



LC-MS/MS bioanalysis of peptides – How to manage non specific binding?

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Outline of this presentation



MEDICINE ON THE BODY'S OWN TERMS

- Background
 - Challenges in LC-MS/MS bioanalysis of peptides
 - Non-specific binding (NSB)
 - Consideration for peptides
- Strategies for minimising NSB in LC-MS/MS bioanalysis of peptides
- Two case studies where NSB have hampered bioanalytical method development
- Lessons learned
- Acknowledgement

Challenges in LC-MS/MS bioanalysis of peptides

- Formation of multiple-charged ions
- Extensive fragmentation in MS/MS
- Adsorption problems → **NSB issue**
- Chromatographic issues → **potential NSB issue**

Non-specific binding (NSB)

- **Definition:** Non-specific binding – analyte losses that occur as a result of binding to any surface to which the analyte is exposed
- **Peptides:** Often demonstrate greater NSB issues than small molecules
- **Cause:** Physical/chemical properties e.g. Van der Waals interactions, ionic interactions
- **Problems:** Sensitivity and robustness. Most troublesome at low concentrations.

Non-specific binding

- NSB can occur to various material throughout the analytical procedure, e.g.
 - Vials
 - Plates

} In pure solution and in evaporation/reconstitution

 - Tubing
 - Columns
 - Injection valves

} In chromatography

Considerations for peptides

- Charged peptides
 - greater solubility in aqueous solution
 - less adsorption to plastics
- Uncharged peptides
 - can adsorb strongly to plastics
 - organic solvent may enhance solubility and prevent adsorption
- Need to be investigated during method development, generic approach will not work

1. Use high concentration stock, spike into matrix and perform serial dilution in the matrix
2. Choice of material in tubes etc.
 - Evaluate glass or polypropylene vessel i.e. high quality/low binding polypropylene material or glass or glass coated material
3. Use additives
 - Organic solvent
 - Protein (e.g. HSA or BSA)
 - Surrogate peptide
 - Detergent (e.g. Triton-X, Tween or C₉-Glu)

20 AA charged peptide, Mw ~ 2400 Da

Start of non-clinical program

Evaluation of sample container material: glass or polypropylene (PP) 96-well plates

- Better precision with PP compared to glass

Dissolve peptide in ACN:5 mM Ammonium acetate (10:90) with and without additives (0.1% HSA + 0.01% Triton-X)

