

Critical evaluation of common sample preparation techniques for bioanalysis on microfluidic LC/MS performance

Paul Rainville
Waters Corporation

NCBI Resources How To

PubMed.gov

US National Library of Medicine
National Institutes of Health

Advanced

Display Settings: Abstract

Send to:

[Rapid Commun Mass Spectrom.](#) 2009 Dec;23(23):3736-42. doi: 10.1002/rcm.4311.

Online nanoelectrospray/high-field asymmetric waveform ion mobility spectrometry as a potential tool for discovery pharmaceutical bioanalysis.

[Hatsis P](#), [Valaskovic G](#), [Wu JT](#).

Department of Drug Metabolism and Pharmacokinetics, Millennium Pharmaceuticals Inc., 35 Landsdowne St, Cambridge, MA 02139, USA.
panos.hatsis@novartis.com

Abstract

Nanoelectrospray ionization (nESI) coupled online with high-field asymmetric waveform ion mobility spectrometry (FAIMS) for small molecule analysis in a discovery pharmaceutical setting was examined. A conventional capillary pump, autosampler and nESI source were used to introduce samples directly into the FAIMS device. The FAIMS device was used to separate gas-phase ions on a timescale that was compatible with the mass spectrometer. The capability of the nESI-FAIMS combination to efficiently remove metabolite interferences from the parent drug, and reduce ion suppression effects, was demonstrated. On average, 85% of the signal intensity obtained from a neat sample was preserved in the extracted plasma samples. Standard curves were prepared for several compounds. Linearity was obtained over approximately 3 to 4 orders of magnitude. Comparison of results from nESI-FAIMS with those from conventional LC/MS for a mouse pharmacokinetic study yielded concentration values differing by no more than 30%.

Copyright 2009 John Wiley & Sons, Ltd.

PMID: 19902415 [PubMed - indexed for MEDLINE]

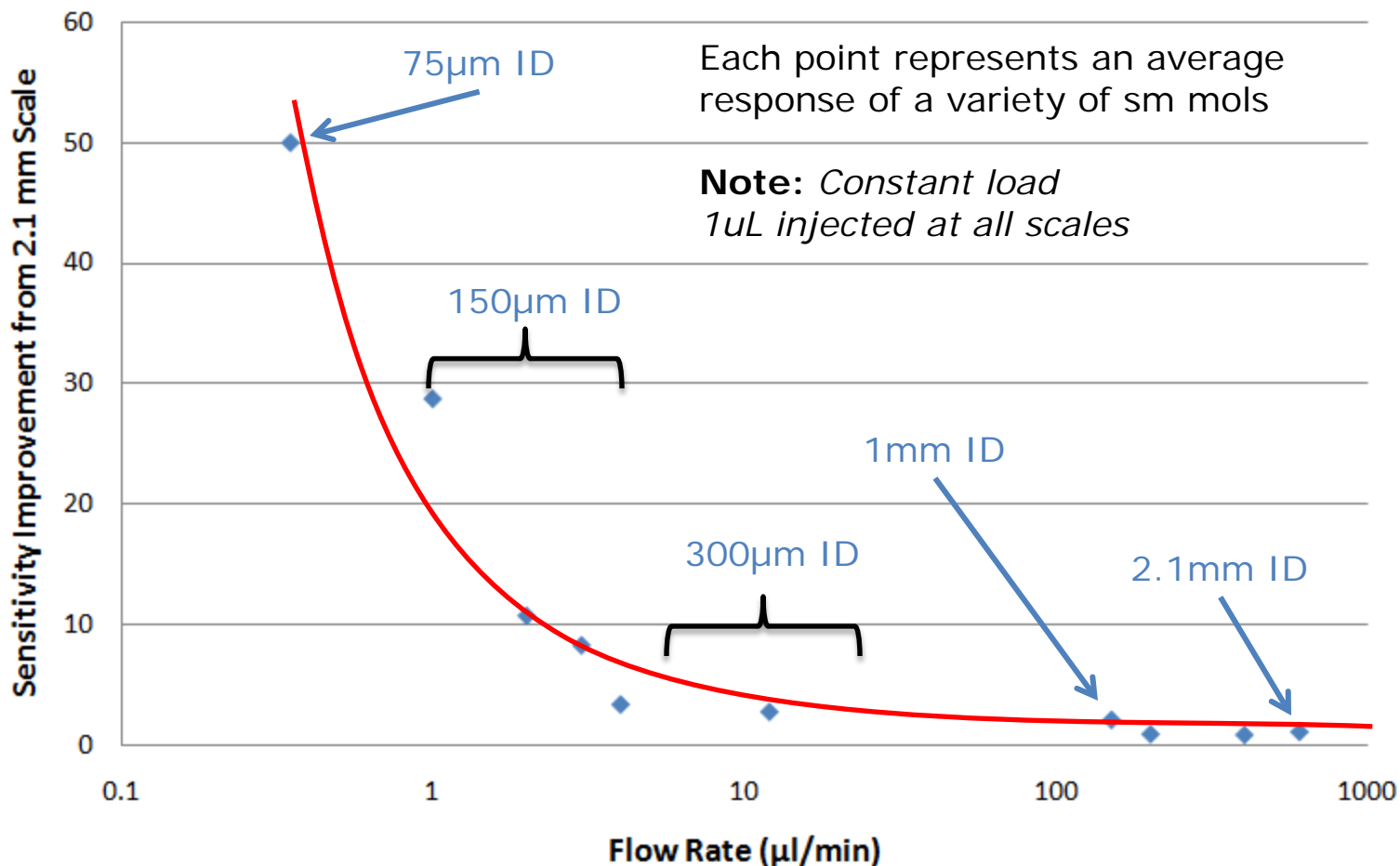
metry
les in
ghput
rotein
oted up
ety in
olites
mass
ature,
speed

CLE

- Why ?
 - Need for sensitivity
 - Limited sample volumes
 - Multiple injections/ same sample
 - Single model PK data
 - “Green” approach
- Why not ?
 - Plenty of sensitivity
 - Plenty sample available
 - Speed on analysis
 - Proteomics not DMPK platform
 - Chromatographic performance*
 - Robustness*

Signal Intensity Improvement in comparison to a 2.1mm format

Small Molecule



Separation Comparison

RAPID COMMUNICATIONS IN MASS SPECTROMETRY
Rapid Commun. Mass Spectrom. 2008, 22: 1053-1061
Published online in Wiley InterScience (www.interscience.wiley.com) DOI: 10.1002/rcm.3472

