

# **DEVELOPMENT OF AFFINITY MICROCOLUMNS FOR HIGH-THROUGHPUT BIOINTERACTION ANALYSIS**

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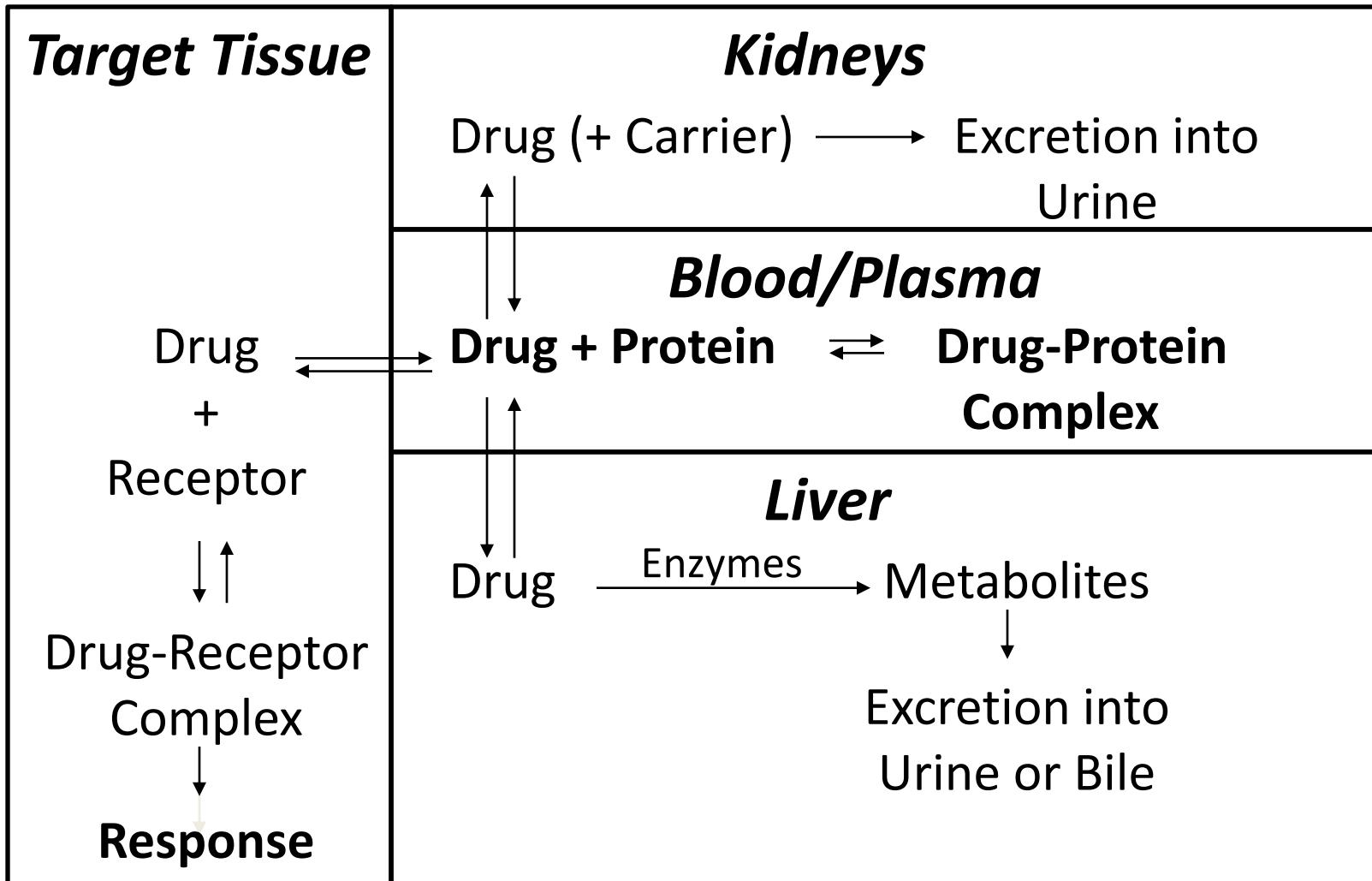
Department of Chemistry