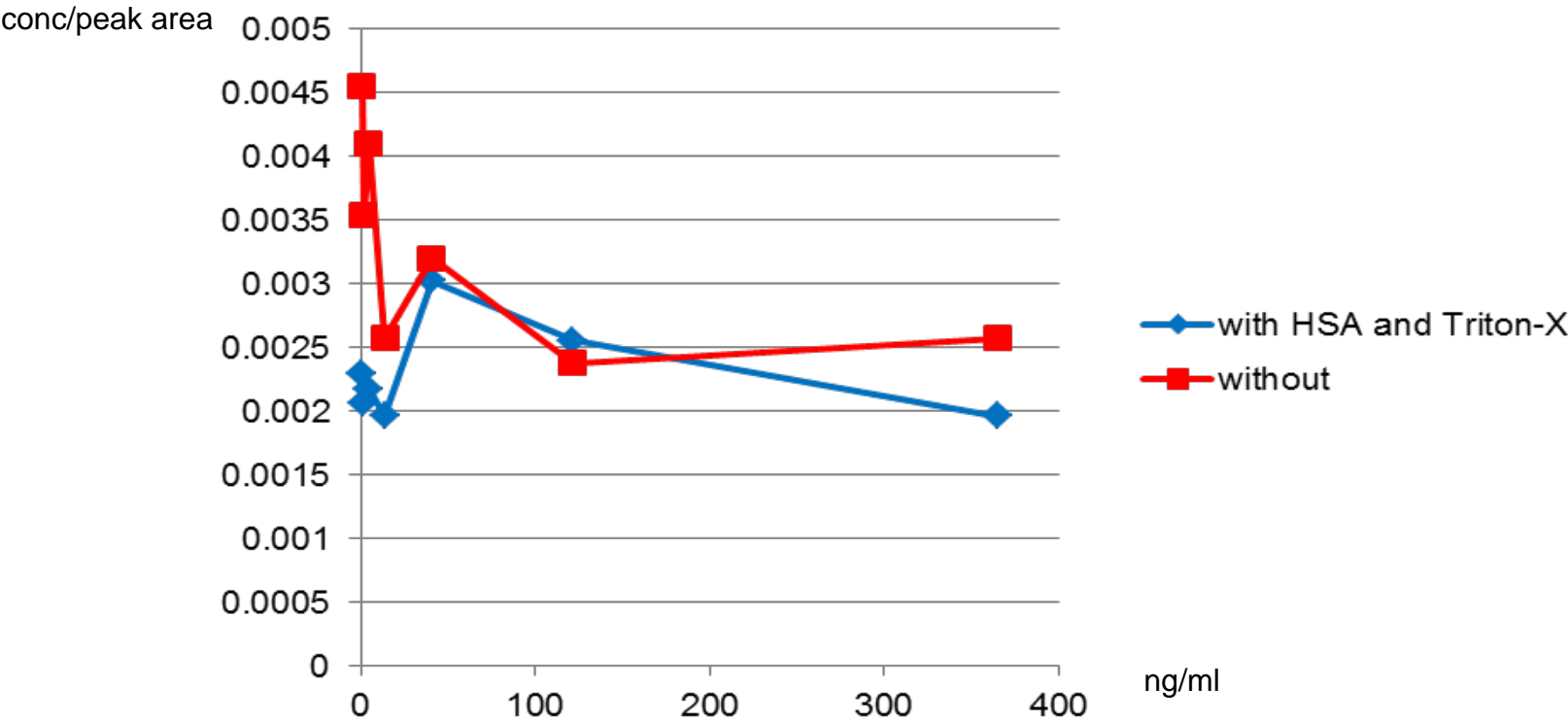
- With additives gave ~ 25% higher response in peak area

Case 1- Sample plate material and use of additives



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- Less absorption with additives (0.1% HSA + 0.01% Triton-X) as judged by looking at response factors



- Better precision and accuracy with additives (0.1% HSA + 0.01% Triton-X)

Case 1- Method for rat plasma validated



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Method for determination of peptide in rat plasma successfully validated

Occasion	Nominal concentration (ng/mL)	0.50	1.50	7.91	106
<i>Validation run 1</i>	Accuracy (% bias)	18	15	6	6
	Precision (%CV)	5	4	2	1
<i>Validation run 2</i>	Accuracy (% bias)	-6	0	-5	-3
	Precision (%CV)	20	6	6	3
<i>Validation run 3</i>	Accuracy (% bias)	17	16	8	7
	Precision (%CV)	6	7	4	1

Study samples analysed successfully.

Case 1 – start of validation for dog plasma



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The same method applied for dog plasma, same sample volume 50 µl

Occasion	Nominal concentration (ng/mL)	5.00	15.00	79.10	1060
QC test run	Accuracy (% bias)	-4	-5	3	-8
	Precision (%CV)	8	8	11	5
Validation run 1	Accuracy (% bias)	-8	-12	-13	-24
	Precision (%CV)	3	7	9	6
Validation run 2	Accuracy (% bias)	4	-14	-17	-20
	Precision (%CV)	19	10	8	9
Validation run 3	Accuracy (% bias)	-23	-10	-11	-16
	Precision (%CV)	4	11	5	4

→ Validation stopped – back to method development

Case 1 – start of validation for dog plasma



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According to EMA and FDA Guidelines on bioanalytical method validation,

The calibration curve should preferably be prepared using freshly spiked samples when appropriate stability data is not available.

→ can cause problems with bias especially for peptides if not handled carefully.