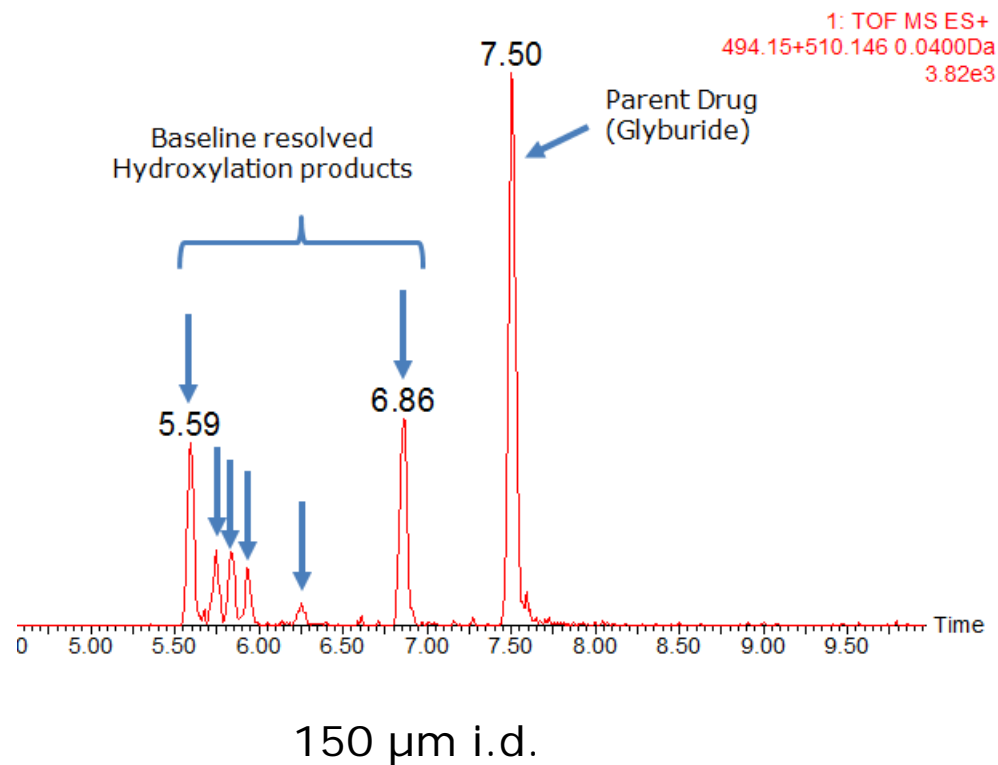
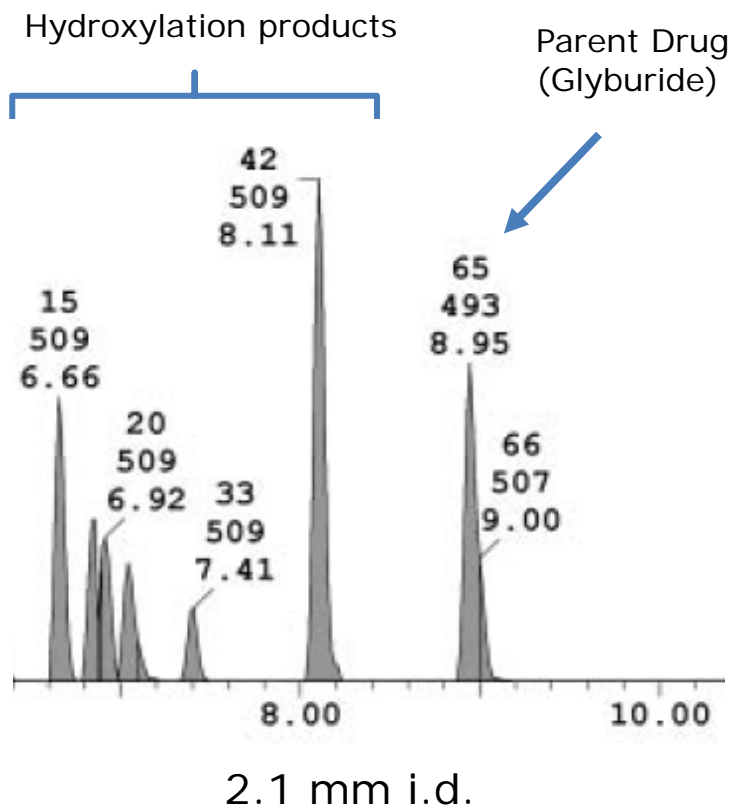
RCM

High-throughput, accurate mass liquid chromatography/
tandem mass spectrometry on a quadrupole time-of-flight
system as a 'first-line' approach for metabolite
identification studies

Philip R. Tiller^{1*}, Sean Yu¹, Jose Castro-Perez², Kerry L. Fillgrove¹ and Thomas A. Baillie¹

¹Drug Metabolism and Pharmacokinetics, Merck Research Laboratories, West Point, PA 19486, USA
²Waters Corporation, Milford, MA 01757, USA

Received 11 October 2007; Revised 28 January 2008; Accepted 29 January 2008

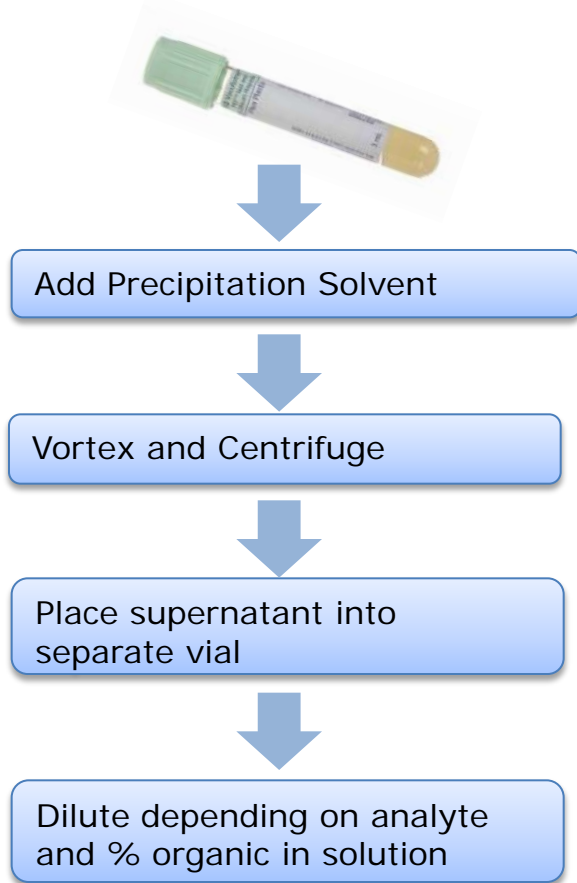


- Place into format acceptable for injection onto LC/MS system.
- Various techniques depending on goal of analysis:
 - Protein precipitation*
 - Liquid-liquid extraction*
 - Solid-phase extraction
 - Solid-liquid extraction
 - Digestion*
 - Affinity
 - Filter
 - Centrifugation
 - Dilution