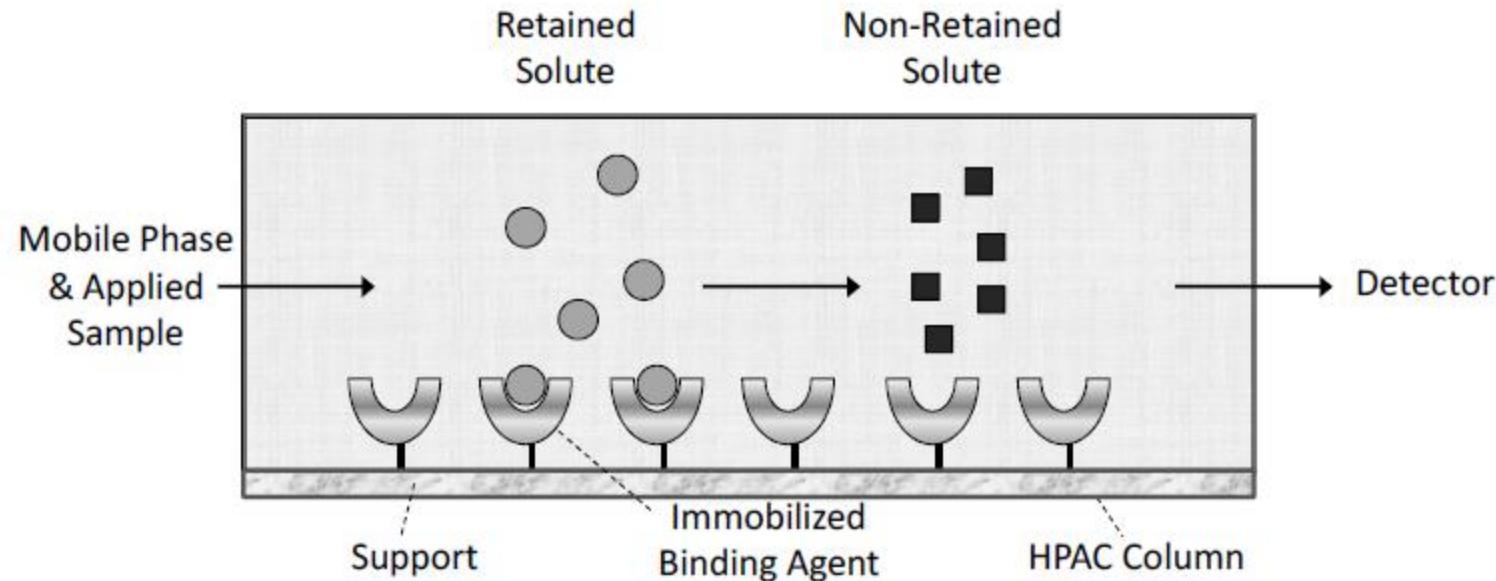
European Bioanalysis Forum 2011  
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# Drug-Protein Interactions

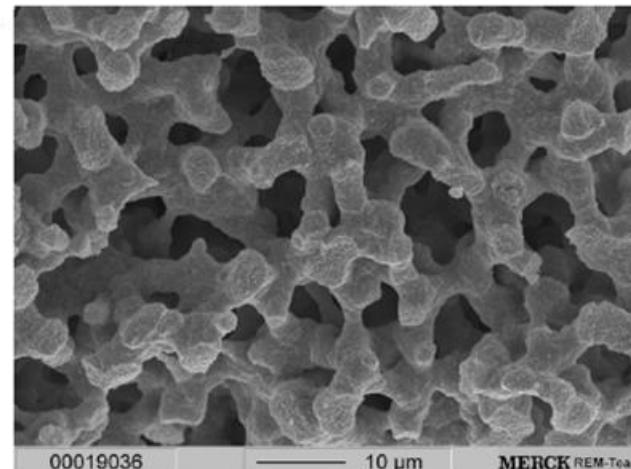
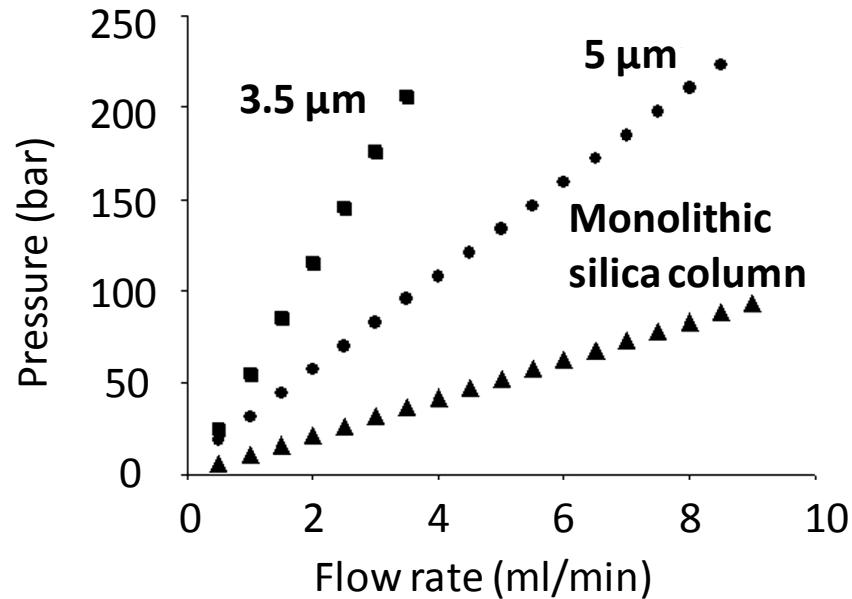


# High-Performance Affinity Chromatography (HPAC)



# Monolithic Supports

- Higher external porosity than particle-based supports
  - Higher permeability
  - Lower backpressures
  - Better efficiency
- Two types of pores
  - Flow-through pores (macropores)
  - Smaller “diffusion” pores (mesopores)



Cabrera, K. *J. Sep. Sci.* 2004, 27, 843-852.

Mallik, R.; Hage D.S. *J. Sep. Sci.* 2006, 29, 1686-1704.

# Silica Particles vs. Silica Monoliths

## Silica Particles

- Good mechanical strength
- High surface area
- Total porosity ~65%

## Silica Monoliths

- Single piece of porous silica
- Total porosity ~80%
- Lower backpressure
- Better mass transfer
- Faster separations
- Shrinkage



from [www.phenomenex.com](http://www.phenomenex.com)

Cabrera, K. *J. Sep. Sci.* **2004**, 27, 843-852.  
Mallik, R.; Hage D.S. *J. Sep. Sci.* **2006**, 29, 1686-1704.