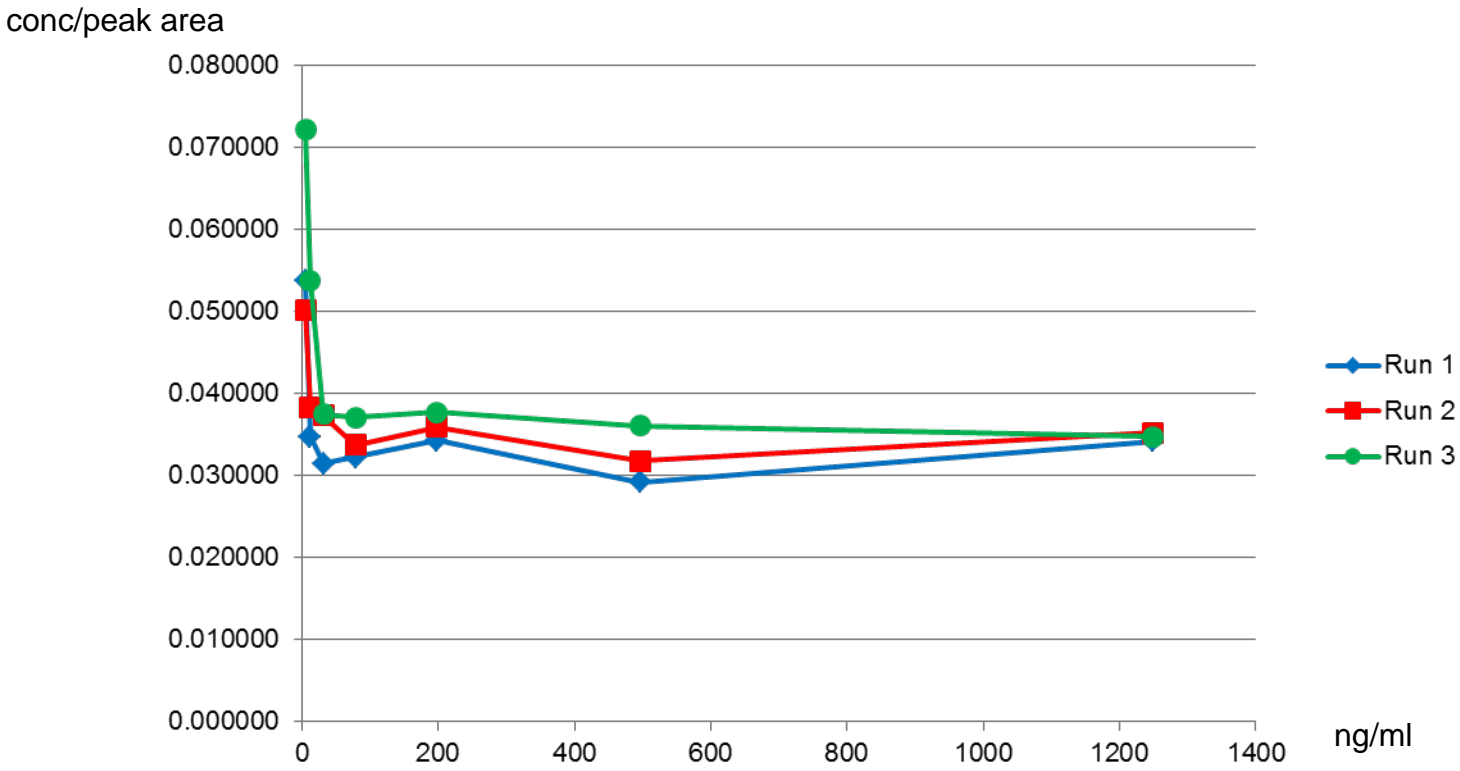
Case 1 - development for method in dog plasma



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The CAL and QC samples were prepared by spiking from dilutions in ACN:5 mM Ammonium acetate (10:90) with additives (0.1% HSA + 0.01% Triton-X)

Response factors for CAL samples



Higher response factor at low concentrations indicted issue with NSB, affected slope on calibration curve with $1/x^2$ weighting factor.