- Various biological matrices tested with small and large molecules:
 - Plasma*
 - Urine
 - Bile
 - Microsomes
- Criteria: 1000 injections (approx. 5 days)
 - Chromatographic peak shape
 - System pressure
- Chromatographic conditions:
 - 150 μm X 50 mm prototype microfluidic device 1.7 μm BEH C₁₈, , temperature controlled, 3-4 $\mu\text{L}/\text{min}$, gradient elution, formic acid/ MeCN , various injection volumes

Robustness Evaluation Test Case 1

A typical Protein Precipitation workflow



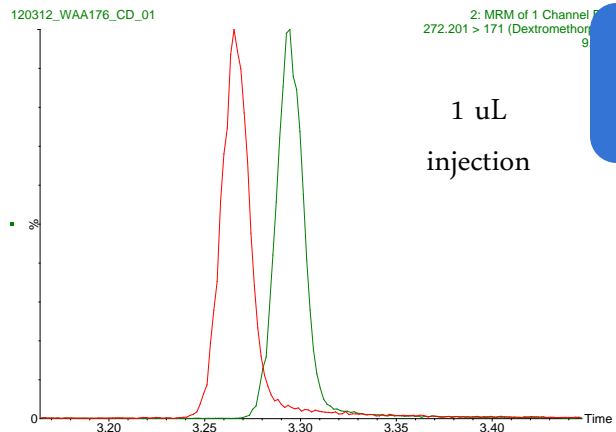
■ Protein precipitation (PPT)

- 1000 1 μ L injections of 2:1 crashed human plasma
- 1000 3 μ L injections of 2:1 crashed human plasma
- 1000 5 μ L injections of 2:1 crashed human plasma
- a standard mixture of dextromethorphan and propranolol (critical pair) was injected every 50 plasma injections

Robustness Testing (PPT data)

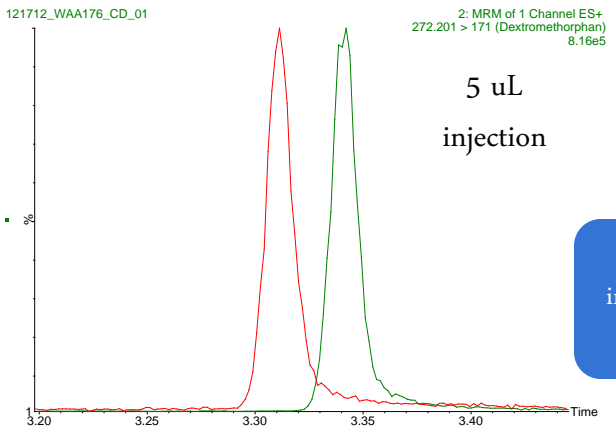
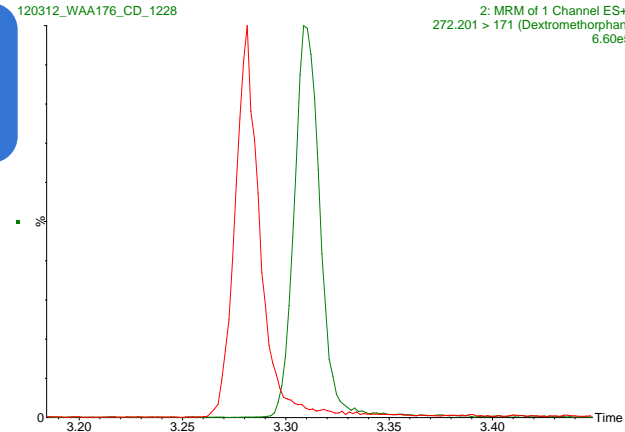
First injection

After 1000 injections of crashed plasma



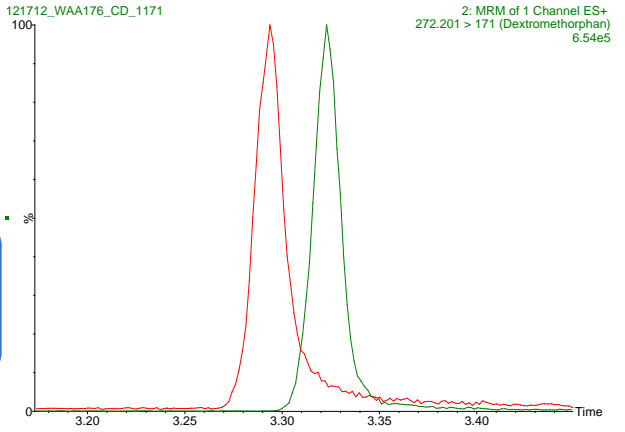
1 uL injection

Equivalent to 200uL injection on 2.1mm scale!