# Affinity Microcolumns

## Advantages

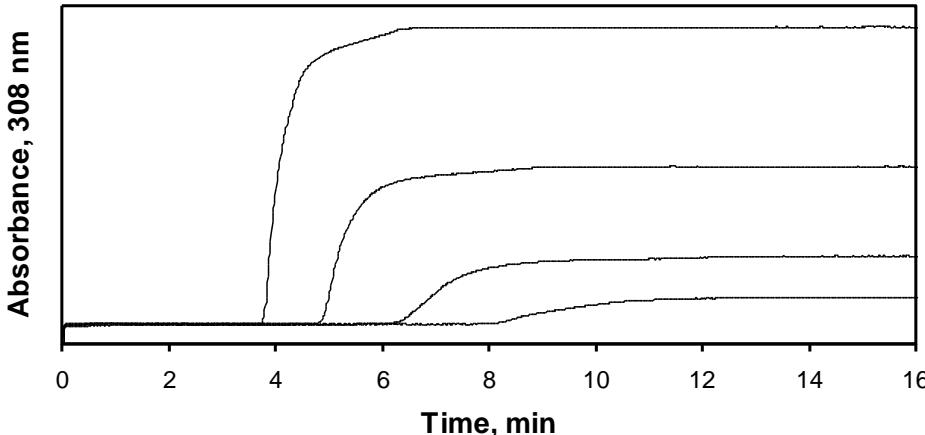
- Require small amounts of ligand
- Reduced surface area, less non-specific binding
- Reduced analysis times

## Challenges

- Need new approaches for preparing and packing columns
- Need new immobilization methods to increase the activity of ligand
- Need better understanding of properties and limitations

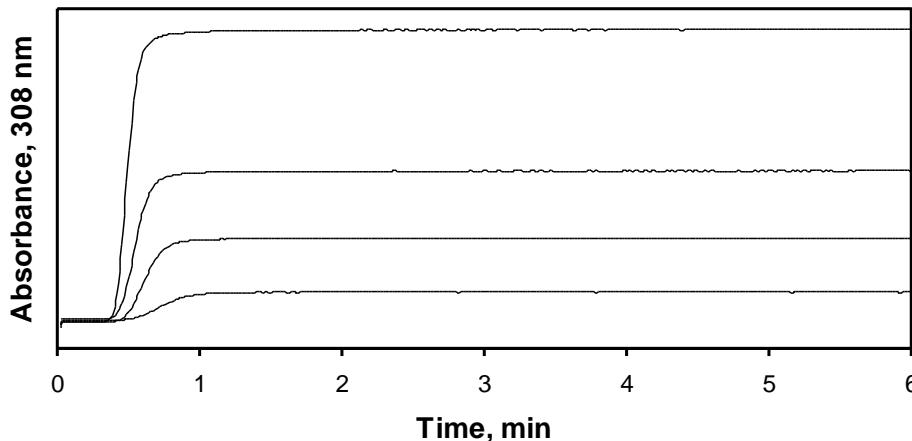
# Frontal Analysis Measurements

2 cm x 2.1 mm i.d.



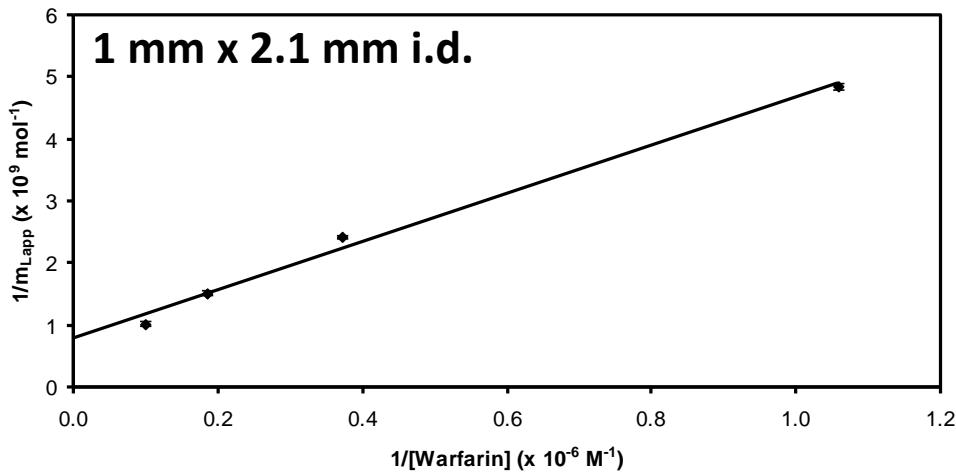
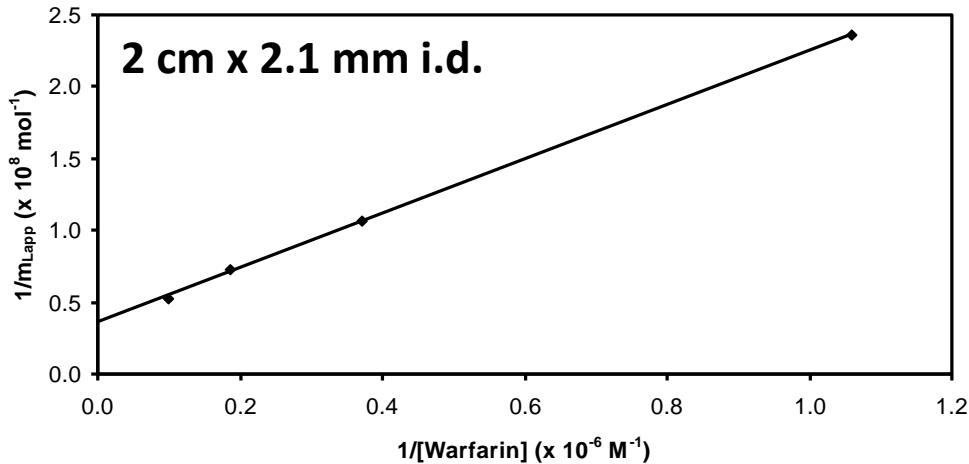
- Breakthrough times:  
4.2-9.5 min
- Breakthrough volume:  
2.8 mL

