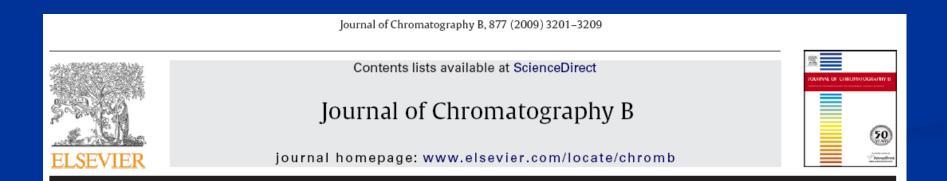


#### Leveraging Successful Troubleshooting Experiences for the Prevention or Reduction of Internal Standard Response Variations during LC-MS Bioanalysis

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# You might have experienced different IS response variations just as I did in the past!



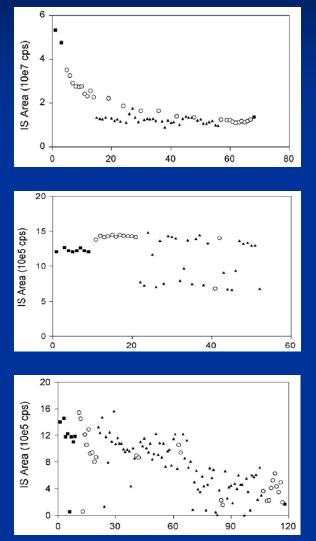
Internal standard response variations during incurred sample analysis by LC–MS/MS: Case by case trouble-shooting

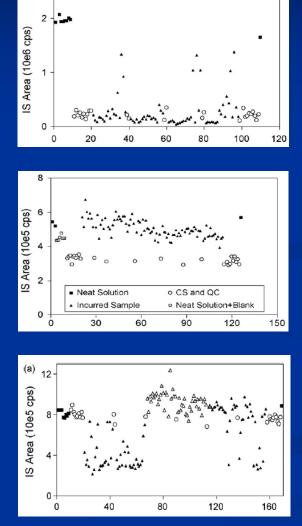
Aimin Tan<sup>a, b,\*</sup>, Saleh Hussain<sup>a</sup>, Adrien Musuku<sup>b</sup>, Robert Massé<sup>b</sup>

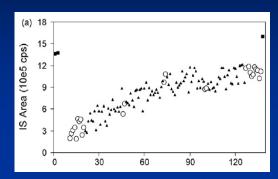
<sup>a</sup> Anapharm, Richmond Hill, Ontario, Canada

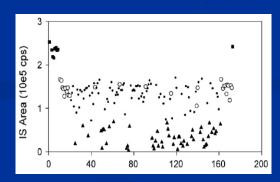
<sup>b</sup> Anapharm, Ste-Foy, Québec, Canada

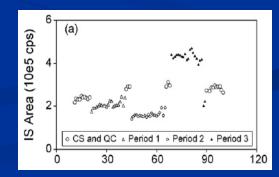
### Look familiar?











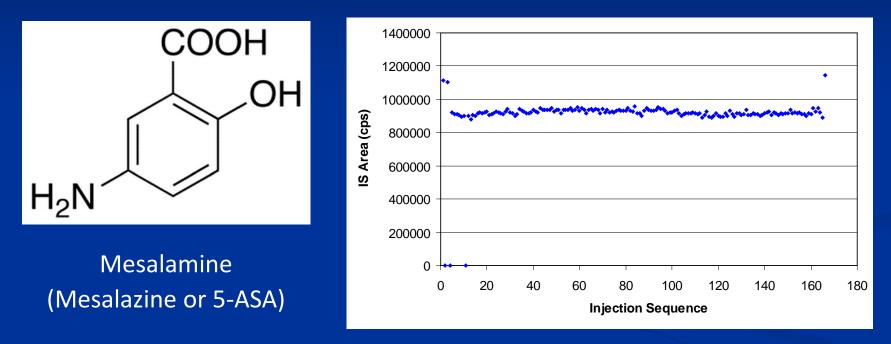
# Can we reduce or prevent IS response variations?