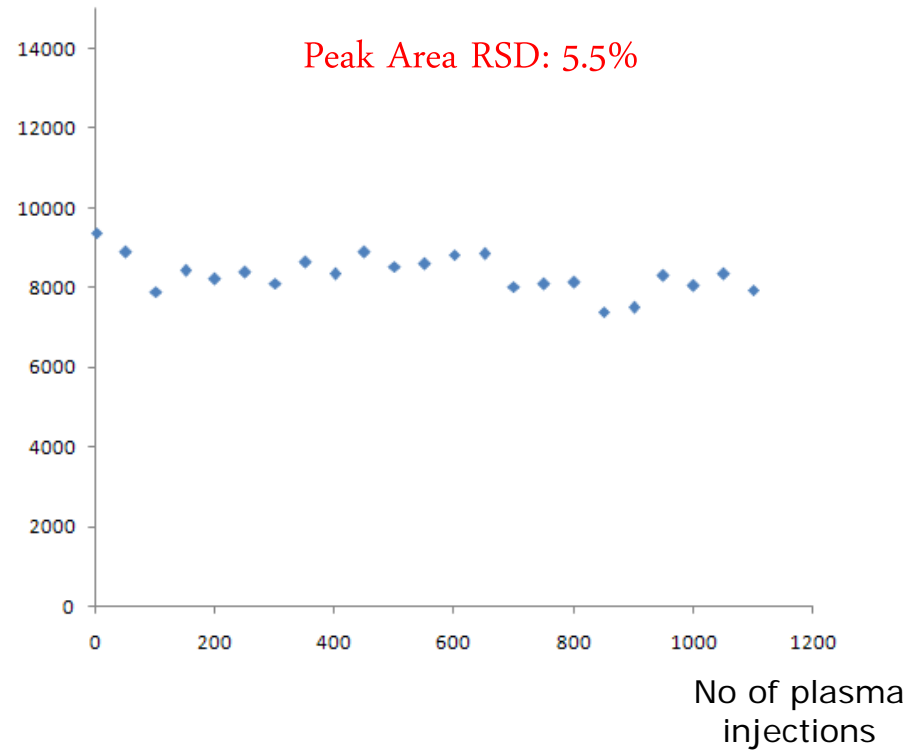
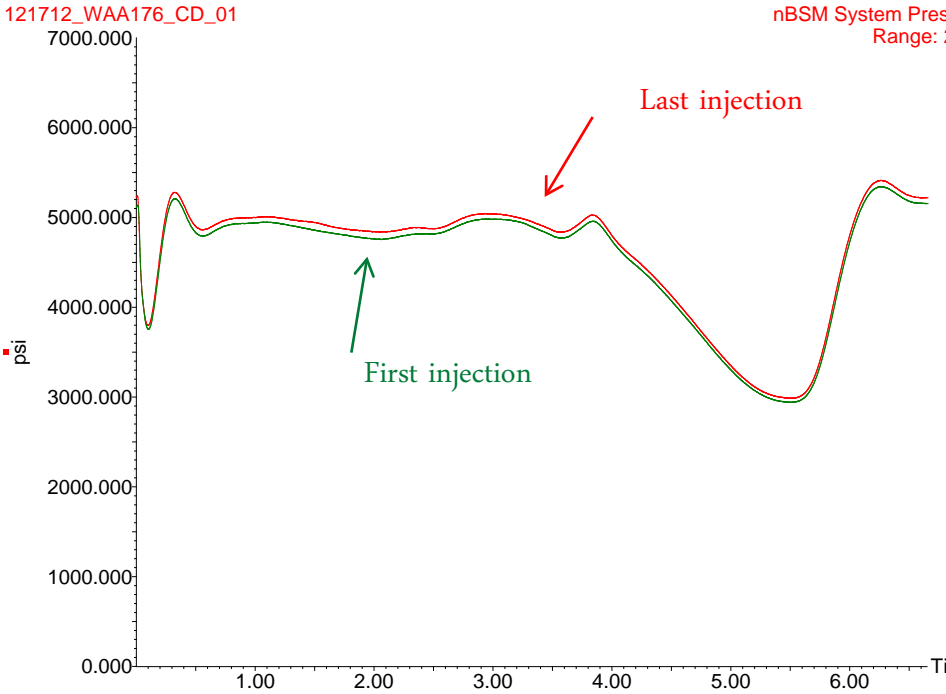


5 uL injection

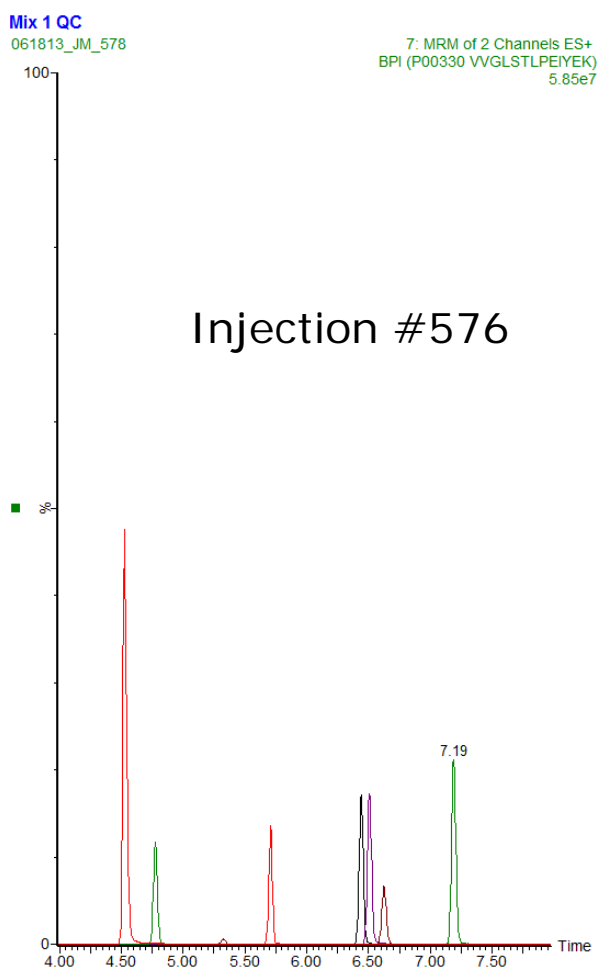
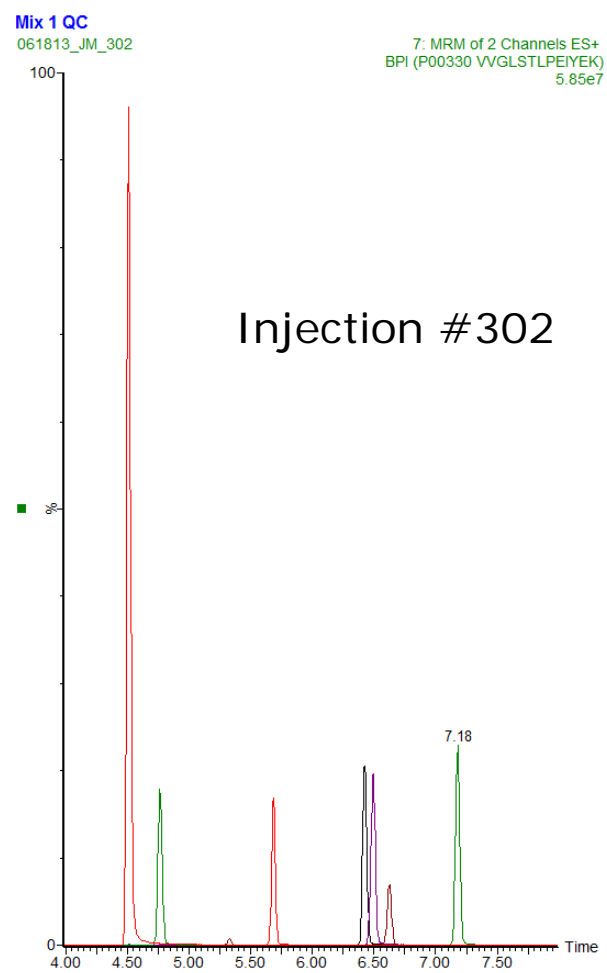
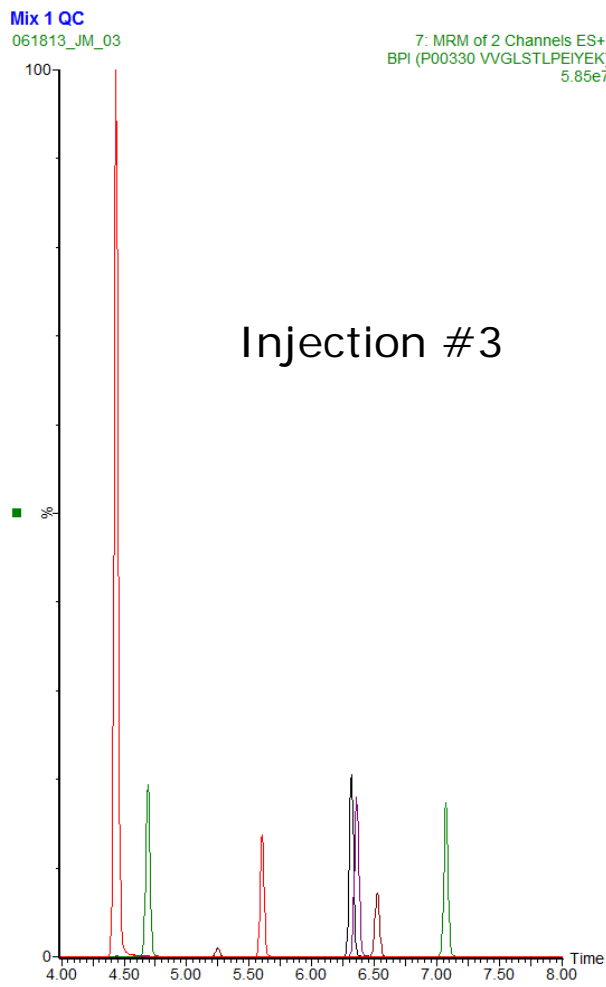
Equivalent to 1mL injection on 2.1mm scale!