1 mm x 2.1 mm i.d.



- Breakthrough times:  
30-45 s
- Breakthrough volume:  
0.29 mL

# Frontal Analysis Measurements



$$\frac{1}{m_{\text{Lapp}}} = \frac{1}{K_A m_L [A]} + \frac{1}{m_L}$$

# Frontal Analysis Results

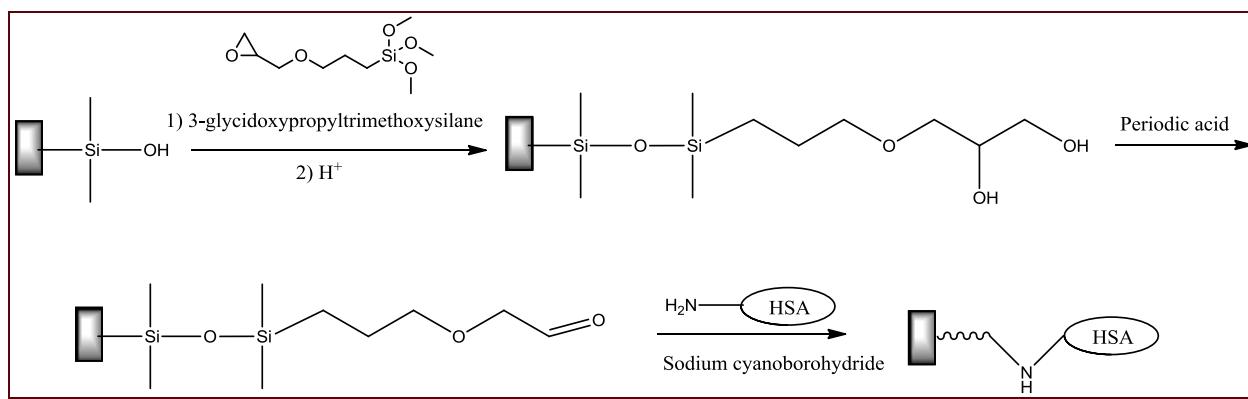
Column Length (mm)	$m_L$ (nmol)	Relative Activity (versus 20 mm column)	$K_a$ ( $\times 10^5 \text{ M}^{-1}$ )
20	27.6 ( $\pm 0.2$ )	1.00	2.0 ( $\pm 0.1$ )
3	3.8 ( $\pm 0.3$ )	0.9 ( $\pm 0.1$ )	2.3 ( $\pm 0.1$ )
2	2.6 ( $\pm 0.3$ )	0.9 ( $\pm 0.1$ )	2.1 ( $\pm 0.2$ )
1	1.2 ( $\pm 0.1$ )	0.8 ( $\pm 0.1$ )	2.6 ( $\pm 0.1$ )

Literature  $K_a$  values:  $2.1 - 2.6 \times 10^5 \text{ M}^{-1}$

- Binding capacity decreased in proportion to column size
- Decrease in precision in  $K_a$  values (2 cm:  $\pm 5\%$ , 1 mm:  $\pm 25\%$ ) due to decrease in amount of time and stationary phase available for drug binding