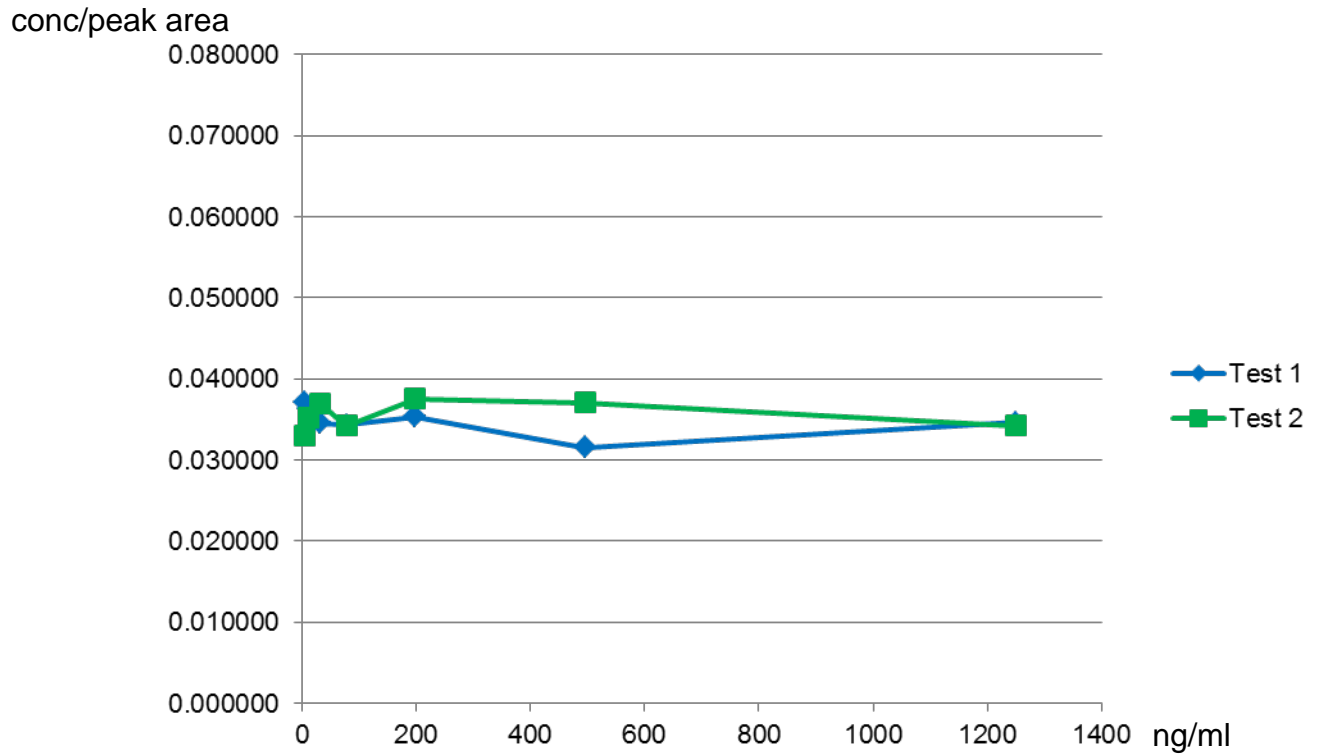
Case 1 - development for method in dog plasma



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Preparation of CALs and QC:s from high concentration stock (5 times higher) with increased HSA concentration (from 0.1 % to 0.5 % HSA) and dilution in sample matrix (dog plasma).

Response factors for CAL samples



Case 1 - development for method in dog plasma



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Validation restarted

Occasion	Nominal concentration (ng/mL)	5.00	15.00	79.10	1060
<i>Validation run 1</i>	Accuracy (% bias)	8	3	-2	3
	Precision (%CV)	5	3	3	2
<i>Validation run 2</i>	Accuracy (% bias)	-7	-2	3	8
	Precision (%CV)	10	8	5	4
<i>Validation run 3</i>	Accuracy (% bias)	8	9	4	13
	Precision (%CV)	4	9	4	4

Validation successful for dog plasma

Case 2



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- 9 AA peptide, MW ~ 1000 Da
- Method successfully validated in plasma for three animal species + human (LLOQ - 5 pg/mL)
- However, problem occurred for fourth animal species + human LTS occasion