Robustness Testing (PPT data)



Peptide Test (PPT data)



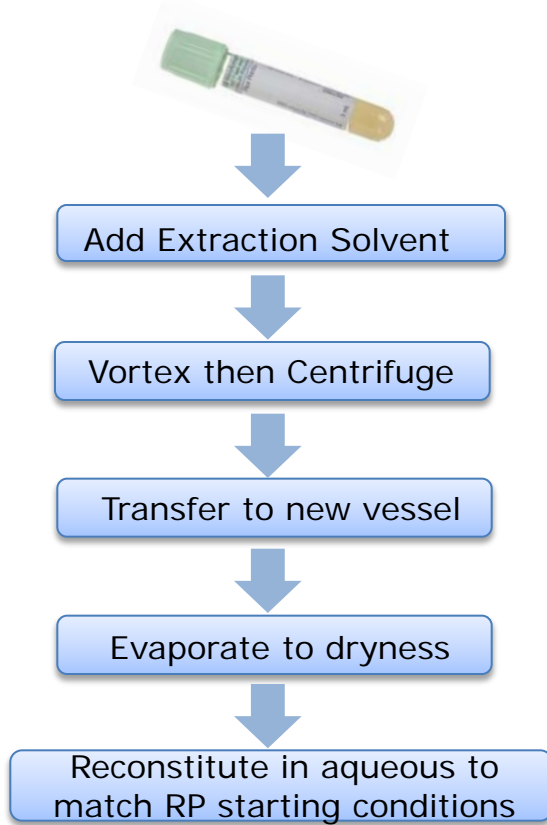
Different devices Different plasma sources (PPT data)

	Peptide P00924
Retention Time	6.64 min
SD of Retention Time	0.09
Average Peak Width at 10%	2.64 s

	Average of all Peptides, all Tiles
Peak Width RSD	4.16%
Retention Time RSD	0.57%
Area RSD	15.5%