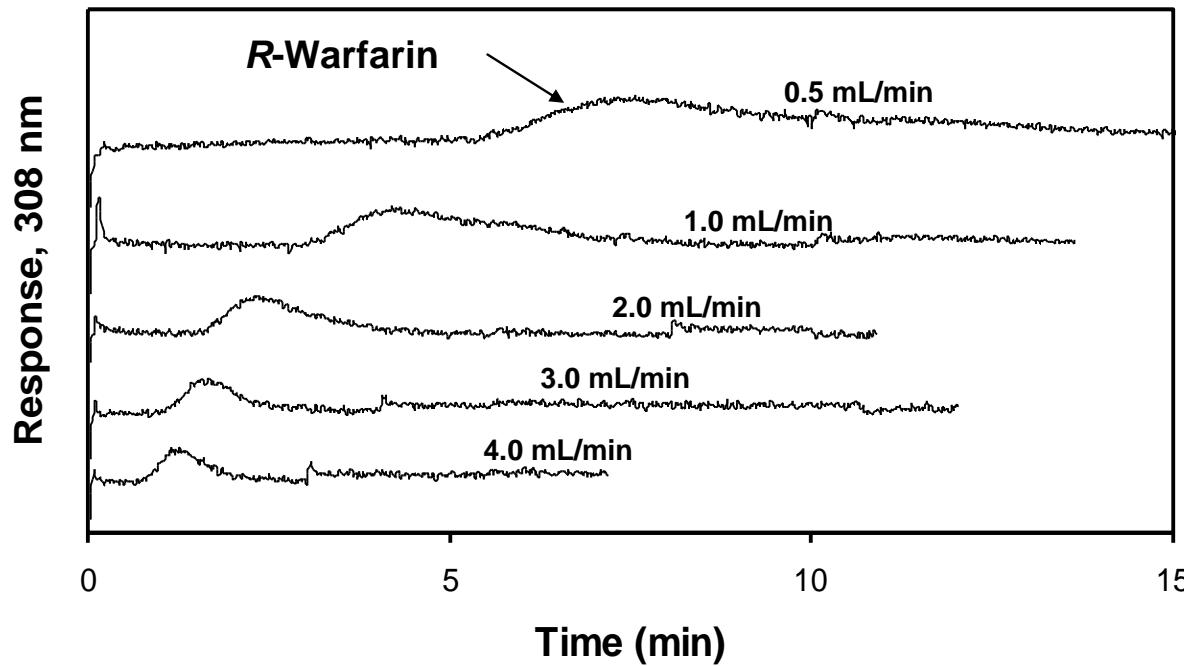
# Preparation of HSA Silica Monoliths

- Chromolith™ Performance Si column was donated by Merck KGaA (4.6 mm i.d. x 10 cm)
  1. Columns cut to lengths from 1 to 5 mm
  2. Converted into diol silica monolith
  3. Immobilized HSA by Schiff base method

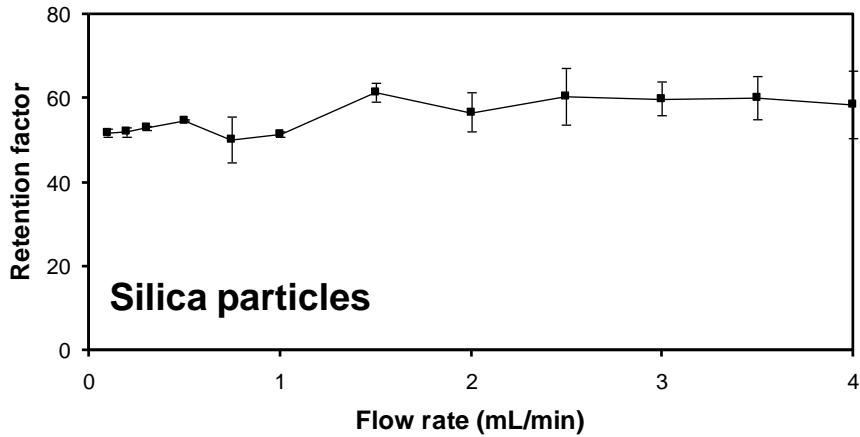
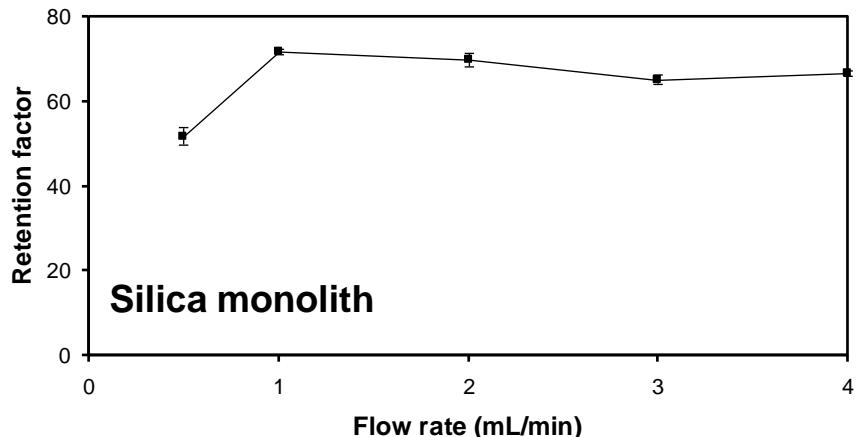
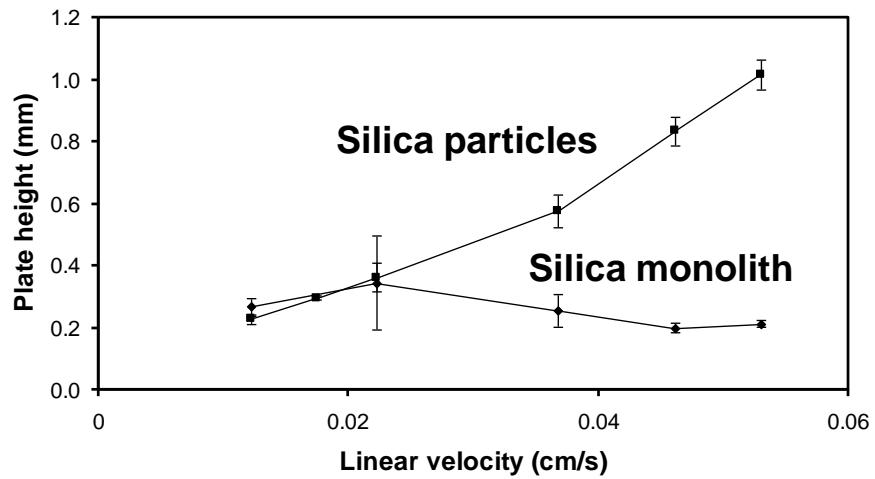