Case 2

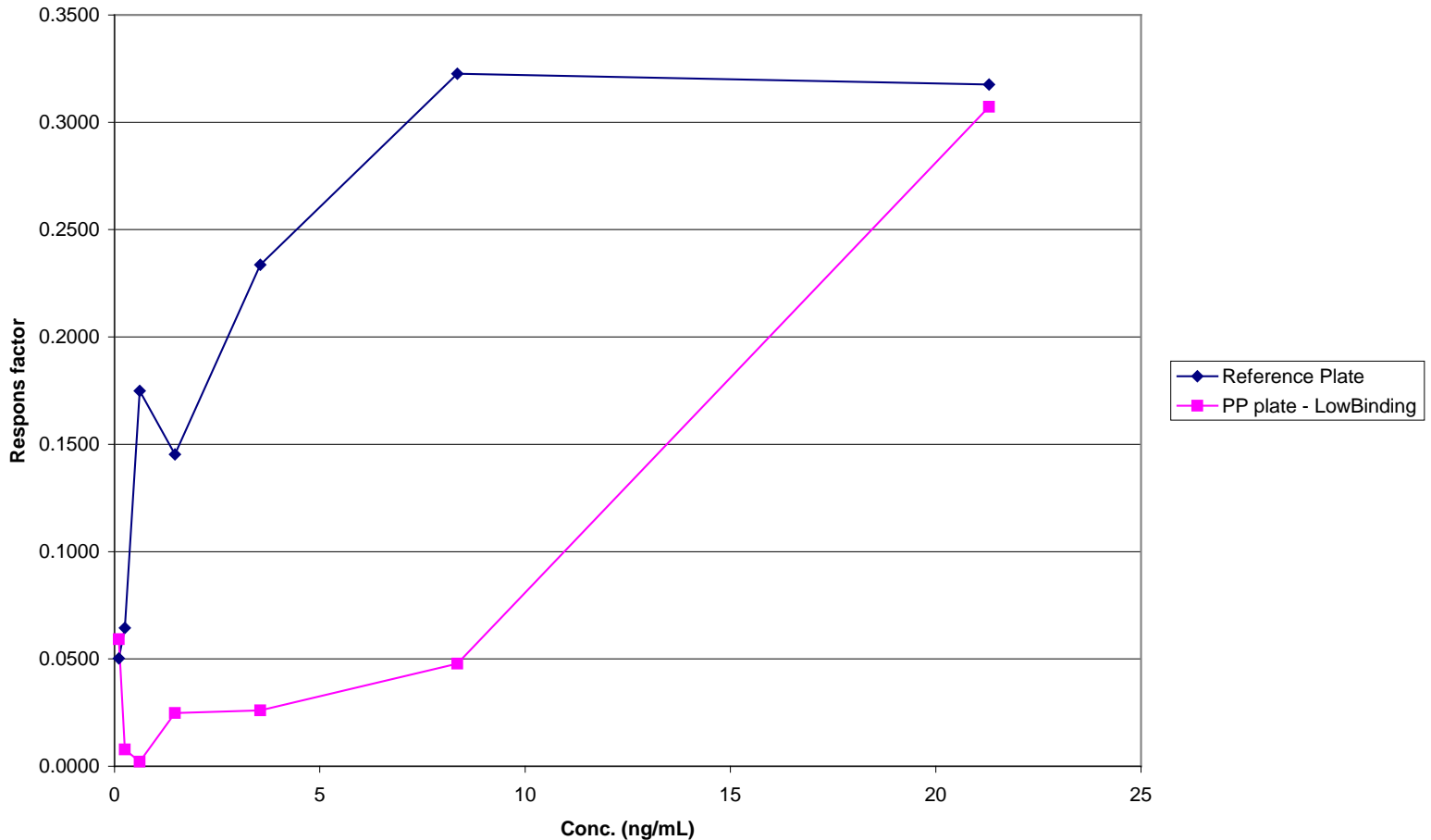
- Sheep plasma, 0.1 – 25 ng/mL
- 0.1% HSA and 0.01% Triton-X as additive in stock solution

Sample	Response per ng/mL
CAL 1	0.016
CAL 2	0.018
CAL 3	0.031
CAL 4	0.039
CAL 5	0.101
CAL 6	0.092
CAL 7	0.097

- Found to be related to PP plates used for preparation of calibration samples, from same manufacture as used for previous studies!