## Yes, we can!

Average % of 15 repeats (study wise)	0.2
Average % of IS repeats (study-wise)	(0-1.9)
Average % of IS repeats (sample-wise)	0.2
% Studies without IS repeats	60
% Studies that need investigation	0

Based on 43 studies and a total of 50k incurred samples that were analyzed at BioPharma Services Inc.

# A typical batch from mesalamine study



Mean: 9.2 E5; Min: 8.9 E5; Max: 9.6 E5

Within ±4% variation in absolute response!

## How did we accomplish that?

### Many factors can cause IS variations!

IS solution	Stability and/or solubility issue	
IS addition	Missed or double addition; imprecise addition	
Extraction	Variation in recovery for IS & co-extracted matrix components	
Injection	Missed or variable injection vol.	
Chromatographic separation	Co-elution with ion-suppression or enhancement component(s); late eluter; column deterioration	
MS detection	Ion suppression/enhancement; ionization/detector saturation; inadequate optimization	
Others	Wrong materials; leakage; inadequate equipment/instrument use, etc.	

1) 2) A. Tan et al., J. Chromatogr. B, 877 (2009) 3201.

A. Tan et al. (2012), Internal Standards for Quantitative LC-MS Bioanalysis, in LC-MS in Drug Bioanalysis (eds. Q.A. Xu and T.L. Madden), Springer, New York, USA, p. 1.

# No approach is perfect for monitoring IS variation.

Setting the upper and lower limits for IS response

e.g. 50-150% of the mean IS response of known samples (CS & QC)

 Performing a trend analysis on IS responses of known samples to define the acceptable variation for unknown samples

- 1) A. Tan et al., J. Chromatogr. B, 877 (2009) 3201.
- 2) M. Jemal et al., Rapid Commun. Mass Spectrom., 17 (2003) 1723.
- 3) R. Bakhtiar, T.K. Majumdar, J. Pharmacol. Toxicol. Methods, 55 (2007), 227.
- 4) Global CRO Council (GCC), Bioanalysis, 3 (2011) 1323.

### But, there is a bottom line...

#### Summary of IS repeats in our lab

Due to low IS response (%)	70
Due to high IS response (%)	30
Unmatched original and reassay results (due to error in IS addition) (%)	30 (<100)
Overall IS addition error rate (%)	0.06

Based on a total of 50k incurred samples that were analyzed at BioPharma Services Inc.

- 1) Unmatchable original results must be singled out for reassay;
- 2) Any IS variation patterns that were not seen in R&D/assay validation should be detected and investigated upon.

# Even reproducible results may not be reportable!

# Response vs. concentration relationship might have changed.

linear for CS  $\rightarrow$  quadratic for samples of abnormal IS responses or vice versa

#### Abnormal IS responses may be outside IS linearity range.

*Ionization or detector saturation;* 

Analyte & IS may not be simultaneously detected inside MS.

- 1) G. Liu, Q.C. Ji, M.E. Arnold, Anal. Chem., 82 (2010) 9671.
- 2) A.K. Hewavitharana, J. Chromatogr. A, 1218 (2011) 359.
- 3) A. Tan and K. Awaiye (2013), Use of Internal Standards in LC-MS Bioanalysis, in Handbook of LC-MS Bioanalysis: Best Practices, Experimental Protocols, and Regulations (eds. W. Li, J. Zhang and F. L.S. Tse), John Wiley & Sons Inc., Hoboken, NJ, USA, p. 217.

#### Strategy 1: Choose a good IS and use it properly

#### Select the best IS possible;

<sup>13</sup>C and/or <sup>15</sup>N > deuterated > structural analogue;
 A stable isotope labeled IS for parent drug is a structural analogue IS for the metabolite!

- One IS for each analyte;
- Determine an appropriate IS concentration;
- Co-elution of analyte and its IS;
- Check working IS solubility & stability;
- Check IS linearity;
- Accurate and reproducible addition of IS.