# Warfarin Retention on HSA Silica Monoliths

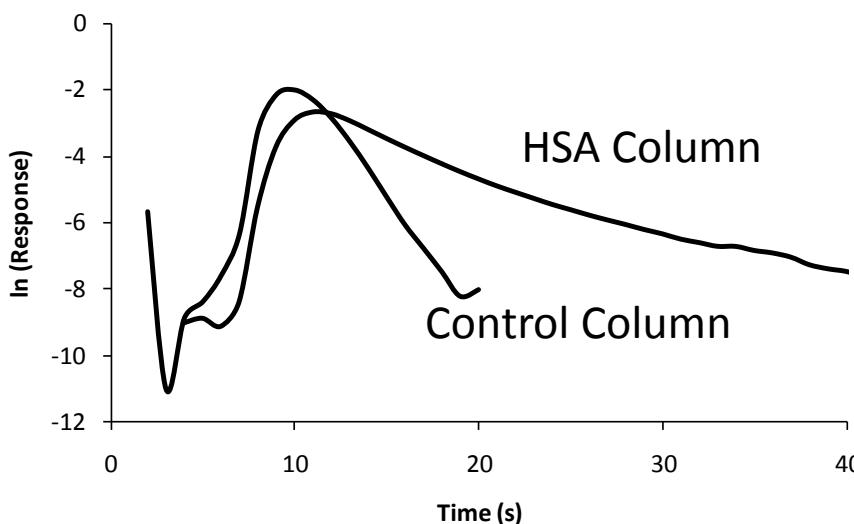
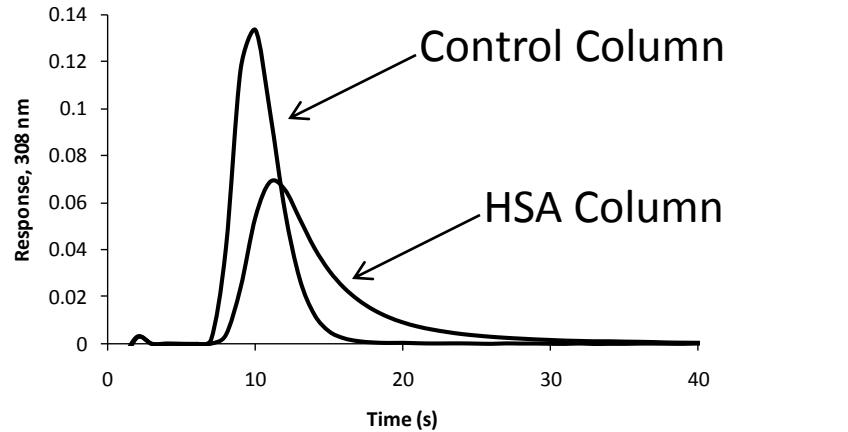
- Longer HSA columns (4-10 cm): 25-150 min
- HSA silica monolith microcolumn (3 mm): 1.4-9.5 min



# Silica Particles vs. Silica Monoliths



# Rapid Determination of Drug-Protein Dissociation Rates



## Control Column

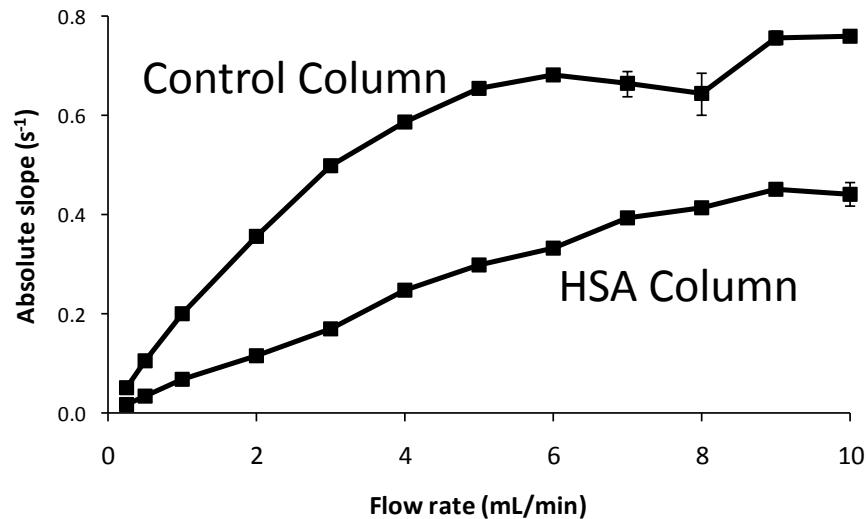
- Washing away of non-bound analyte
- Release of any non-specifically bound analyte

