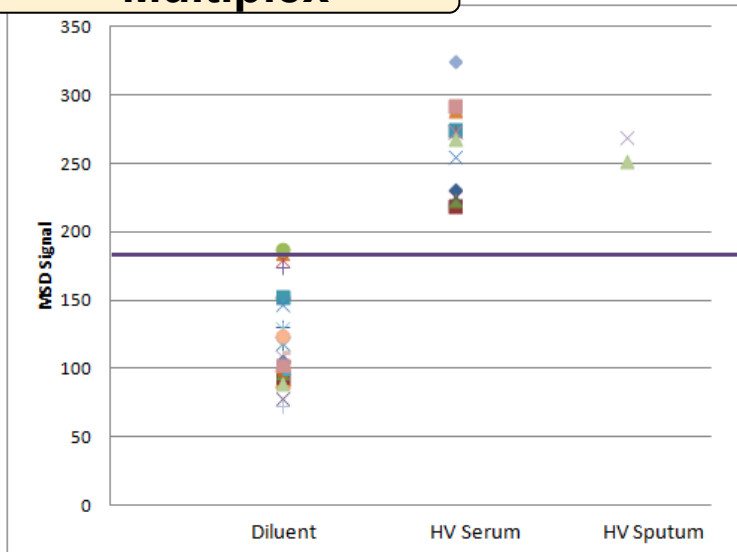


# Sensitivity/Limits of detection: TNF-Alpha

## Singleplex



## Multiplex



# Discussion

## What are the major analytical challenges and considerations?

- Complexity of assessing interference from numerous sources
- Consider ranges for all analytes and proximity to LLOQ
- Understand the precision requirements of the assay and impact if different results are generated from multiplex to singleplex
- Need to discriminate the validation carried out versus intended use e.g. BM fishing, Informative in clinic or Decision making in clinic

## Do we get the same results from a multiplex and singleplex assay?

- **Yes:** Sensitivity, interference, calibration standards, mid/high concentration analytes (IL-8)
- **No:** Low concentration analytes (TNF-Alpha), small differences magnified at the limits (Cal std/DTT effect?)

## Can we use multiplex and singleplex assays seamlessly through the drug development process to measure the same biomarker?

- Yes but need to understand where data can or cannot be compared (continuity)
- Clear strategy around normalisation of data/assays if required

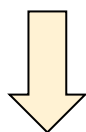




# Thoughts for the future...



- Can we make the transition easier?
  - Use of same platform
  - Use of same calibration standards (normalisation)
- Do we even need to move to singleplex? Do multiplex assays have a utility in later phase work?
  - Potential for exploratory markers
  - Could small (2-plex) assays offer advantages (time, cost, sample volume)



- Assay validation/qualification:
  - What factors need extra consideration?
  - Is there a basic minimum package of work?.... EBF TT-49



# Acknowledgements

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## ANY QUESTIONS?