Robustness Evaluation Test Case 2

A typical LLE workflow



Solvents Tested

Hexane

Ethyl Acetate

Methyl tert Butyl Ether

1:10 ratio plasma:solvent

Dry down

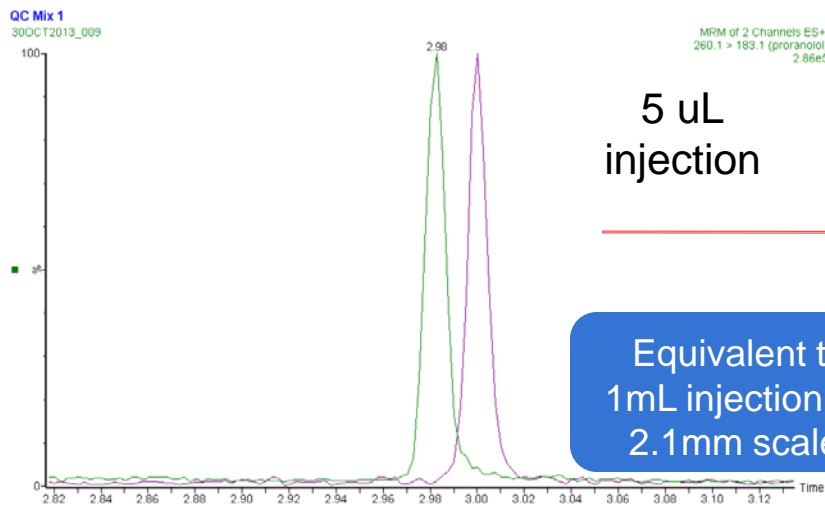
Recon in initial gradient conditions

X 5 enrichment

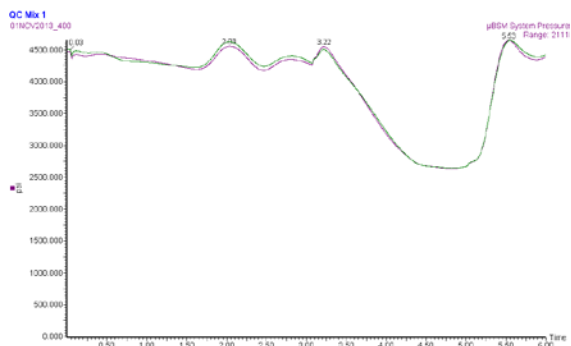
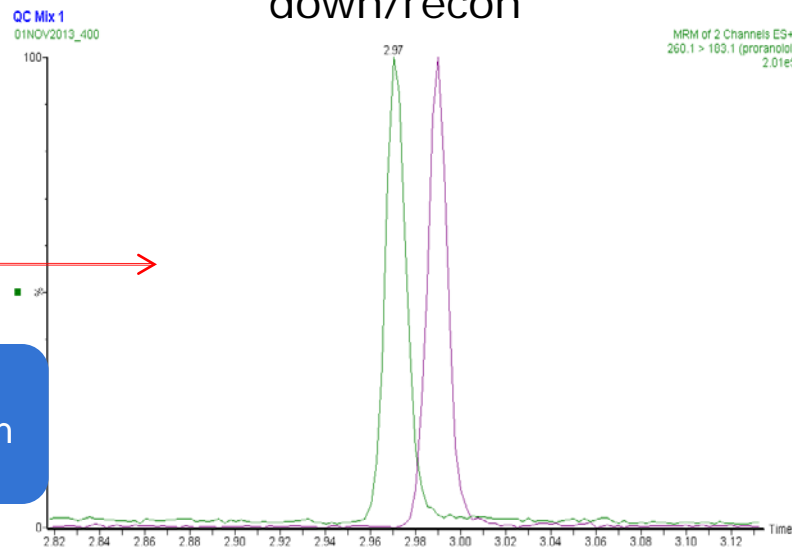
5 μ L injection volume

Robustness Testing (LLE data)

First injection

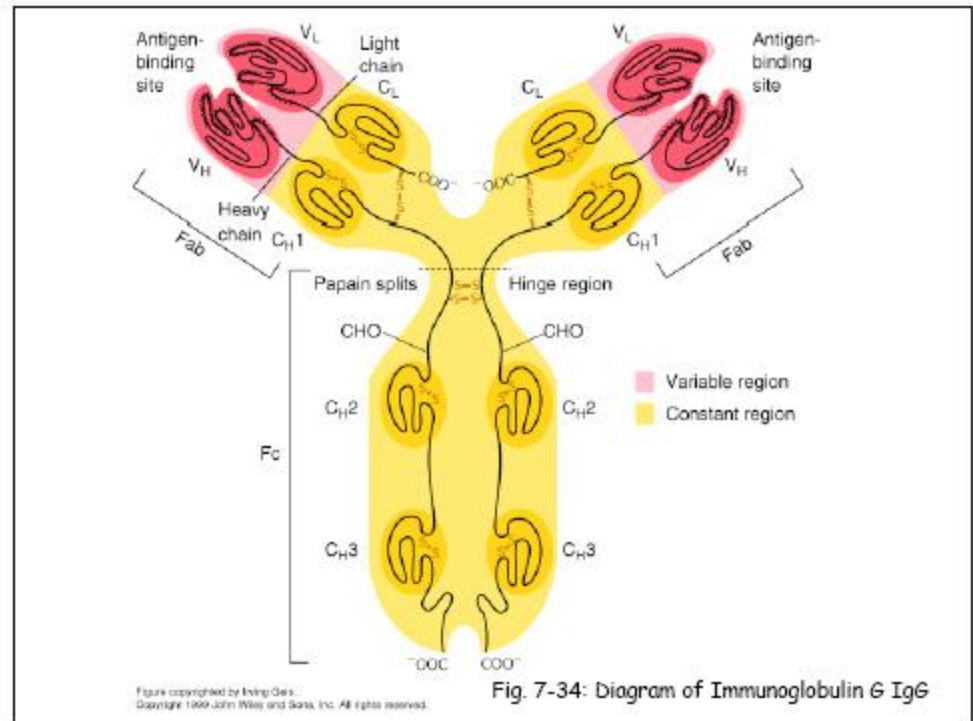
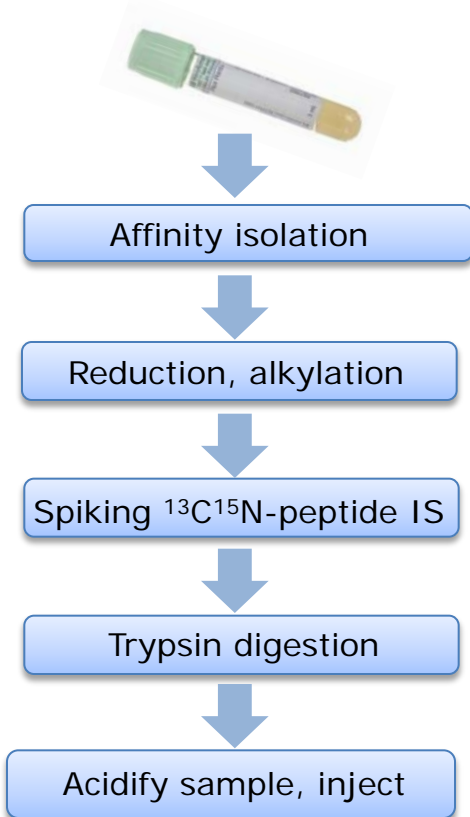