## HSA Column

- Release of the analyte from the immobilized HSA

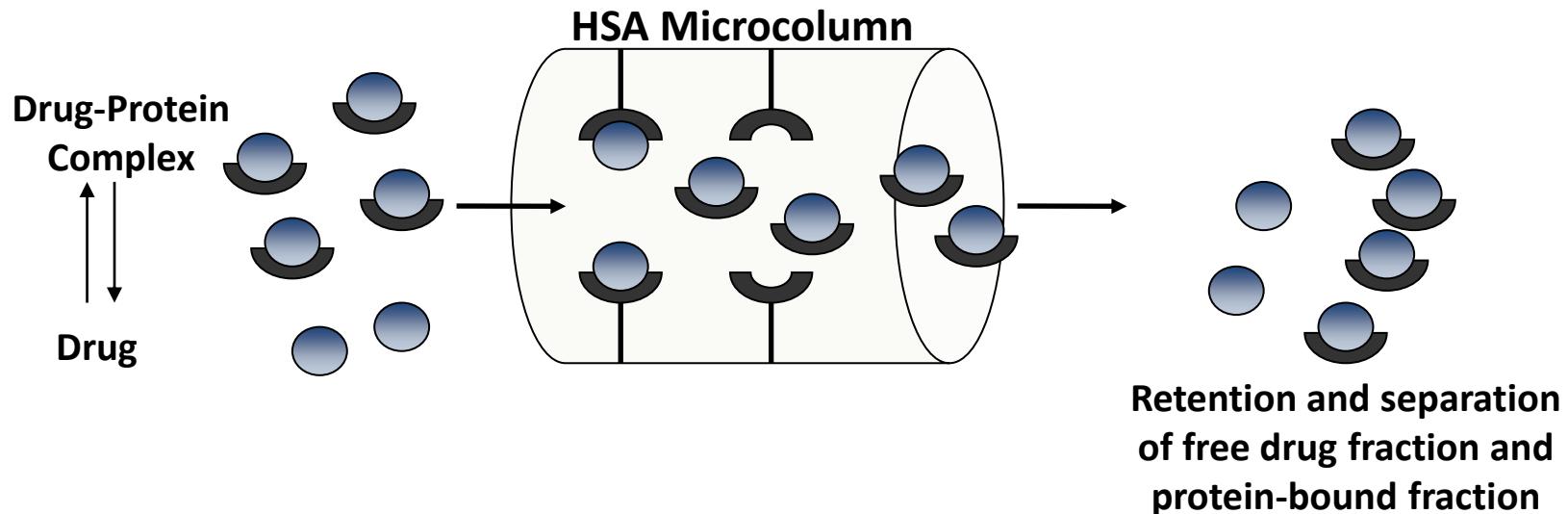
# Typical Results

Drug	$K_a$ (M <sup>-1</sup> )	Binding Site on HSA	Measured Value $k_d$ (s <sup>-1</sup> )	Literature Value $k_d$ (s <sup>-1</sup> )
Warfarin	$2.1\text{-}2.6 \times 10^5$	I	$0.41 (\pm 0.06)$	$0.35 - 0.66$
Imipramine	$1.6 \times 10^5$	II	$0.29 (\pm 0.11)$	$0.41 - 0.67$
Diazepam	$2.2 \times 10^5$	II	$0.44 (\pm 0.11)$	--
Acetohexamide	$0.43\text{-}1.3 \times 10^5$	I, II	$0.58 (\pm 0.02)$	--
Tolbutamide	$5.3\text{-}5.5 \times 10^4$	I, II	$0.49 (\pm 0.15)$	--



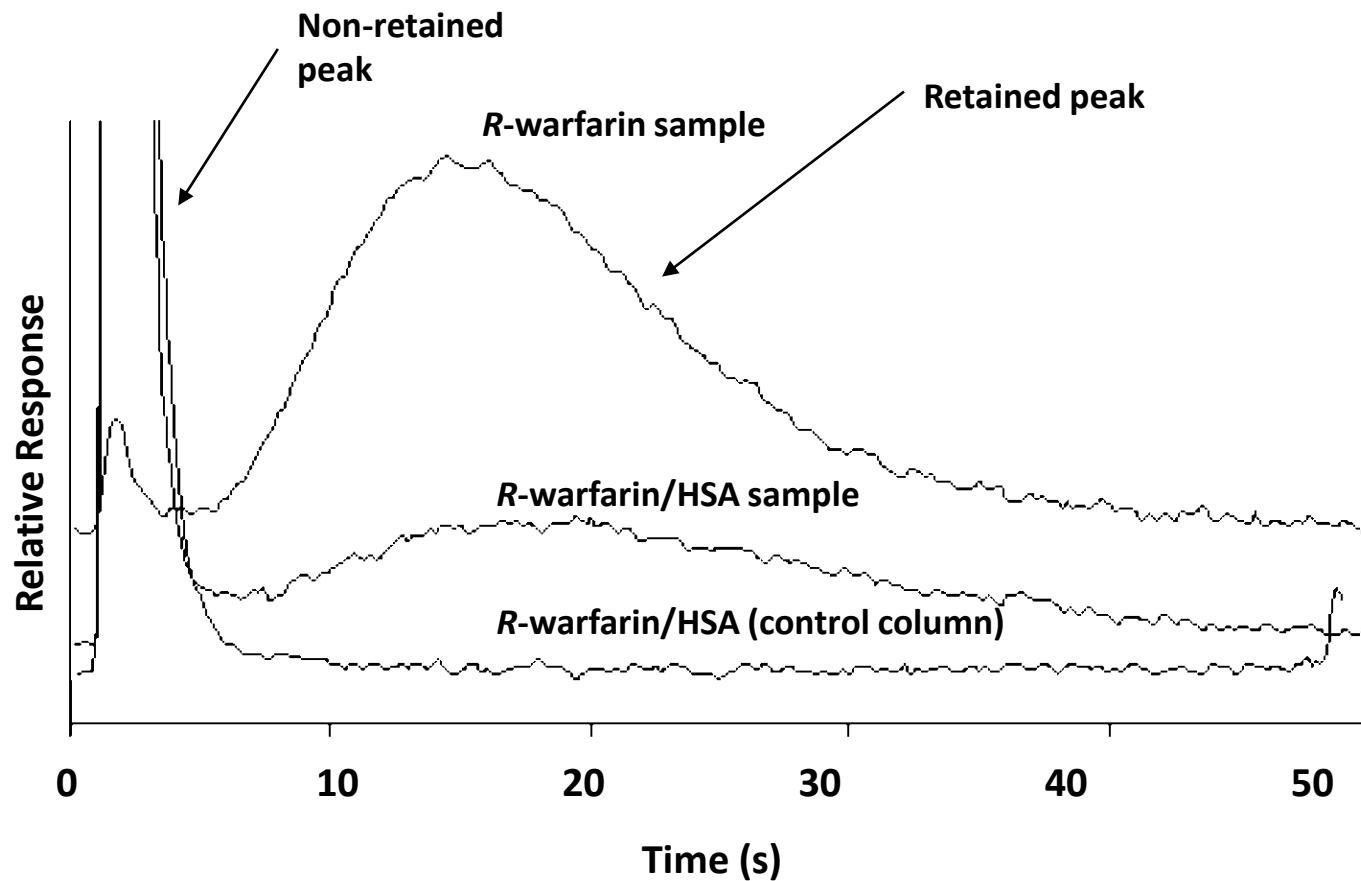
Same general behavior and types of plots were seen for the other drugs that were examined.