Case 2

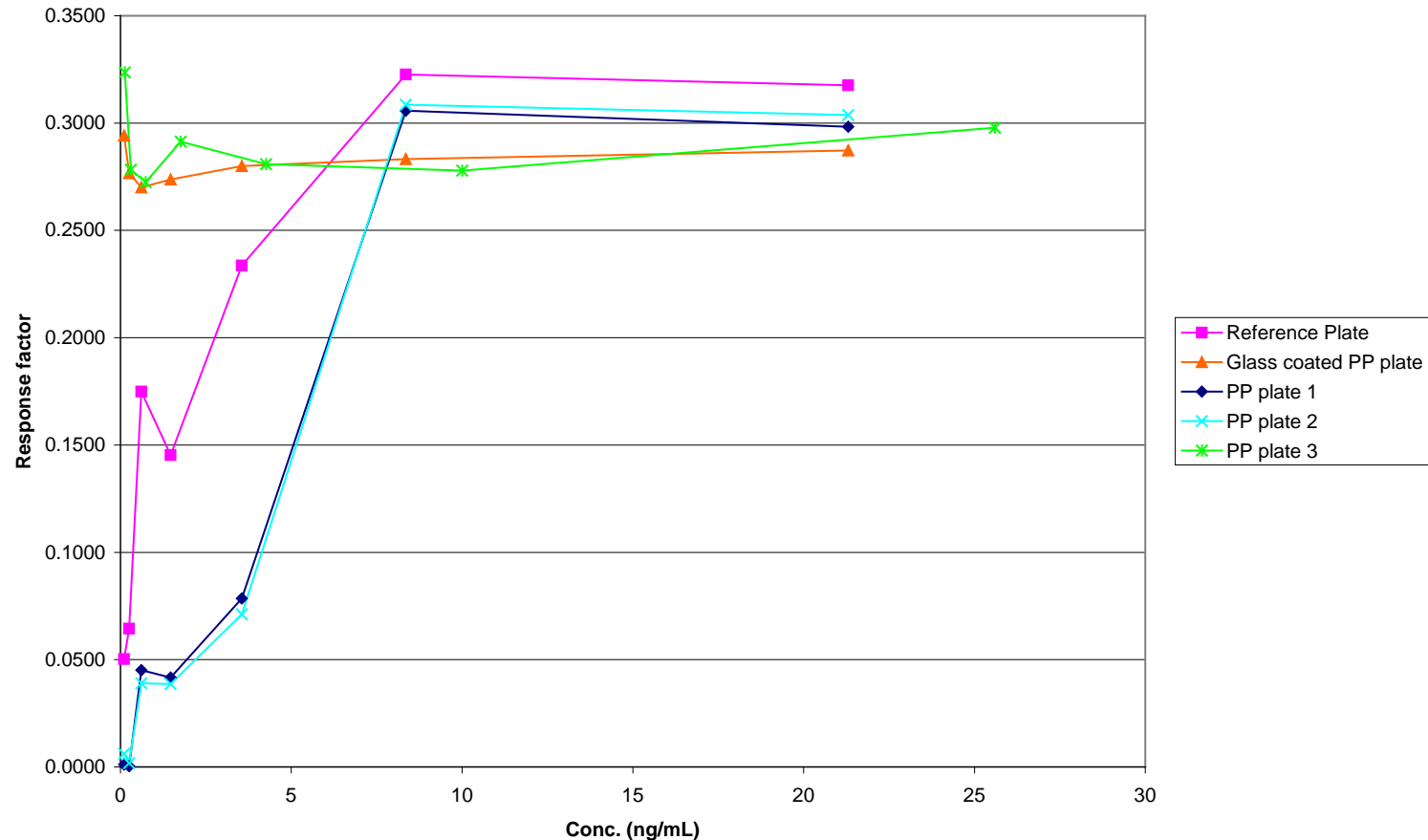
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Severe NSB also found when using specific low binding PP plate, worse than in our reference PP plate!

Case 2

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- Selecting another supplier of PP plate or using a glass coated PP plate solved the NSB issue.
- Method for sheep plasma was successfully validated with PP plate 3.

- Careful selection of material and additive might minimize problem
 - Test during method development
- Even for well established methods, NSB to material can all of sudden occur
 - As part of trouble shooting, good practice to start looking at the material involved (e.g. by looking at the response factors)
- Calibration standards and quality control samples of peptides should be prepared by spiking from a high concentration stock solution directly into plasma and subsequently serial-diluted in the same sample matrix.

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