After 1000 injections of LLE/dry down/recon



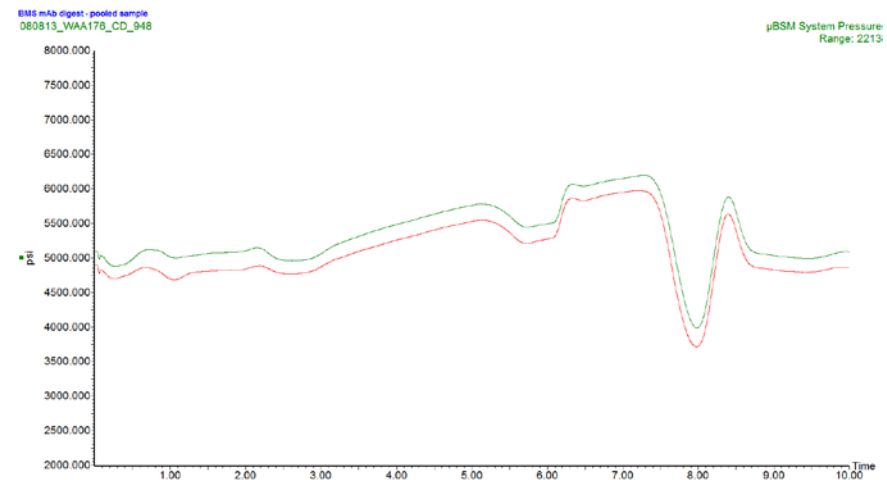
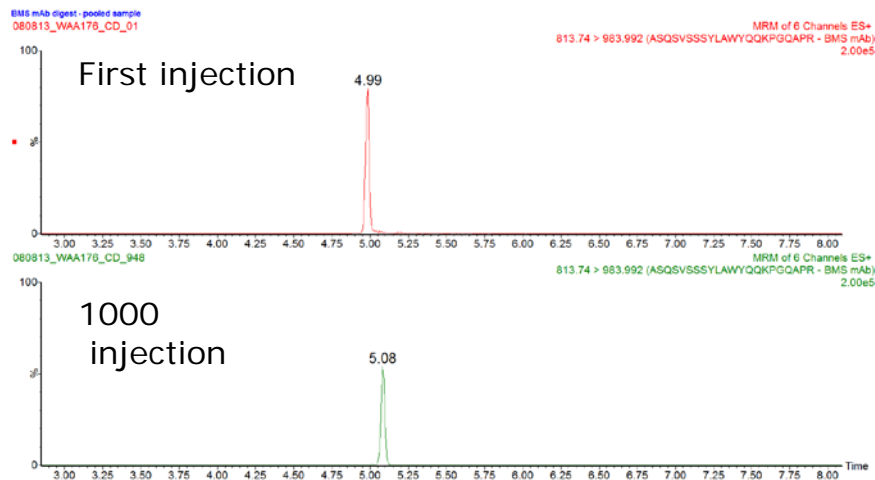
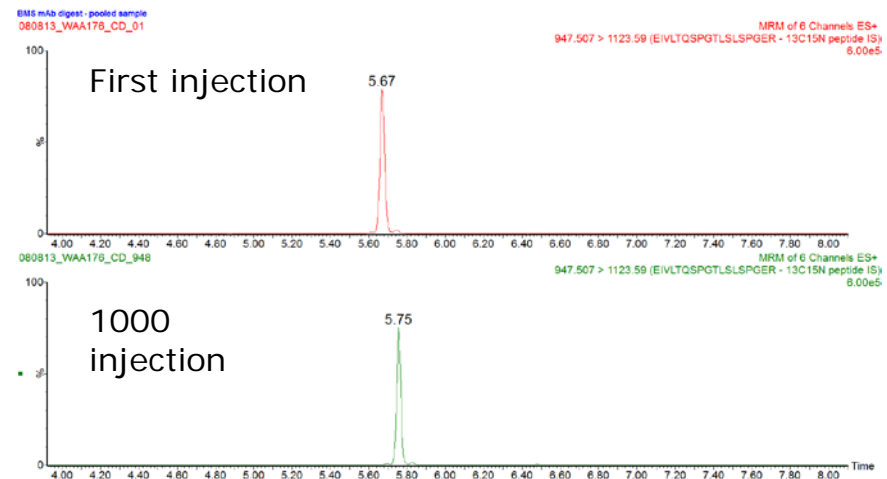
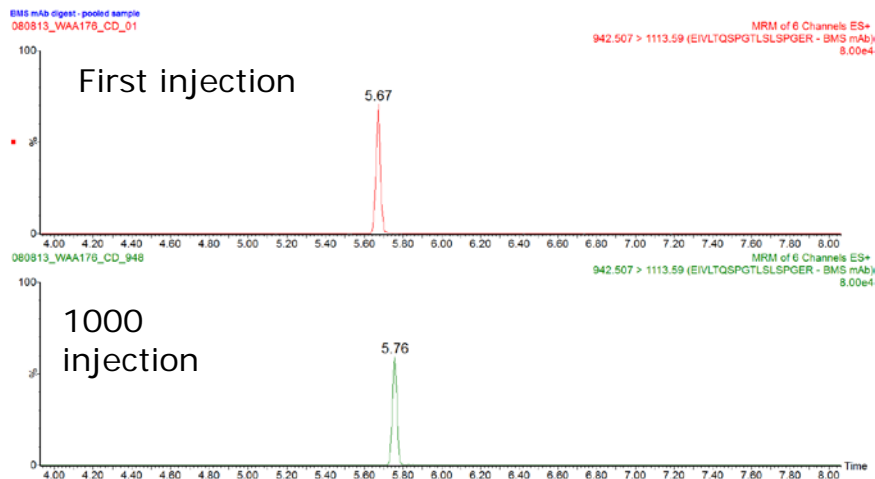
Robustness Evaluation Test Case 3