# Analysis of Free Drug Fractions



- HSA microcolumns were used in ultrafast extraction and free drug fraction measurements
  - HSA is most abundant serum protein and important binding agent for a variety of drugs-use for more than analyte
  - Lower association equilibrium constants of drugs for HSA ( $10^3$ - $10^5$ ) versus antibodies ( $10^6$ - $10^{12}$ ) allows for the use of isocratic conditions
  - Less expensive to perform compared to using columns based on immobilized antibodies

# Analysis of Free Drug Fractions



Analysis times of only 40 s per injection at 4.5 mL/min

Mallik, R.; Yoo, M.J.; Briscoe, C.J.; Hage, D.S. *J. Chromatogr. A*, 2010, 1217, 2796-2803.



# Analysis of Free Drug Fractions

Immobilization Method and Sample	HSA Microcolumns (Ultrafast Extraction)	Ultrafiltration (Reference Method)
<b>Schiff Base Method</b>		
S-Warfarin + HSA	0.20 ( $\pm$ 0.03)	0.20 ( $\pm$ 0.02)
R-Warfarin + HSA	0.31 ( $\pm$ 0.05)	0.28 ( $\pm$ 0.02)
<b>SMCC Method</b>		
S-Warfarin + HSA	0.23 ( $\pm$ 0.02)	0.21 ( $\pm$ 0.03)
S-Ibuprofen + HSA	0.28 ( $\pm$ 0.02)	0.25 ( $\pm$ 0.07)
Imipramine + HSA	0.92 ( $\pm$ 0.02)	0.86 ( $\pm$ 0.07)

# Summary

- Affinity microcolumns provide comparable results to those obtained with longer columns and can be used in rapid analysis of drug-protein binding
  - Smaller amount of protein (low nmol to upper pmol) needed for column preparation
  - Faster analysis times (net retention times are 100 times faster than traditional 10 cm column)
  - Affinity silica monolith microcolumns have better efficiency (higher flow rates, larger number of theoretical plates)
- Ultrafast extraction based on HSA microcolumns can be used for measuring the free fractions of some drugs in drug-protein mixtures.
  - Good agreement with reference method
  - Fast analysis times (40 s per injection)



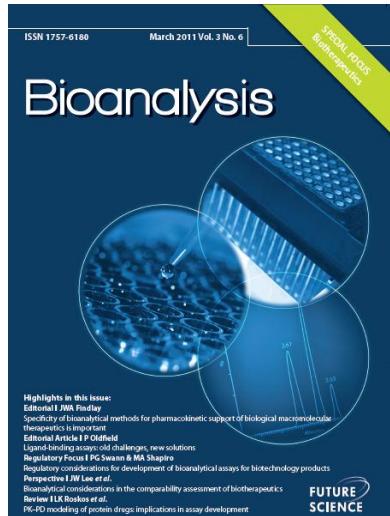
# References

- Hage, D.S. et al. *J. Sep. Sci.* **2009**, *32*, 835-853.
- Hage, D.S. et al. *Curr. Drug Metab.* **2011**, *12*, 313-328.
- Cabrera, K. *J. Sep. Sci.* **2004**, *27*, 843-852.
- Mallik, R.; Hage D.S. *J. Sep. Sci.* **2006**, *29*, 1686.
- Yoo, M.J. et al. *J. Chromatogr. B*, **2010**, *878*, 1707-1713.
- Yoo, M.J.; Hage, D.S. *J. Sep. Sci.*, **2009**, *32*, 2776-2785.
- Yoo, M.J.; Hage, D.S. *J. Chromatogr. A*, **2011**, *1218*, 2072-2078.
- Mallik, R.; Yoo, M.J.; Briscoe, C.J.; Hage, D.S. *J. Chromatogr. A*, **2010**, *1217*, 2796-2803.



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