A. Tan and K. Awaiye (2013), Use of Internal Standards in LC-MS Bioanalysis, in Handbook of LC-MS Bioanalysis: Best Practices, Experimental Protocols, and Regulations (eds. W. Li, J. Zhang and F. L.S. Tse), John Wiley & Sons Inc., Hoboken, NJ, USA, p.217.

#### Strategy 2: Achieve high & consistent recovery

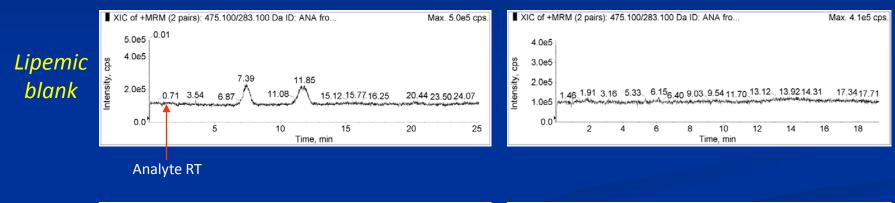
Determine the best extraction strategy;

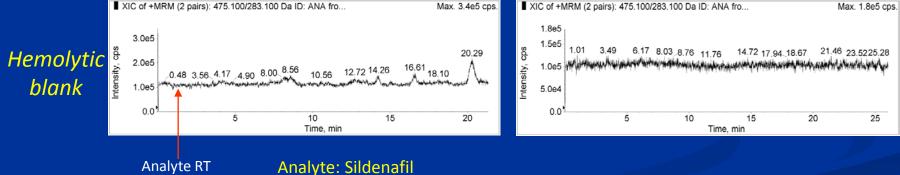
- Make sure there is sufficient buffering capacity!
- Adequate transfer volumes;
- Adequate reagent volumes.

#### Strategy 3: Don't stop at successful matrix factor test!

#### Mobile Phase A

#### Mobile Phase B





Matrix: Human EDTA plasma Extraction: SLE

## **Other Strategies**

- Test different LC columns and different LC–MS systems;
- Add extra tests as needed;

e.g. check autosampler stability for hemolyzed samples if the analyte is a phenolic compound\*

- Precise execution of validated assay;
- Close monitoring of assay performance;
- Adequate maintenance of lab equipment and instruments;

### Hire the best research scientists!

<sup>\*</sup> E.-R. Bérubé, M.-C. Lacasse, M. Furtado, F. Garofolo, Bioanalysis, 5 (2013) 1491.

#### Quiz: What's the cause of IS variation? What could be improved in this method?

Analyte:	Weak base	Aliquot 300 µl plasma sample
IS: Matrix:	D <sub>7</sub> Human K <sub>2</sub> EDTA plasma	+ Add 700 μl 50 mM Na <sub>2</sub> HPO <sub>4</sub>
Conc. range:	15-4000 pg/ml (linear, 1/C <sup>2</sup> )	Add 100 µl IS in 50% MeOH
Extraction:	SPE, Bond Elut-C18 100 mg/1 ml	↓ Mix by vortexing ↓
Activation	: 1 ml MeOH & 1 ml H <sub>2</sub> O>	Load on activated cartridge
Scenario:		Wash with 1 ml H <sub>2</sub> O
1) Successfu recovery	l assay validation (analyte 66-71%);	Wash with 1 ml 50% MeOH
<ol> <li>Low &amp; variable IS responses mainly with study samples;</li> </ol>		Elute with 0.5 ml ACN
<ol> <li>20% of IS repeats still had low IS responses.</li> </ol>		Evaporate at 40°C
4) Good agreement between reassay and initial results, including ISR.		↓ Reconstitute in 200 μl MP

## Conclusions

- Many different factors can cause IS response variation.
- Once observed, investigation should be done to find the root cause. It is important to demonstrate that the accuracy of quantitation has not been impacted.
- Sometimes, reproducible results may still not be reportable.
- IS response variation is preventable or at least it can be significantly reduced through thoughtful assay development and precise execution of validated assays.

#### Acknowledgements

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# Thank you for your attention! Questions or comments?

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