A typical Protein BioA workflow

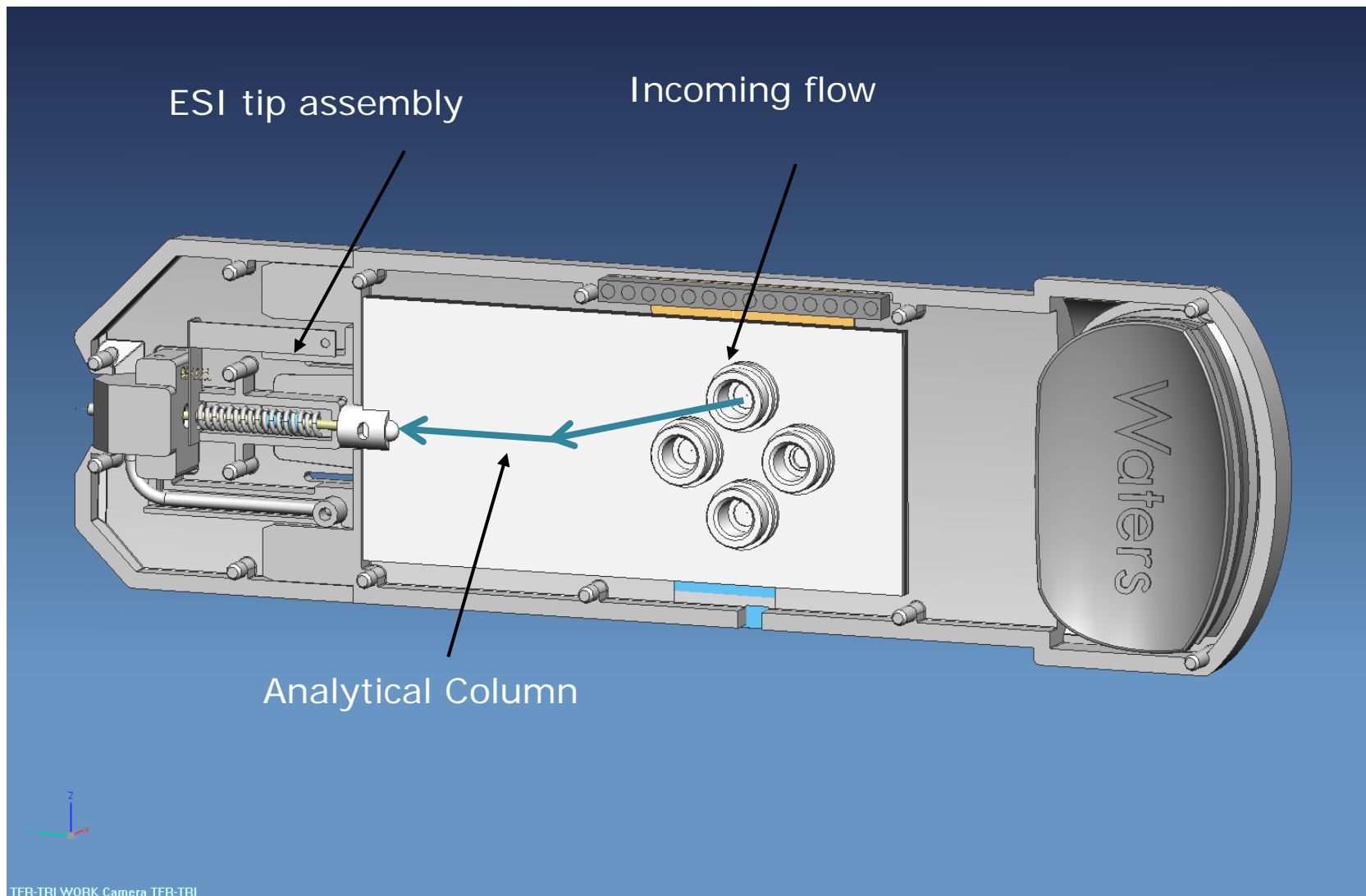


https://www.google.com/search?q=mAb&source=Inms&tbm=isch&sa=X&ei=gwN5UsSWK_jfsASZg4DwBA&ved=0CAcQ_AUoAQ&biw=1024&bih=577

Robustness Testing (mAb digest data)



Separations Device



TFR-TRI WORK Camera TFR-TRI

Ceramic Microfluidic Device

Waters
THE SCIENCE OF WHAT'S POSSIBLE.™





Research Article

Received

Accepted: 14 March 2013

Published online in Wiley Online Library

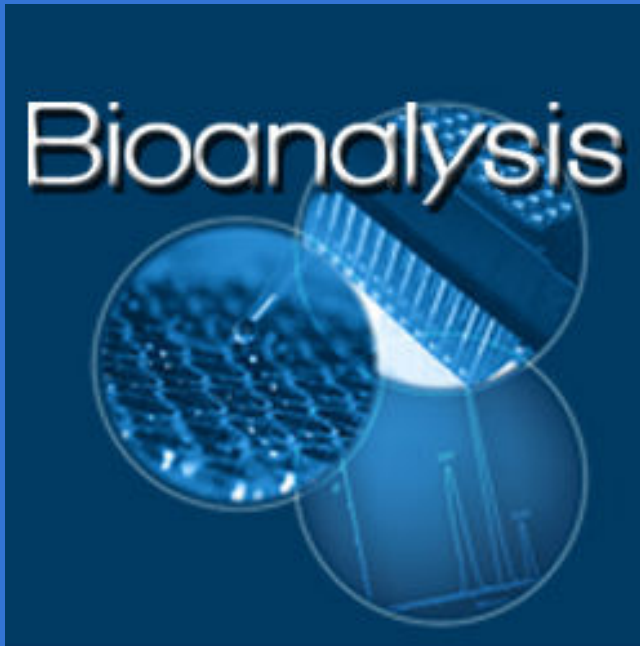
Rapid Commun
(wileyonlinelibrary.com)

Mass Spectrometry

High Resolution
Thermally Stable
Bridged
Gills

¹Molecular
²Clinical
³Clinical
⁴Colorectal
⁵Division
Philadelphia

RAT
factor



M Lassman *et al.*

Measurement of Lp(a) in human subjects using a stable isotope labeled tracer and mass spectrometry

M. Lassman^{2†}, Tiffany Thomas⁴, Wenyu Li¹,
John A. Wagner³, David E. Gutstein³,
Henry N. Ginsberg⁴,
Thomas P. Roddy¹ and Steven F. Previs^{1*}

¹Whitehouse Station, NJ, USA

²Whitehouse Station, NJ, USA

³Whitehouse Station, NJ, USA

⁴School of Medicine, University of Pennsylvania,

Measurement of lipoprotein(a) [Lp(a)] in human subjects using a stable isotope labeled tracer and mass spectrometry is a component of lipoprotein(a) [Lp(a)], an independent risk factor for cardiovascular disease. The role of Lp(a) in blood is poorly understood in part due to technical challenges in measuring Lp(a) kinetics. Improvements in the ability to readily and reliably measure the kinetics of apo(a) using a stable isotope labeled tracer is expected to facilitate studies of the role of Lp(a) in cardiovascular disease. Since investigators typically determine the isotopic labeling of protein-bound amino acids following acid-catalyzed hydrolysis

- A 150 μm i.d. ceramic micro-fluidics prototype device was successfully implemented in the analysis of biofluid samples with good chromatographic performance.
- Common sample preparation techniques already utilized in BioA workflows can be readily implemented when scaling down chromatography to 150 μm i.d.
 - PPT
 - LLE
 - IA/Digestion

Acknowledgements

- Jim Murphy
- Jay Johnson
- Mark Wrona
- Catalin Doneanu
- Erin Chambers